

# The Integration of Osteopathic Approaches on Visceral Health

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International Diploma in Animal Osteopathy

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January 5<sup>th</sup>, 2025

## **Introduction**

This thesis emphasizes the connection between somatic dysfunction and visceral disease in osteopathic care. Understanding viscerosomatic reflexes is vital, as it demonstrates how issues in internal organs can present as physical symptoms, aiding practitioners in uncovering underlying health problems. Detailed assessments encompassing patient history and environmental factors are essential for revealing these relationships. Incorporating visceral techniques into Osteopathic Manual Therapy (OMT) has been shown to enhance treatment effectiveness and improve overall patient outcomes. By embracing a holistic approach that views the body as an interconnected system, osteopathic practitioners can more effectively address the complexities of patient's health concerns and promote patient-centered care. Research on animal anatomy has been extensive; however, studies on viscerosomatic reflexes, somatic dysfunction, and osteopathic evaluations and treatment approaches primarily focus on humans. Given that the studies may present similar indications, this thesis utilizes findings from both animal and human research to develop its conclusions.

### **1.0 Understanding Visceral Function & Anatomy**

"Viscera" encompasses the soft internal organs within the thorax, abdomen, and pelvic cavities, excluding those of the central nervous system, head and neck, skin, or musculoskeletal components. The viscera's functions include respiratory, circulatory, digestive, and reproductive processes, are regulated involuntarily, and crucial for maintaining homeostasis and health. To understand visceral dysfunction from an osteopathic perspective, it is necessary to examine the anatomical structures of the viscera. The organ system is a vast topic and extends beyond the overall scope of this thesis. For simplicity, the basic anatomy is outlined below.

### 1.0.1 Development of Viscera

An organ is a collection of two or more tissues that work together to form larger functional units. Organ development begins early in embryonic growth as cells organize germ layers. These germ layers contain active cell populations for forming essential vertebrate organ systems (MacCord, 2013). The layers are further categorized into somatic layers, which create body walls, and splanchnic layers, which develop into internal organs (O'Neill, 2024).

### 1.0.2 Tissues of the Viscera

An organ comprises various types of tissues organized into distinct layers. Most organs primarily consist of a main tissue corresponding to their primary function and additional supportive tissues. Although each organ has a specific type of tissue linked to its primary function, all organs are surrounded by muscles, connective tissue, and nervous tissues that contribute to their overall function. Smooth muscle fibers are present in the walls of hollow visceral organs, except for the heart, which comprises cardiac muscle. These smooth muscle fibers are one of the viscera's most essential components when discussing organ tissue. Smooth muscle differs from skeletal muscle in several fundamental ways, the most notable being its capacity to contract involuntarily. This ability is crucial for maintaining homeostasis within the organs. Moreover, smooth muscle exhibits greater elastic properties than skeletal muscle, allowing the organs to tense and relax while preserving their contractile tone continuously (Hafen, 2023). Without smooth muscle, the body could not perform its most fundamental functions.

Fascia is another essential part of the connective tissue system, consisting of interwoven membranous tubular layers of collagen and elastic fibers rich in fluid. It encapsulates every muscle, bone, gland, and cell within the organism, including the tissues that enclose the nervous system, vascular structures, and all visceral organs (Horton, 2015). Functionally, the fascia operates as a mechanical support system and a communication network within the body (Langevin, 2006). Additionally, it attaches to and stabilizes visceral and somatic structures, providing robust support and strength (Gatt et al., 2023). It also functions as a neural, lymphatic, and vascular systems channel (Horton, 2015). Furthermore, fascia facilitates the separation of body cavities and muscles, envelopes various organs, and establishes sliding surfaces that promote movement while minimizing friction (Horton, 2015). In addition to fascia, muscle and nervous tissue, all organs in the body receive blood and have lymphatic components from the vascular system, ensuring that tissues remain healthy and alive.

### 1.0.3 Structure of Visceral Spaces

The viscera are housed and organized within specific spaces in the body known as cavities. These cavities are filled with fluid, contain various internal organs and are categorized based on the cavity they occupy, such as the pleura (thoracic cavity), pericardium (surrounding the heart and blood vessels), peritoneum (abdominopelvic cavity) and the pelvic cavity.

The structure of visceral spaces is characterized by an outer layer of fibrous fascia that encases each organ, referred to as parietal fascia (Gatt et al., 2023). This outer layer is complemented by a serous membrane composed of specialized cells. The serous layer adheres to the parietal fascia, creating a structure that acts as a "wall" for the visceral space and extends

inward to form the "skin" of the organ, known as visceral fascia (Gatt et al., 2023; Hedley, 2023). Nestled between the organ's "skin" and its "wall" is a delicate layer of serous fluid produced by the serous membrane (Hedley, 2023). This fluid facilitates differential movement, fosters a connection among organs, minimizes friction, facilitates gliding motion, and permits high mobility within the space (Horton, 2015). This structural arrangement is consistent across various visceral spaces within the body's cavities (Figure 1 and Figure 2).

#### Gil Hedley: The Structure of Visceral Spaces

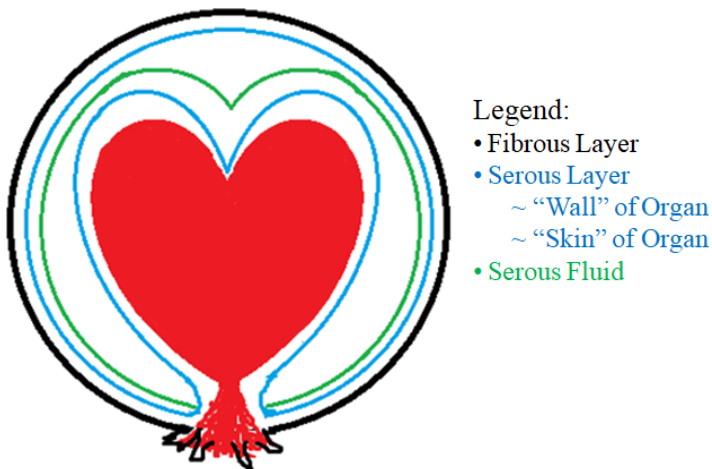


Figure 1. Recreation of Gil Hedley's drawing of visceral spaces (Hedley, 2023).

#### Gil Hedley: The Structure of Visceral Spaces

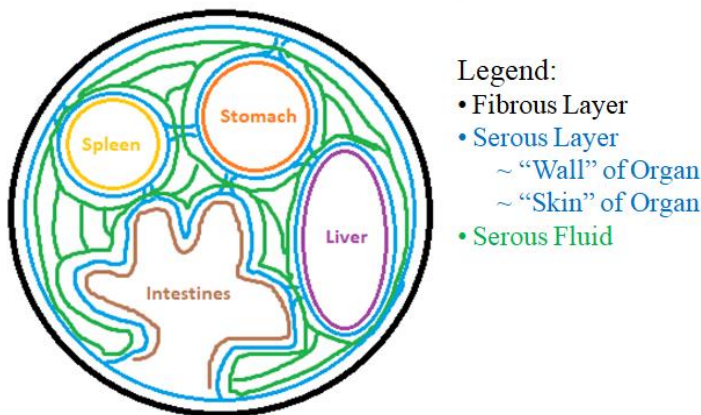


Figure 2. Recreation of Gil Hedley's drawing of visceral spaces (Hedley, 2023).

#### 1.0.4 Innervation of Viscera

The body's organs are intricately linked to the autonomic nervous system, which controls involuntary muscle functions. Every organ is controlled by two distinct nervous systems: the sympathetic system, which governs the fight or flight response, and the parasympathetic system, which manages the rest and digest response. Most organs receive parasympathetic signals through the vagus nerve and sympathetic signals via the greater and lesser splanchnic nerves. Both branches contain nociceptors that detect harmful stimuli and relay signals to the brain and spinal cord. Together, these elements of the autonomic nervous system continuously regulate bodily functions, playing an essential role in maintaining physiological balance and homeostasis.

