PREOMICS

iST 4x

P.O.00061; P.O.00176

Pelleted cells & precipitated protein

exorein preparation Solutions

Introduction

Sample preparation is one of the essential steps of bottom-up proteomics. The PreOmics® iST sample preparation kit is designed to streamline this process, enabling researchers to achieve optimal results with reduced steps and hands-on time. For sample-specific protocols and optimization, visit www.preomics.com/resources or contact info@preomics.com.

Kit Contents

The kit includes all essential components for proteomic sample preparation: denaturing, reducing, and alkylating agents, enzymes, cartridges, and wash buffers for peptide cleanup.

Component	Сар	Quantity	Buffer Properties		s	Description	Storage	
			Organic	Acidic	Basic	Neutral		
DIGEST		1x					Trypsin/LysC mix to digest proteins.	-20°C
RESUSPEND	\bigcirc	1x 2 mL				•	For reconstituting lyophilized proteolytic enzymes.	RT
LYSE		1x 1 mL			•		For denaturing, reducing, and alkylating proteins.	RT
STOP		1x 1 mL	•	•			For stopping enzymatic activity.	RT
WASH 1		1x 2 mL	•	•			For removing hydrophobic contaminants.	RT
WASH 2		1x 2 mL		•			For removing hydrophilic contaminants.	RT
ELUTE		1x 2 mL	•		•		For eluting peptides from the cartridge.	RT
LC-LOAD	\circ	1x 1 mL		•			For loading peptides on reversed-phase LC-MS column.	RT
CARTRIDGE		4x					Cartridges with SPE sorbent for peptide purification from 1–100 μg protein starting material.	RT
WASTE TUBE		4x					2 mL tube for collecting waste after washing steps.	RT
COLLECTION TUBE		4x					1.5 mL tube for collecting peptides after elution.	RT
ADAPTER		4x					Enables a cartridge to be placed into a tube.	RT

Pre-Requisites

Common lab equipment is required for the sample preparation.

Equipment	Quantity and Description
PIPETTE	Careful sample handling and pipetting reduces sample contaminations and improves quantification.
SAMPLE	Pelleted cells or precipitated protein $(1-100~\mu g$ protein starting material; max. $10~\mu L$ sample volume). For higher sample volumes, visit our FAQs. For other sample types, contact PreOmics for adapted protocols.
REACTION TUBE	1.5 mL microcentrifuge low protein binding tubes are recommended (e.g., Eppendorf Protein LoBind® Tubes, catalogue number EU: 0030108116, US: 022431081).
HEATING SHAKER	Two heating shakers for tubes are recommended for protein denaturation and digestion.
CENTRIFUGE	Benchtop centrifuge for 1.5 or 2 mL tubes is required for peptide loading, washing, and elution.
SONICATOR or NUCLEASE	If the sample is viscous, use a sonicator to shear DNA (e.g., Diagenode Bioruptor®) or add nuclease (e.g., Benzonase®) to degrade both DNA and RNA. Visit our FAQ for more information.
VACUUM EVAPORATOR	To evaporate volatile buffers from the eluate before LC-MS.
ULTRASONIC BATH	Optional: can be used to resuspend peptides.

Procedure



Method

1. LYSE

- 1.1. Add 50 μL LYSE to 1–100 μg of protein sample in a REACTION TUBE, place it in a pre-heated HEATING SHAKER (95°C; 1,000 rpm; 10 min). *NOTE1*
- 1.2. Optional: Spin down droplets (RT; max. 300 rcf; 10 sec).
- 1.3. If sample is viscous, use a SONICATOR or add NUCLEASE.
- 1.4. Allow samples to cool down to room temperature.

2. DIGEST

- 2.1. Optional: Spin down lyophilized enzyme mix in the DIGEST tube (RT; max. 300 rcf; 10 sec).
- 2.2. Add 210 μL **RESUSPEND** to **DIGEST** (1 tube for 4 reactions), shake (RT; 500 rpm; 10 min) and pipette up/down. *NOTE2*
- 2.3. Add 50 μL **DIGEST** to REACTION TUBE and place it in a pre-heated HEATING SHAKER (37°C; 500 rpm; 1–3 h).
- 2.5 Assemble ADAPTER and CARTRIDGE to the top of WASTE TUBE. Label all tubes and transfer sample to the CARTRIDGE.

3. PURIFY

- 3.1. Centrifuge the CARTRIDGE (RT; 3,800 rcf; 1-3 min). If needed, adjust time to ensure complete flow-through.
- 3.2. Add 200 μL WASH 1 to CARTRIDGE, repeat step 3.1.
- 3.3. Add 200 μL WASH 2 to CARTRIDGE, repeat step 3.1. *SP*
- 3.4. Transfer assembled ADAPTER and CARTRIDGE onto clean COLLECTION TUBE. Discard WASTE TUBE. Label all tubes.
- 3.5. Add 100 μL ELUTE to CARTRIDGE, repeat step 3.1, keep flow-through in COLLECTION TUBE.
- 3.6. Repeat step 3.5, keep flow-through in the same COLLECTION TUBE.
- 3.7. Discard CARTRIDGE and place COLLECTION TUBE in a VACUUM EVAPORATOR (45°C; until completely dry). *SP*
- 3.8. Reconstitute peptides by adding LC-LOAD to COLLECTION TUBE. Adjust the volume according to specific requirements. For example, add 50 µL LC-LOAD to 100 µg protein starting material.
- 3.9. Sonicate COLLECTION TUBE in an ULTRASONIC BATH (5 min) or shake (RT; 500 rpm; 5 min).
- 3.10 Spin **COLLECTION TUBE** in a CENTRIFUGE (RT; 16,000 rcf; 5 min). Transfer the supernatant to a clean vial and avoid touching the bottom of the **COLLECTION TUBE** during transfer. *NOTE3*

NOTE1 Buffer volumes can be adjusted according to protein starting amounts.

Lysis temperature should be between 60-95°C.

Visit our FAQ website for more information.

NOTE2 Resuspended DIGEST can be stored for up to two weeks at 4°C. For longer storage periods, visit our FAQ.

NOTE3 At this point, peptide concentration can be measured, or the sample can be directly injected for LC-MS analysis.

Visit our FAQ for recommendations on peptide quantitation assays.

SP - Storage Point: At this point, close the peptide containing COLLECTION TUBE or CARTRIDGE using the silicone lid.

Peptides can be frozen at -20°C for two weeks. Dried peptides, prior to reconstitution in LC-LOAD, can

also be stored long-term at -80°C.

Data analysis

Consider the following as fixed modifications in your database search:

MODIFICATION	DESCRIPTION	COMPOSITION	SPECIFICITY	MASS	UNIMOD #
ALKYLATION	Carbamidomethyl on cysteine	C ₂ H ₃ NO	[C]	+57Da	4

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