## **2.0 Visceral-Somatic Interactions**

### 2.0.1 Visceral Attachment to the Somatic Frame

The viscera are interconnected with the somatic structure via various connective tissue forms, including fascia and suspensory ligaments. Collectively, the tissues collaborate to ensure stability by anchoring organs to the skeletal framework, which acts as a support system. Besides attaching body cavities to the somatic structure, fascia and ligaments link various organs within each cavity. For example, a fibrous fascial pouch encases the heart's parietal pericardium and connects to ligaments that link bones and other organs (Figure 3) (Barral et al., 2006). Furthermore, the hepatogastric ligament connects the liver to the stomach (Birchard, 2006). This connective tissue enables the movement of neural, lymphatic, and vascular structures, thereby ensuring effective communication and support among the internal organs and their association with the skeletal system.

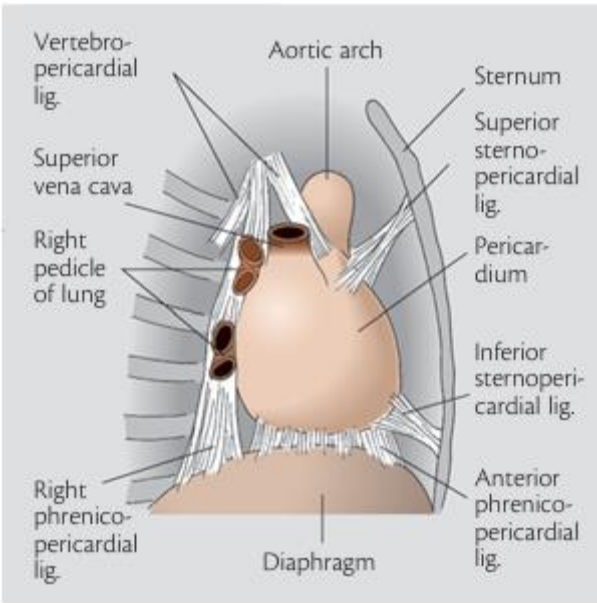


Figure 3. *The Pericardial Ligaments* (Barral et al., 2006).

## 2.0.2 Myofascial Kinetic Lines

The skeletal system has a strong visceral connection through myofascial kinetic lines, which are continuous strands of fascia and connective tissue linking the cranium to the distal limbs and connecting muscles, joints, and organs. The lines run cranially and caudally, connecting movements and providing stability, support, and compensatory posture.

A comparative dissection study has revealed the complex fascial connections between humans and horses, emphasizing the importance of suspensory ligaments as vital links bridging the visceral and parietal systems (Elbrønd et al., 2021). This connection is facilitated by a deep fascial line that traverses the body cavities, establishing a link between the somatic body and the viscera, linked to the stomatognathic system. The findings in horses resemble those in humans, but the unique quadrupedal structure leads to the term Deep Ventral Line (DVL). The DVL begins at the hindlimbs and tail, extending cranially through the pelvic cavity, into the abdominal

cavity, enveloping the gastrointestinal tract, then moving into the thoracic cavity and along the neck to the cranium (Figure 4). This pathway includes the nasal and oral cavities within the skull. It surrounds the vessels and nerves that supply the organs, forming a continuous network linking the inner organs, muscles, vessels, nerves, and skeletal system (Figure 5) (Elbrønd et al., 2021).

In the realm of osteopathy, if one segment of a myofascial line becomes tight, that tension can create a cascading effect throughout the interconnected cavities and organs, ultimately affecting the entire skeletal structure and its function.



*Figure 4. An overview of the deep equine myofascial kinetic lines - DDL (purple) and DVL (red) myofascial kinetic lines painted on a horse (Elbrønd et al., 2021).*

**Table 1.** Overview of some of the major fascia connections between the subserosal insertional fascia and the somatic/locomotor system in the horse.

Organ	Connecting structures	Connections to the somatic/locomot. system
Liver	<i>Lig. triangulare dexter</i> <i>Lig. Sinister</i>	<i>Diaphragma, Centrum tendineum</i> <i>M. transversus abdominis and aponeurosis</i> <i>Diaphragma pars costalis</i>
	<i>Lig. coronarius</i> <i>Lobus caudatus, proc. caud.</i>	<i>Centrum tendineum diaphragmatis</i> <i>M. psoas major et minor</i>
	<i>Lig. teres hepatis/falciformis</i>	<i>M. rectus abdominis</i> (the ventral part of Diaphragma)
Stomach	<i>Esophagus/Cardia,</i> <i>Omentum majus, lig. Gastrophrenicum</i>	The ventral part of the <i>crus dexter diaphragmatis</i> <i>Centrum tendineum, crus sinister diaphragmatis</i>
Intestinal tract		
<i>Intestinum tenue</i>	<i>Mesoduodenum, jejunum, ileum</i>	<i>M. psoas minor/major, Crus dexter/sinister diaphragmatis</i>
<i>Intestinum crassum</i>	<i>Caput caecum</i>	<i>Mm. psoas major et minor dexter,</i> <i>M. transversus abdominis</i>
<i>Rectum</i>	<i>Adventitia</i>	<i>M. sphincter ani externus</i> <i>M. levator ani</i> <i>M. sacrococ. ventralis</i> Indirectly to <i>M. biceps femoris</i> via connection to <i>Lig. sacroiliaca</i>
Lien/spleen	<i>Omentum majus, Lig. phrenicocolleale</i>	<i>Centrum tendineum, crus sinister diaphragmatis</i>
<i>Gl. adrenales</i>	<i>Adventitia</i>	<i>Mm. psoas major and minor, prox part</i>
Kidney	Subserosal fascia, dorsal part of the <i>capsula adiposa</i> , trabecular attachments from the <i>capsula fibrosa renalis</i>	<i>Mm. psoas major et minor</i> <i>M. transversus abdominis</i> <i>Diaphragma pars lumbalis</i> <i>Crus dexter diaphragmatis</i> (right kidney)
<i>Ureteres</i>	<i>Adventitia</i>	Ventrally along the <i>Mm. psoas major and minor</i>
<i>Ovarium</i>	Subserosal fascia of <i>Processus vaginalis Mesovarium</i>	<i>Mm. obliquus internus, externus and transversus abdominis</i> via <i>Anulus inguinalis</i>
	<i>Lig. ovarii proprii</i>	Via the <i>Lig. latum uteri</i> and to the <i>M. transversus abdominis</i> and ventral surface of <i>Mm. psoas major and minor</i>
<i>Testis</i>	Subserosal fascia of <i>Processus vaginalis</i> and <i>mesorchidium</i>	<i>Mm. obliquus internus, externus and transversus abdominis</i> via <i>Anulus inguinalis</i>
	Proximal connections from the site of origin, <i>mesonephros</i>	Connections alongside the <i>A. testicularis</i> to <i>Mm. psoas major and minor</i> to the offspring from the <i>Aorta</i> , cranial to the kidney
<i>Vesica urinaria</i>	<i>Ligg. vesicae lateralis</i> <i>Lig. vesicae mediana</i>	<i>M. transversus abdominis</i> <i>Fascia rectus, M. rectus abdominis</i>
<i>Uterus</i>	<i>Lig. latum uteri</i> <i>Lig. teres uteri</i>	Mid part of <i>M. transversus abdominis</i> <i>Mm. obliquus internus, externus and transversus abdominis</i> and <i>Anulus inguinalis</i>
<i>Ductus deferens</i>	<i>Plica ductus deferentis</i>	<i>Mm. obliquus internus, externus and transversus abdominis</i>

Figure 5. Key facial links between the viscera and the somatic system in horses (Elbrønd et al., 2021).

### 2.0.3 How Fascia Impacts Movement

Fascia is a web-like tissue that envelops and supports blood vessels, muscles, bones, and organs, facilitating bodily movement. It is rich in nociceptors and mechanoreceptors, responsible for detecting various stimuli and delivering sensory feedback to both the autonomic and somatic nervous systems. This network of sensory information provides proprioception and body awareness by relaying information to the brain about the position and movement of body parts. When unrestricted, fascia allows for effective force transmission and coordinated movement. However, adhesions within the fascia can restrict mobility and interfere with physical activities.

Fascia exhibits a remarkable ability to adapt in response to various mechanical stimuli such as contraction, stretching, and distension. It alters its tension to counteract external forces, thus maintaining an intra- and intramuscular balance throughout the body. Research indicates that myofascial tension generated by contracting muscles can significantly impact related tissues, potentially imposing limitations on the mobility of adjacent tendons and ligaments (Ajimsha et al., 2020; Krause et al., 2016; Leonard, 2013).

Consequently, given that fascia encompasses the entire body, it operates similarly to a pulley system; alterations in tension within one region can influence other areas, thereby facilitating optimal posture and motion. Under normal physiological conditions, myofascial lines interact smoothly, allowing for free movement among body structures. This harmonious interaction is important for maintaining functional integrity and mobility within the musculoskeletal and visceral system.

## 2.0.4 Viscerosomatic & Somaticvisceral Reflexes

Reflexes are vital for maintaining homeostasis, balance, and the body's overall functioning. They allow rapid adaptation to adverse situations, aiding in protection from harm through nerve involvement.

Myron C. Beal defines a viscerosomatic reflex as an autonomic response to stimuli from a visceral disorder that affects somatic tissues (Beal, 1985). The reflex arc begins with afferent sensory impulses from visceral receptors, sending information to an integrating center in the spinal cord through a spinal segment (Figure 6). This center then relays the sensory information to motor efferents, leading to responses in skeletal muscles, viscera, blood vessels, and skin. Somatic-visceral reflexes occur when somatic stimulation affects related viscera. Somatic reflexes involve skeletal muscles, while visceral reflexes involve soft tissue organs, with the former mediated by the somatic nervous system and the latter by the autonomic nervous system (Beal, 1985).

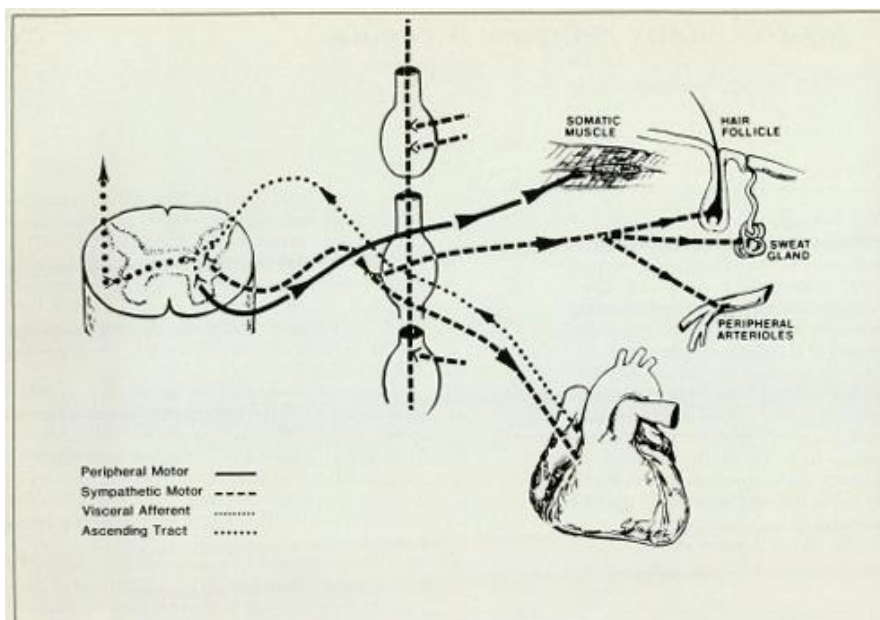


Figure 6. Schematic representation of the viscerosomatic reflex (Beal, 1985).

### **3.0 Integrative Approaches to Evaluation and Treatment**

The osteopathic profession emphasizes a holistic approach, viewing patients as dynamic units where somatic disturbances can affect local and distant areas. Key concepts include viscerosomatic and somatovisceral interactions, though the approach to understanding these interactions between somatic dysfunction and visceral disease remains debated. Additionally, much of the existing evidence is based on studies conducted in humans rather than animals. Nevertheless, it is believed that in the field of animal osteopathy, findings from human studies may have similar implications for animals.

#### **3.0.1 Osteopathic Evaluation of Viscera-Somatic Dysfunction**

For centuries, osteopathic neurophysiology has revolved around osteopathic lesions, a concept introduced by Dr. Andrew Taylor Still, the founder of osteopathy (Liem, 2016). He suggested that disruptions in the flow of life can lead to disease processes, which he described as lesions—changes in tissue texture, structure, or position (Liem, 2016). Over time, our understanding of these lesions has progressed into what we currently recognize as somatic dysfunction, a complex and multidimensional phenomenon.

Today, the concept of somatic dysfunction is characterized by impaired function in the skeletal, arthrodiar, and myofascial structures, along with their vascular, lymphatic, and neural components, as well as physiological adaptation (Consorti et al., 2023). Despite the evolving definitions, the osteopathic profession has consistently emphasized the structural-function relationships between the musculoskeletal and visceral systems. Some osteopathic theorists believe visceral diseases might manifest in the somatic system even before symptoms emerge,

providing diagnostic clues about visceral abnormalities within musculoskeletal structures. This phenomenon is believed to arise from segmental facilitation, changes in muscular activity, alterations in tissue texture, and changes in circulation (Bath et al., 2023; Beal, 1985; Eble, 1960; Kelso, 1971; Korr, 1947; Liem, 2016). Ultimately, segmental facilitation occurs through viscerosomatic convergence, emphasizing the biological connections between visceral and musculoskeletal components in cases of somatic dysfunction. This understanding can assist in diagnosing underlying visceral diseases when clinical signs are unclear—a topic that often sparks debate in contemporary osteopathic care.

Historically, manual practitioners believed that many cases of visceral disease or injury could lead to common somatic reflex patterns that can be identified through palpation. Early studies by J. Stedman Denslow, DO, and Irvin Korr, PhD, focused on examining sympathetic reflex responses involving somatic structures in humans, aiming to validate a spinal facilitation theory with a straightforward cause-and-effect model (Beal, 1985; Denslow et al., 1947; Korr, 1947). This concept has further been challenged over time. However, historical evidence shows that visceral stimulation can cause localized neurologic and functional changes in somatic structures due to segmental effects. Abnormal sensory signals originating from damaged or pathological areas of internal organs travel through afferent pathways to the spinal cord via viscerosomatic reflexes (Bath et al., 2023). This process alters neuronal activity in the corresponding vertebral segments and can also affect adjacent areas where the nerve supply to that organ or muscle originates. Additionally, it may lead to referred muscle pain (Giamberardino, 1999).

For instance, Beal and Wilson's research indicated that somatic dysfunction in the T1-T5 region was associated with cardiac disease, while dysfunction in the T5-T12 region was linked to gastrointestinal issues in humans (Beal, 1983; Beal, 1985). Beal also proposed that facilitation could occur when a reflex arc responds abnormally due to sustained overstimulation; if the involved neurons are already partially excited, less stimulation would be required to trigger an impulse (Beal, 1985). The reflex arc receives input from muscle somatic resources, such as Golgi tendons, but can also be influenced by signals from the brain. Additionally, these alterations could affect other spinal segments that may not be directly connected to the original injury. Therefore, through multiple studies, Korr, Beal, and Klieber observed that presymptomatic signs emerging from changes in viscerosomatic reflexes in individuals with diseases or developing visceral conditions were present (Beal, 1985; Beal et al., 1985; Korr, 1947). This leads to an important question for evaluation and treatment. Given that our understanding of anatomy shows a link between somatic dysfunction and the viscera, the theory of viscerosomatic relationships is likely valid, and patterns of palpatory fixation should be evident.

Motion palpation is typically recognized as the most widely used technique by manual therapists for differentiating abnormal from normal spinal mobility. Researchers in manual medicine have been investigating the reliability of various diagnostic tests based on palpation for many years. Previous clinical studies conducted on humans and small animals indicated that spinal facilitation associated with visceral diseases is connected to the autonomic nervous system's innervation and organ supply (Figure 7) (Beal, 1985). When plotting the autonomic reference sites, somatic reference areas related to the viscera were also identified (Figure 8) (Beal, 1985).

TABLE 3. VISCEROSOMATIC REFERENCE SITES FROM THE OSTEOPATHIC LITERATURE.		
Observer	Findings	Remarks
<b>HEART</b>		
Barstow (in Patriquin <sup>39</sup> )	T3-T5, upper ribs left	
Beal <sup>24,26</sup>	C2, T1-T5 left	
Beasley <sup>40</sup>	Ribs 1-2 left	
Becker <sup>41</sup>	T1-T6, ribs 3-5 left	
Burchett <sup>42</sup>	T1-T4	
Burns <sup>43</sup>	T3, T4	
Champlin and Champlin <sup>44</sup>	C2, C4, C5, T3, T4, ribs 3-6 left	A single case
Cox and associates <sup>45</sup>	C3, T4, T5	
Glascook (in Northup <sup>46</sup> )	T3, T4	
Hart <sup>47</sup>	C7-T5, most prominent T1-T3	
Johnson <sup>48</sup>	Rib 2 left	
Kelso and associates <sup>9</sup>	T1-T4	
Koch <sup>49</sup>	T2-T6	
Korr <sup>5</sup>	T1-T4	Low skin resistance areas
Larson <sup>25</sup>	C2 left, T2-T5	Left side predominance
Long <sup>50</sup>	T1-T10	
MacBain <sup>51</sup>	C1-C2, T2-T5	
Patriquin <sup>39</sup>	T1-T4, ribs 1-4 left	
Robuck <sup>52</sup>	T3, T4	
Singleton <sup>53</sup>	T1-T3, ribs 1, 2, 5	
Snyder <sup>54</sup>	T1-T4, T5 occasionally, ribs also	
Steunenberg <sup>55</sup>	C6-T5, upper ribs left	Myocarditis
A.T. Still symposium <sup>56</sup>	T3, T4	
Waitley <sup>57</sup>	T1, T6	
Walton <sup>58</sup>	T1-T4 left	
Wilson <sup>23</sup>	T1-T5, ribs left	
<b>LUNG</b>		
Bolton <sup>59</sup>	C3-C4, T4-T9	Chronic pulmonary disease
Burns <sup>60,61</sup>	T1-T3 T3-T5 T2-T7  T7-T10 T8-T10  T8-T11 T6-T10	Asthma 27 cases asthma Nervous control of lungs Tuberculosis 19 cases laryngeal tuberculosis 15 cases upper lobe tuberculosis 38 cases middle and lower lobe tuberculosis
Bush <sup>62</sup>	Ribs 5-8 right	Asthma
Crane <sup>63</sup>	T3-T4	Lobar pneumonia
Deming and Kruener <sup>7</sup>	C2-C4, T1-T6	183 cases disorder of the respiratory tract
Facto <sup>64</sup>	C2, C3, T3, T6	1 case bronchitis
Goode <sup>65</sup>	Rib 5	Asthma
Grainger <sup>66</sup>	T3-T4	Lobar pneumonia
Gravett <sup>67</sup>	C5, C6, rib 1, clavicle	Acute and chronic bronchitis
Hoag <sup>68</sup>	Upper thoracic area, especially T1-T6	Chronic lung disease
Howell and associates <sup>69,70</sup>	T3, right costotransverse articulation, T2 left Thoracic spine—extreme restricted	Chronic obstructive lung disease 1 case pulmonary disease
Keene <sup>71</sup>	T2-T5, ribs 1, 2 depressed T4-T5 right, ribs 4, 5 depressed right	1 case tuberculosis 1 case tuberculosis
Kline <sup>72</sup>	T2 right, T6-T7 left, rib 2	1 case tuberculosis
Koch <sup>73</sup>	T2-T8	Respiratory infections
Koch <sup>73</sup>	Cervical and upper thoracic spine	Asthma
Magoun <sup>74</sup>	T2-T4	Bronchi and lungs
McWilliams <sup>76</sup>	C6, C7, T3, T4, sacrum	Asthma
Wilson <sup>76-78</sup>	T4, T5 Occiput, T4, T5, ribs 4-5 bilaterally Rib 5 left	20 cases asthma Asthma 1 case asthma
<b>STOMACH</b>		
Northup <sup>79</sup>	T5	1 case chronic gastritis
A.T. Still symposium <sup>56</sup>	T5	
Burns <sup>80</sup>	T5-T10 C6, C7, T2-T4  C1, C2, T2, T6, T12  T9, 10, ribs involved T5-T7 left	Acute gastritis 1 case chronic gastritis 1 case chronic gastritis 1 case Ulcer, inflammation of stomach
Brigham <sup>81</sup>	T5-T9, inflammation of T5-T7	
Conley <sup>82-84</sup>	T5-T9, particularly T5, T6 T9-T12	Peptic ulcer Peptic ulcer
Gibson <sup>85</sup>	T5-T9 left	
Glascook (in Northup <sup>46</sup> )	T3 right T1, T3 right T6-T8, ribs right	Acute indigestion
Halladay <sup>86</sup>	T8	Gastric ulcer
Kranz <sup>87</sup>	T5-T7	Gastritis
Mattern <sup>88</sup>	T6, T7, upper cervical area, and T5-T10	
Magoun <sup>89</sup>	T5, T6 right T6-T8 left T5-T7 left	
Meyers <sup>90</sup>	T5, spastic neck	1 case gastric ulcer
Muttart <sup>91</sup>	T6-T10	Gastric ulcer
Waitley <sup>57</sup>	T5-T9, most importantly T5-T7	
<b>SMALL INTESTINE</b>		
Brigham <sup>81</sup>	T8, T9 T5-T7 left	Pyloric ulcer
Gibson <sup>85</sup>	T8-T10 left  T6-T9 left	Pylorus, inflammation of Duodenum, areas of referred pain
Kranz <sup>87</sup>	T5-T7	Duodenum, inflammation of
Magoun <sup>89</sup>	T5-T12, particularly T8, T9 T5-T7 right	Duodenum, functional conditions of
Martin <sup>93</sup>	T6, T7 left	Duodenal ulcer with cholecystitis

Figure 7. Viscerosomatic reference sites with associated visceral diseases (Beal, 1985).

Lindberg <sup>94</sup> Northup <sup>95</sup>	<b>LARGE BOWEL</b> T7,T8,T10,L5 Lower dorsal or lumbar area	Colitis Colitis			
Waitley <sup>97</sup> Woods <sup>96</sup>	T5,L3 T12-L2		1 case strangulated inguinal hernia, left indirect		
<b>APPENDIX</b>					
Brigham <sup>97</sup> Gibson <sup>83</sup> Glascock (in Northup <sup>46</sup> ) Kani <sup>98</sup> Laughlin <sup>99</sup> Magoun <sup>74,80,92</sup>	T9,T10 T9-T11 right S1 right  T11 Rib 10 right T9-T11 right T10-L2,tender spot T11 right		1 case appendicitis		
Millard <sup>101</sup>	Lowest thoracic and lumbar areas				
Smith <sup>102</sup> Wilson <sup>103</sup>	T9-T12 Group curve right, apex at T10	1 case appendicitis			
Woods <sup>96</sup>	T11-L1 bilaterally				
<b>LIVER</b>					
Magoun <sup>74,92</sup>	T6-T8 right T5-T7 right				
McWilliams <sup>76</sup> Peckham <sup>104</sup> Waitley <sup>97</sup> Wilson <sup>105</sup>	T8, midthoracic region right T9=2 segments (T7-T11) T5-T9 T10 right		1 case		
<b>GALLBLADDER</b>					
Becker <sup>106</sup>	Ribs,T5-T12	Vertebral lesion (flexion type)			
Bell <sup>107</sup> Brigham <sup>81</sup> Burns <sup>108</sup>	T6-T8 T5-T7 right T9-T10, vertebral and costal lesions, frequently rib of T10	1 case gallstones			
Conley <sup>84,106,110</sup>	Rib 9, T12, T5-T9,T7 most frequent location T7, pain to right of T8-T11 spinous process				
Denslow <sup>111</sup>	T8, L4-L5, left sacroiliac joint	1 case cholecystitis and colitis			
Downing <sup>112</sup>	T8-T10, including ribs, especially costovertebral joint right				
Gibson <sup>85</sup> Magoun <sup>74,80,92</sup>	T6-T8 right T7 right T5-T7 right T6-T8 right				
Malone <sup>113</sup> McWilliams <sup>76</sup>	T5-T8, ribs bilaterally T7,T8 midthoracic region right				
Northup <sup>79,114</sup> Starks <sup>115</sup>	Rib 8 right T8-T10 rigidity and immobility, including ribs	1 case cholecystitis			
A.T., Still symposium <sup>86</sup> Townsend <sup>116</sup> Waitley <sup>97</sup> Wilson <sup>117,118</sup>	T10  T5-T10 and ribs T5-T9 T6-T9, ribs right C3,T8				
<b>SPLEEN</b>					
Gibson <sup>80</sup>	T8-T11 left				
<b>PANCREAS</b>					
Gutensohn <sup>119</sup>	T6-T11	Paravertebral tenderness			
Magoun <sup>80</sup> Wilson <sup>120</sup>	T5-T10 left Occiput,T7,T10			Diabetes	
<b>KIDNEY</b>					
Burns <sup>61</sup> Barstow (in Conn <sup>121</sup> ) Blackslee (in Conn <sup>121</sup> ) Ellis <sup>122</sup>	T12-L1 T11  T12  Lower thoracic area, upper lumbar region, 3 lower ribs, frequently rib 12			Renal tuberculosis Nephritis	11 cases
Gibson <sup>85</sup> Magoun <sup>74,80,92</sup>	T9-T11 T11-T12 T10-L1 T10-T12				
Nelson <sup>123,124</sup>	T5-L3, principal area T9-T11, most specifically T10				
Smith <sup>125</sup>	T7-L3, flat, extreme rigidity				
Strachan <sup>126</sup>	T10-T12				
<b>URINARY BLADDER AND URETER</b>					
Gibson <sup>85</sup>	Lumbar and sacral areas bilaterally			Bladder	
	Lumbar and sacral areas			Ureter	
Wilson <sup>127</sup>	L5				
<b>PROSTATE GLAND</b>					
Gibson <sup>85</sup>	Sacrococcygeal area bilaterally				
Glascock (in Northup <sup>46</sup> ) Wilson <sup>127</sup>	L3,L4 L5			1 case prostate and bladder disease	
<b>UTERUS AND OVARIES</b>					
Burns <sup>61</sup>	T8-T10			Tuberculosis of the uterine tubes	
Detwiler <sup>128</sup>	C2,C3,T9-T11			1 case of dysmenorrhea	
Gibson <sup>86</sup>	L5 left innominate L4-S2			Ovarian and tubal disease	
	Sacrococcygeal junction			Uterus	
Glascock (in Northup <sup>46</sup> ) Hitchcock <sup>129</sup>	L1 T8-T12,L1-L2, S3,S4			Menstruation	Primary dysmenorrhea
Laughlin <sup>99</sup>	Sacroiliac C1,C2,ribs 1,2 right L1 spastic muscles of lower thoracic and lumbar areas			1 case fibroids 1 case cystic right ovary	1 case left ovarian and tubal disease
Magoun <sup>74</sup> Northup <sup>79</sup>	12 rib right, L1 Lumbar area T12,L1,L2			Retroverted uterus	Pelvic organs
Simmons <sup>130</sup>	T9-T12,L1 L3-L5,sacroiliac			1 case	dysmenorrhea
	T10,11 right			Ovaries, dysfunction of	Uterus, dysfunction of
Woods <sup>96</sup>	T10-L1,L4,L5			1 case left ovarian and tubal disease	1 case ruptured left ectopic pregnancy, right ovarian cyst

Figure 7. Viscerosomatic reference sites with associated visceral diseases (Beal, 1985).

Author(s)	Heart	Lungs	Esophagus	Stomach	Small intestine	Large intestine (splenic flexure)	Large intestine (splenic flexure to rectum)	Appendix	Liver	Gallbladder
Gray <sup>32</sup>	T1-T5	T2-T4	T5-T6	T6-T10	T9-T10	T11-L1	L1-L2		T7-T9	T7-T9
House and Pansky <sup>33</sup>	T1-T5	T2-T5		T6-T10	T6-T10	T6-T10	T6-T10		T5-T6	
Crosby and associates <sup>34</sup>	T1-T5 (T6)	T3-T4 (T5)	T5-T6	T7-T9	T9-T12	T9-T12			T7-T8	T9-T10 (R)
Bhagat and associates <sup>35</sup>	T1-T5	T2-T7	T5-T6	T6-T9	T9-T10	T11-L1	L1-L2	T10-T12	T9-T10	T9-T10
Pottenger <sup>36</sup>	T2-T8	T4-T9		T7-T9	T9-T12				T7-T10	T8-T9
Brodal <sup>8</sup>	T1-T4 (T5)	T2-T7 (T2-T4)		T6-T10 (T5-T11) (T5-T9L)	T6-T10 (T5-T11)		L1-L2	T10-T12	T7-T9 (R)	T7-T9 (R)
White <sup>37</sup> (afferent)	T1-T3			(T6) T7,T8 (T9)	T9,T10 (T11)	(T11) T12-L1	S2-S4		(T6) T7,T8 (T9)	(T6) T7,T8 (T9)
Bonica <sup>30</sup> (afferent)	T1-T4 (T5)	T2-T7	Upper T2-T7 (T8)	T6-T9	T6-T8 (T10) duodenum T9-T11 jejunem ileum	T12-L1	L1-L2	T10-T12	T5-T9	T5-T9
Bonica <sup>30</sup> (efferent)	T1-T4 (T5)	T2-T7	Upper T2-T4 Lower T5-T7	T6-T9 (T10)	T6-T11	(T11) T12-L1	L1-L2	T10-T12	T6-T9 (T10)	T5-T9 (T10)

\* The segments shown in parentheses indicate less frequent findings. L = left; R = right.

Figure 8. A summary of viscerosomatic referral zones according to segmental nerve distribution (Beal, 1985).

Spleen	Pancreas	Kidney	Ureter	Adrenal cortex	Adrenal medulla	Testes, Ovaries	Epididymis	Urinary bladder	Prostate gland	Uterus	Ureterine tube
T6-T10	T6-T10	T10-L1	T11-L2	T8-L1		T10-T11	T11-T12	T11-L2	T12-L1	T12-L1	T10-L1
	T5-T6 T7-T9	T10-T11 T11, T12-L1	T11, T12-L2			T9-T10		T11-L2 T11, T12-L1 (L2)	L1-L2 (L3)	T10-L2	
T6-T10	T6-T10	T11-L1 T11-L1	L1-L2			T10-T11 T10		T11-L2		T10-L1 T12-L1	
T6-T10 (T5-T11)	T6-T10 (T5-T11)	T12-L1 (T11-L2)	L1-L2 (T11-T12)		T11-L1 (T10-L2)			L1-L2 (T11-L2)			
		(T11) T12-L1	L1-L2					S2-S4		T11-L1 S2-S4	
	T6-T10	T11, T12-L1	T11-L2			T10-L1		T11-L1	T10-L1	T10-T12 (L1)	
	T6-T10	(T10) T11-L1 (L2)	T11-L2	T6-L2		T6-L1		T11-L2	T6-L1	T6-L1	

Figure 8. A summary of viscerosomatic referral zones according to segmental nerve distribution (Beal, 1985).

The research demonstrates that dysfunction in thoracic cavity organs is reflected in the lower cervical to upper thoracic vertebrae, abdominal organs through the mid-thoracic vertebrae, large colons via the lower thoracic vertebrae, and pelvic cavity organs through the lumbar and sacral regions. In addition to changes in spinal segments, Korr, Larson, and Eble found that continuous hyperirritability of stimulated viscera is reflected in altered muscular activity, leading to changes in the texture of muscles, paravertebral tissues, connective tissue, and skin, along with temperature variations (Beal, 1985; Eble, 1960; Korr, 1947; Larson, 1976). The location of the muscular response also varies depending on the stimulated organ (Beal, 1985; Eble, 1960). Additionally, Kelso's examination showed that many visceral diseases exhibit multiple areas with more segmental findings (Kelso, 1971). It was also noted that unpaired organs would likely show more findings on one side (Kelso, 1971). Furthermore, the extent of affected spinal segments correlates with the disease duration, and reflex patterns may intensify over time (Beal, 1985). Common physical stress responses are seen in acute cases, while chronic cases may show increased muscle contraction, skin thickening, and hypersensitivity (Beal, 1985).

From history, one might assume irrefutable evidence supports palpation as a diagnostically valuable tool for identifying direct visceral dysfunctions. However, this assumption is not currently substantiated. The ability of manual therapists to accurately identify true visceral dysfunction remains controversial. This may be due to studies that have found significant associations between spinal TART (Tissue, Asymmetry, Restriction, Tenderness) dysfunction and visceral dysfunction, yet the absence of standard control groups makes it challenging to establish meaningful relationships between specific visceral abnormalities and

somatic dysfunction in various body regions. As a result, these studies are considered to be inadequately researched.

Moreover, somatic dysfunction can create overt signs and symptoms that mimic or simulate internal organ disease, even though the source may be elsewhere in the body (Nansel, 1995). For example, in a horse, a fascial line runs from the stifle to the kidney on the same side. If dysfunction originates at the stifle, it can lead to restrictions in the fascia that extend up to the lower back, which may also cause a pain response in this area. This restriction may hinder the proper movement of the kidneys, leading to an abnormal dysfunction in visceral movement. However, the root cause of this dysfunction would be the stifle, not a visceral disease. Although there may not be a neurological explanation for the stifle's impact on the vertebrae, there is a valid fascial reason for this connection.

Additionally, strains in the connective tissue of the viscera can result from various factors, including surgical scars, adhesions, illnesses, posture, or injury. These strains lead to tension patterns within the deep fascial network of the body, which can create far-reaching effects and necessitate compensatory mechanisms. This compensation produces fixed, abnormal points of tension that the body must navigate around, and the resulting chronic irritation can lead to both functional and structural issues, potentially simulating organ disease.

Somatic responses to visceral disease can also vary uniquely among individuals. These include the location and number of spinal segments involved, the intensity of muscle contractions and tissue changes, pain levels, and whether the involvement is unilateral or

bilateral (Beal, 1985). There are also phenomena that science cannot fully explain. This may be why several theories correlating somatic dysfunction patterns with visceral dysfunction have been proposed in an attempt to elucidate the possible mechanisms behind patients presumed to be suffering from various internal organ diseases.

Consequently, the osteopathic profession emphasizes a holistic approach, viewing the patient as a dynamic unit where disturbances in the somatic system can have local and distant effects. Thus, considering the concepts of viscerosomatic and somatovisceral interactions are key elements that reinforce this holistic perspective.

### 3.0.2 Osteopathic Approaches to Treatment

Differentiating somatic and visceral dysfunction is challenging for osteopathic physicians and manual therapists, with ongoing debate about its clinical relevance. While simplistic cause-and-effect models are often criticized, it may be wise to stick to osteopathic principles until more substantial evidence is available, despite the anatomical connections between somatic dysfunction and visceral organs. Arguably, manual therapy can be divided into two primary conceptual approaches: 1) specific adjustments to correct anatomical or biomechanical issues that affect function through established musculoskeletal or nerve compromises, and 2) specific adjustments intended for physiological regulation that may involve alterations in internal organ function.

Musculoskeletal issues are common patient complaints in osteopathy and can be either a primary issue or a secondary effect of other conditions. How these treatments are applied

involves an understanding of various specialties and integrating OMT methods, enabling practitioners to see the body as a complete entity. Among the multiple approaches, the Cranial Sacral method involves practitioners who believe they can feel and influence the intrinsic rhythmic movements of the brain, which leads to fluctuations in cerebrospinal fluid and specific changes among dural membranes, cranial bones, and the sacrum (Guillaud, et al. 2016). However, the structural approach is arguably the most widely recognized aspect of osteopathy for addressing musculoskeletal issues, focusing on the musculoskeletal system to correct structural problems in joints, muscles, and tissues through techniques such as massage, stretching, and manipulation.

Low back pain is commonly seen in horses, and specific treatment methods have been shown to manage biomechanical disorders of the spine and alleviate pain effectively (Haussler et al., 2021). A study examined the effects of musculoskeletal mobilization in horses suffering from disease-related back pain, measuring thoracolumbar pain sensitivity, trunk stiffness, spinal reflexes, joint range of motion, thermographic imaging, and muscle hypertonicity. The results indicated low to moderate-quality evidence that structural manipulation could alleviate pain, stiffness, and muscle hypertonicity. However, among actively competing horses with acute back pain, using a precision thrust technique alone proved less beneficial than combining various structural treatment methods (Haussler et al., 2021). Therefore, osteopaths often employ additional assessment and treatment approaches to enhance patient outcomes.

Core osteopathic principles emphasize that an integrated approach to patient evaluation and treatment involves understanding structural connections between the viscera and the

musculoskeletal system. In the past fifty years, newer treatment modalities focusing on assessing and managing visceral organs have emerged (Guillaud et al., 2018; Roberts et al., 2022). While structural OMT significantly influences internal organ function by manipulating the musculoskeletal system, Osteopathic Visceral Manipulation (VOMT) specifically addresses the functioning of visceral organs and their surrounding environments. This technique can impact structural and physiological dysfunction across various body systems, including the musculoskeletal, vascular, nervous, urogenital, respiratory, digestive, eliminatory, neuroendocrine, and lymphatic systems (The Barral Institute, (n.d.)). Research on VOMT in animals remains limited, but many human studies have been conducted that could also inform practices with animals. Evidence suggests that visceral techniques have a positive effect on conditions such as irritable bowel syndrome, constipation, and gastroesophageal reflux (Fleischmann et al., 2020; Ignatowicz et al., 2017; Lotfi et al., 2023; Snider et al., 2016). Additionally, research has explored the impact of visceral techniques on musculoskeletal issues like low back pain, concussion, pregnancy-related back pain, and cervical spine pain (Franke et al., 2017; Horton, 2015; Tamer et al., 2017; Verhaeghe et al., 2018; Wetzler et al., 2017).

Each of the osteopathic treatment approaches show benefits for somatic dysfunction when applied individually. Still, their combined use, especially with an emphasis on visceral function, has significantly enhanced outcomes in pain relief, physical functionality, and overall health. A study by Tamer, Oz, and Ugler evaluated the effectiveness of OMT against VOMT for chronic non-specific low back pain (Tamer et al., 2017). The results suggested that OMT influences corticospinal changes related to somatic function and pain by altering the sensitivity of reflex pathways due to biomechanical stress on muscle spindles. This response quickly

reduces pain, aligning with the gate control theory (Dickenson, 2002). Moreover, research on VOMT revealed that muscle contractions between the thoracic vertebrae and the lumbosacral joint occur due to internal organ stimulation (Tamer et al., 2017). Thus, the added stimuli from visceral applications may assist in easing spasms in associated segments and modulate both peripheral and central pathways through the visceral-somatic reflex arc, promoting healing. Utilizing VOMT techniques alongside visceral interventions improved blood and fascial circulation, aided in removing bodily fluids, and boosted overall energy levels, leading to favorable physiological changes (Tamer et al., 2017). Therefore, when examining the relationship between visceral function, fascial limitations, and musculoskeletal dysfunction, the integrative approach of OMT paired with a focus on VOMT proves beneficial.

## **Conclusion**

In conclusion, the various influences on somatic findings indicate that facilitation, recruitment, variation in visceral-afferent innervation, and learning physical signs should be part of osteopathic theories of disease. Recognizing the connection between somatic dysfunction and visceral disease is essential for a holistic approach to patient care. The interplay of viscerosomatic reflexes highlight how issues within internal organs can lead to physical manifestations, guiding practitioners to pinpoint underlying health concerns. Practitioners can enhance treatment effectiveness and improve patient outcomes by conducting thorough assessments and employing various osteopathic techniques focused on visceral health. Embracing these principles reinforces the commitment to patient-centered care and aligns with ongoing research to deepen our understanding of complex health challenges. Ultimately,

combining visceral techniques into OMT promotes a comprehensive view of the body as an interconnected system, optimizing patient care.

## **References**

- Ajimsha, M. S., Shenoy, P. D., & Gampawar, N. (2020). *Role of fascial connectivity in musculoskeletal dysfunctions: A narrative review*. *Journal of Bodywork and Movement Therapies*, 24(4), 423–431. <https://doi.org/10.1016/j.jbmt.2020.07.020>
- Barral, J.-P., & Mercier, P. (2006). Chapter 2 / Thoracic Cavity. In *Visceral Manipulation* (pp. 32–38). essay, Eastland Press. chrome-extension://efaidnbmnnnibpcajpcglclefindmkaj/<https://www.barralinstitute.com/docs/marketing-materials/vmiexcerpt.pdf>
- Bath, M., & Owens, J. (2023). Physiology, Viscerosomatic Reflexes. In StatPearls. StatPearls Publishing. <https://www.ncbi.nlm.nih.gov/books/NBK559218/>
- Beal, M. C. (1983). *Palpatory testing for somatic dysfunction in patients with cardiovascular disease*. *The Journal of the American Osteopathic Association*, 82(11), 822–831. [https://www.researchgate.net/publication/16846530\\_Palpatory\\_testing\\_for\\_somatic\\_dysfunction\\_in\\_patients\\_with\\_cardiovascular\\_disease](https://www.researchgate.net/publication/16846530_Palpatory_testing_for_somatic_dysfunction_in_patients_with_cardiovascular_disease)
- Beal, M. C. (1985). *Viscerosomatic reflexes: a review*. *The Journal of the American Osteopathic Association*, 85(12), 786–801. [https://www.researchgate.net/publication/19321049\\_Viscerosomatic\\_reflexes\\_A\\_review](https://www.researchgate.net/publication/19321049_Viscerosomatic_reflexes_A_review)
- Beal, M. C., & Kleiber, G. E. (1985). *Somatic dysfunction as a predictor of coronary artery disease*. *The Journal of the American Osteopathic Association*, 85(5), 302–307. [https://www.researchgate.net/publication/19321049\\_Viscerosomatic\\_reflexes\\_A\\_review](https://www.researchgate.net/publication/19321049_Viscerosomatic_reflexes_A_review)

- Birchard, S. J. (2006). Chapter 72 - Surgery of the Liver and Biliary Tract. In Saunders Manual of Small Animal Practice (Third Edition, pp. 810–810). essay, W.B. Saunders. Retrieved from chrome-extension://efaidnbmnnnibpcajpcglclefindmkaj/https://www.sciencedirect.com/sdfe/pdf/download/eid/3-s2.0-B0721604226500747/first-page-pdf.
- Consorti, G., Castagna, C., Tramontano, M., Longobardi, M., Castagna, P., Di Lernia, D., & Lunghi, C. (2023). Reconceptualizing Somatic Dysfunction in the Light of a Neuroaesthetic Enactive Paradigm. *Healthcare (Basel, Switzerland)*, 11(4), 479. <https://doi.org/10.3390/healthcare11040479>
- Denslow, J. S., Korr, I. M., & Krems, A. D. (1947). *Quantitative studies of chronic facilitation in human motoneuron pools*. *The American journal of physiology*, 150(2), 229–238. <https://doi.org/10.1152/ajplegacy.1947.150.2.229>
- Dickenson, A. H. (2002). *Gate control theory of pain stands the test of time*. *British journal of anaesthesia*, 88(6), 755–757. <https://doi.org/10.1093/bja/88.6.755>
- Eble, J. N. (1960). *Patterns of response of the paravertebral musculature to visceral stimuli*. *The American journal of physiology*, 198, 429–433. <https://doi.org/10.1152/ajplegacy.1960.198.2.429>
- Elbrønd, V.S. and Schultz, R.M. (2021) *Deep Myofascial Kinetic Lines in Horses, Comparative Dissection Studies Derived from Humans*. *Open Journal of Veterinary Medicine*, 11, 14-40. <https://doi.org/10.4236/ojvm.2021.1111002>

- Fleischmann, M., Vaughan, B., Fitzgerald, K., & Grace, S. (2020). *Use of manual therapy applied to the viscera: Secondary analysis of a nationally representative sample of Australian osteopaths*. *International Journal of Osteopathic Medicine*, 36, 19–25.  
<https://doi.org/10.1016/j.ijosm.2020.05.002>
- Franke, H., Franke, J. D., Belz, S., & Fryer, G. (2017). *Osteopathic manipulative treatment for low back and pelvic girdle pain during and after pregnancy: A systematic review and meta-analysis*. *Journal of bodywork and movement therapies*, 21(4), 752–762.  
<https://doi.org/10.1016/j.jbmt.2017.05.014>
- Fryer, G. (2017). *Integrating osteopathic approaches based on biopsychosocial therapeutic mechanisms. part 2: Clinical approach*. *International Journal of Osteopathic Medicine*, 26, 36–43. <https://doi.org/10.1016/j.ijosm.2017.05.001>
- Gatt, A., Agarwal, S., & Zito, P. M. (2023). *Anatomy, Fascia Layers*. In StatPearls. StatPearls Publishing. <https://pubmed.ncbi.nlm.nih.gov/30252294/>
- Giamberardino, M. A., Affaitati, G., Iezzi, S., & Vecchiet, L. (1999). *Referred Muscle Pain and Hyperalgesia from Viscera*. *Journal of Musculoskeletal Pain*, 7(1–2), 61–69.  
[https://doi.org/10.1300/J094v07n01\\_07](https://doi.org/10.1300/J094v07n01_07)
- Guillaud, A., Darbois, N., Monvoisin, R., & Pinsault, N. (2016). *Reliability of Diagnosis and Clinical Efficacy of Cranial Osteopathy: A Systematic Review*. *PloS one*, 11(12), e0167823. <https://doi.org/10.1371/journal.pone.0167823>

- Guillaud, A., Darbois, N., Monvoisin, R., & Pinsault, N. (2018). Reliability of diagnosis and clinical efficacy of visceral osteopathy: a systematic review. *BMC complementary and alternative medicine*, 18(1), 65. <https://doi.org/10.1186/s12906-018-2098-8>
- Hafen, B. B., & Shook, M. (2023, July 17). Anatomy, Smooth Muscle. StatPearls. <https://www.ncbi.nlm.nih.gov/books/NBK532857/>
- Haussler, K. K., Hesbach, A. L., Romano, L., Goff, L., & Bergh, A. (2021). *A Systematic Review of Musculoskeletal Mobilization and Manipulation Techniques Used in Veterinary Medicine*. *Animals: an open access journal from MDPI*, 11(10), 2787. <https://doi.org/10.3390/ani11102787>
- Hedley, G. (2023, May 8). How Do Visceral Fasciae Structure Visceral Space? Learn Integral Anatomy with Gil Hedley. YouTube. <https://www.youtube.com/watch?v=rYtADunTly8&t=139s>
- Horton, Ramona. (2015). *The anatomy, biological plausibility and efficacy of visceral mobilization in the treatment of pelvic floor dysfunction*. *Journal of Pelvic, Obstetric Gynecological Physiotherapy*. 117. 5-18. [https://www.researchgate.net/publication/295549504\\_The\\_anatomy\\_biological\\_plausibility\\_and\\_efficiency\\_of\\_visceral\\_mobilization\\_in\\_the\\_treatment\\_of\\_pelvic\\_floor\\_dysfunction](https://www.researchgate.net/publication/295549504_The_anatomy_biological_plausibility_and_efficiency_of_visceral_mobilization_in_the_treatment_of_pelvic_floor_dysfunction)
- Ignatowicz, A., & Berkowitz, M. R. (2017). *Imaging evidence demonstrating effectiveness of osteopathic visceral manipulation techniques in treating pseudo-obstruction*. *The AAO Journal*, 27(1), 7–10. <https://doi.org/10.53702/2375-5717-27.1.7>

Kelso, A. F. (1971). *A double-blind clinical study of osteopathic findings in hospital patients: progress report*. The Journal of the American Osteopathic Association, 70(6), 570–592.

<https://ostemed-dr.contentdm.oclc.org/digital/api/collection/myfirst/id/4727/download>

Korr, I. M. (1947). *The neural basis of the osteopathic lesion*. The Journal of the American Osteopathic Association, 47(4), 191–198.

[chrome-extension://efaidnbmninnkjkpcjpcglclefindmkaj/https://www.claudio-lopez-osteopata.it/documenti/C.Lopez%20Osteopata%20-%20Osteopatia%20e%20Neurologia%20-%20Bibliografia%20\[7\].pdf](chrome-extension://efaidnbmninnkjkpcjpcglclefindmkaj/https://www.claudio-lopez-osteopata.it/documenti/C.Lopez%20Osteopata%20-%20Osteopatia%20e%20Neurologia%20-%20Bibliografia%20[7].pdf)

Krause, F., Wilke, J., Vogt, L., & Banzer, W. (2016). *Intermuscular force transmission along myofascial chains: a systematic review*. Journal of anatomy, 228(6), 910–918.

<https://doi.org/10.1111/joa.12464>

Langevin, H. M. (2006). *Connective tissue: a body-wide signaling network?*. Medical hypotheses, 66(6), 1074–1077. <https://doi.org/10.1016/j.mehy.2005.12.032>

Larson, N. J. (1976). *Summary of site and occurrence of paraspinal soft tissue changes of patients in the intensive care unit*. The Journal of the American Osteopathic Association, 75(9), 840–842.

<https://ostemed-dr.contentdm.oclc.org/digital/api/collection/myfirst/id/5594/download>

Leonard, J. (2013). *Importance of Considering Myofascial Force Transmission in*

*Musculoskeletal Surgeries*. Journal of Surgical Academia. Retrieved from [chrome-extension://efaidnbmninnkjkpcjpcglclefindmkaj/https://www.jsurgacad.com/sites/default/files/article/2013/01-Editorial%20\(1\)\\_0.pdf](chrome-extension://efaidnbmninnkjkpcjpcglclefindmkaj/https://www.jsurgacad.com/sites/default/files/article/2013/01-Editorial%20(1)_0.pdf).

- Liem, T. (2016). *A.T. Still's Osteopathic Lesion Theory and Evidence-Based Models Supporting the Emerged Concept of Somatic Dysfunction*. The Journal of the American Osteopathic Association, 116(10), 654–661. <https://doi.org/10.7556/jaoa.2016.129>
- Lotfi, C., Blair, J., Jumrukovska, A., Grubb, M., Glidden, E., & Toldi, J. (2023). Effectiveness of Osteopathic Manipulative Treatment in Treating Symptoms of Irritable Bowel Syndrome: A Literature Review. *Cureus*, 15(7), e42393. <https://doi.org/10.7759/cureus.42393>
- MacCord, K. (2013, September 17). Germ layers. \*Embryo Project Encyclopedia\*. <https://hdl.handle.net/10776/6273>
- Nansel, D., & Szlazak, M. (1995). *Somatic dysfunction and the phenomenon of visceral disease simulation: a probable explanation for the apparent effectiveness of somatic therapy in patients presumed to be suffering from true visceral disease*. *Journal of manipulative and physiological therapeutics*, 18(6), 379–397. [https://chiro.org/Subluxation/Visceral\\_Disease\\_Simulation.shtml](https://chiro.org/Subluxation/Visceral_Disease_Simulation.shtml)
- O'Neill, K. (2024, April 16). The Peritoneum. TeachMeAnatomy. <https://teachmeanatomy.info/abdomen/areas/peritoneum/>
- Roberts, A., Harris, K., Outen, B., Bukvic, A., Smith, B., Schultz, A., Bergman, S., & Mondal, D. (2022). Osteopathic Manipulative Medicine: A Brief Review of the Hands-On Treatment Approaches and Their Therapeutic Uses. *Medicines (Basel, Switzerland)*, 9(5), 33. <https://doi.org/10.3390/medicines9050033>

Snider, K. T., Schneider, R. P., Snider, E. J., Danto, J. B., Lehnardt, C. W., Ngo, C. S., Johnson, J. C., & Sheneman, T. A. (2016). *Correlation of Somatic Dysfunction With Gastrointestinal Endoscopic Findings: An Observational Study*. *The Journal of the American Osteopathic Association*, 116(6), 358–369.

<https://doi.org/10.7556/jaoa.2016.076>

Tamer, S., Öz, M., & Ülger, Ö. (2017). *The effect of visceral osteopathic manual therapy applications on pain, quality of life and function in patients with chronic nonspecific low back pain*. *Journal of back and musculoskeletal rehabilitation*, 30(3), 419–425.

<https://doi.org/10.3233/BMR-150424>

The Barral Institute. (n.d.). Therapeutic value of visceral manipulation.

<https://www.barralinstitute.com/therapies/>

Verhaeghe, N., Schepers, J., van Dun, P., & Annemans, L. (2018). Osteopathic care for low back pain and neck pain: A cost-utility analysis. *Complementary Therapies in Medicine*, 40, 207–213. <https://doi.org/10.1016/j.ctim.2018.06.001>

Wetzler, G., Roland, M., Fryer-Dietz, S., & Dettmann-Ahern, D. (2017). *Craniosacral Therapy and Visceral Manipulation: A New Treatment Intervention for Concussion Recovery*. *Medical acupuncture*, 29(4), 239–248. <https://doi.org/10.1089/acu.2017.1222>