PREOMICS

iST8x

Mammalian Tissue



Introduction

Sample preparation is one of the essential steps of bottom-up proteomics. The PreOmics® iST sample preparation kit is designed to streamline this process, enabling researchers to achieve optimal results with reduced steps and hands-on time. For specific recommendations and optimization, visit www.preomics.com/resources or contact info@preomics.com.

Kit Contents

The kit includes all essential components for proteomic sample preparation: denaturing, reducing, and alkylating agents, enzymes, cartridges, and wash buffers for peptide cleanup.

Component	Сар	Quantity	E	Buffer P	ropertie	S	Description	Storage
iST 8x (P.O.00001)		Organic	Acidic	Basic	Neutral			
DIGEST		2x					Trypsin/LysC mix to digest proteins.	-20°C
RESUSPEND	\bigcirc	1x 2 mL				•	For reconstituting lyophilized proteolytic enzymes.	RT
LYSE		1x 1 mL			•		For denaturing, reducing, and alkylating proteins.	RT
STOP		1x 1 mL	•	•			For stopping enzymatic activity.	RT
WASH 1		1x 2 mL	•	•			For removing hydrophobic contaminants.	RT
WASH 2		1x 2 mL		•			For removing hydrophilic contaminants.	RT
ELUTE		1x 2 mL	•		•		For eluting peptides from the cartridge.	RT
LC-LOAD	\circ	1x 1 mL		•			For loading peptides on reversed-phase LC-MS column.	RT
CARTRIDGE		8x					Cartridges with SPE sorbent for peptide purification from 1–100 µg protein starting material.	RT
WASTE TUBE		8x					2 mL tube for collecting waste after washing steps.	RT
COLLECTION TUBE		8x					1.5 mL tube for collecting peptides after elution.	RT
ADAPTER		8x					Enables a cartridge to be placed into a tube.	RT

Pre-Requisites

Common lab equipment is required for the sample preparation.

Equipment	Quantity and Description				
PIPETTE	Careful sample handling and pipetting reduces contaminations and improves quantification.				
SAMPLE	Mammalian tissue, $1-2$ mg wet weight. For most mammalian tissue types, $1-2$ mg of wet-weight tissue sample is equivalent to $100-200~\mu g$ protein starting amount. Visit our FAQ for more information on tissue starting amounts.				
REACTION TUBE	1.5 mL microcentrifuge low protein binding tubes are recommended (e.g., Eppendorf Protein LoBind® Tubes, catalogue number EU: 0030108116, US: 022431081)				
SONICATOR	For tissue homogenization using bead-based sonication (e.g., Diagenode Bioruptor®). Alternatively, use BeatBox® for tissue homogenization. Consult the BeatBox protocols at www.preomics.com/resources for details.				
GLASS BEADS	Protein extraction glass beads (Diagenode #C20000021; Ø<1 mm) to facilitate tissue lysis.				
HEATING SHAKER	Two heating shakers for tubes are recommended for protein denaturation and digestion.				
CENTRIFUGE	Benchtop centrifuge for 1.5 or 2 mL tubes is required for loading, washing, and elution.				
VACUUM EVAPORATOR	To evaporate volatile buffers from the eluate before LC-MS.				
ULTRASONIC BATH	Optional: can be used to resuspend peptides.				

Procedure



Method

1. LYSE

- 1.1. Place SAMPLE in a clean REACTION TUBE. Add 40–50 mg GLASS BEADS and 100 μL LYSE to sample. *NOTE1*
- 1.2. Shear sample in a SONICATOR (10 cycles; 30 sec ON/OFF).
- 1.3. Place sample in a pre-heated HEATING SHAKER (95°C; 1,000 rpm; 10 min). For harder tissue like heart or muscle, repeat steps 1.2 and 1.3 once.
- 1.4. Optional: Spin down droplets (RT; max. 300 rcf; 10 sec).

2. DIGEST

- 2.1. Determine protein concentration of homogenate. Visit our FAQ for recommendations on protein quantitation assays. Proceed with digestion of up to 100 μ g of extracted protein in a maximum total volume of 50 μ L (if the sample volume is less than 50 μ L, adjust to 50 μ L with LYSE).
- 2.2. Optional: Spin down lyophilized enzyme mix in the DIGEST tube (RT; max. 300 rcf; 10 sec).
- 2.3. Add 210 μL **RESUSPEND** to **DIGEST** (1 tube for 4 reactions), shake (RT; 500 rpm; 10 min) and pipette up/down. *NOTE2*
- 2.4. Add 50 μL **DIGEST** to REACTION TUBE and place it in a pre-heated HEATING SHAKER (37°C; 500 rpm; 3 h).
- 2.5. Add 100 μL STOP to REACTION TUBE (precipitation may occur), shake (RT; 500 rpm; 1 min) and pipette up/down.*SP*
- 2.6. Spin sample in CENTRIFUGE (16,000 rcf; 1 min).
- 2.7. Assemble ADAPTER and CARTRIDGE to the top of WASTE TUBE. Label all tubes and transfer supernatant to the CARTRIDGE.

3. PURIFY

- 3.1. Centrifuge the **CARTRIDGE** (RT; 3,800 rcf; 1–3 min). If needed, adjust time to ensure complete flow-through.
- 3.2. Add 200 μL WASH 1 to CARTRIDGE, repeat step 3.1.
- 3.3. Add 200 μL WASH 2 to CARTRIDGE, repeat step 3.1. *SP*
- 3.4. Transfer assembled ADAPTER and CARTRIDGE onto clean COLLECTION TUBE. Discard WASTE TUBE. Label all tubes.
- 3.5. Add 100 μL ELUTE to CARTRIDGE, repeat step 3.1, keep flow-through in COLLECTION TUBE.
- 3.6. Repeat step 3.5, keep flow-through in the same **COLLECTION TUBE**.
- 3.7. Discard CARTRIDGE and place COLLECTION TUBE in a VACUUM EVAPORATOR (45°C; until completely dry). *SP*
- 3.8. Reconstitute peptides by adding LC-LOAD to COLLECTION TUBE. Adjust the volume according to specific requirements. For example, add 50 µL LC-LOAD to 100 µg protein starting material.
- 3.9. Sonicate COLLECTION TUBE in an ULTRASONIC BATH (5 min) or shake (RT; 500 rpm; 5 min).
- 3.10. Spin **COLLECTION TUBE** in a CENTRIFUGE (RT; 16,000 rcf; 5 min). Transfer the supernatant to a clean vial and avoid touching the bottom of the **COLLECTION TUBE** during transfer. *NOTE3*

NOTE1	Alternatively, use BeatBox® for tissue homogenization and lysis. Consult the BeatBox protocols at			
	www.preomics.com/resources for details.			
NOTE 2	Resuspended DIGEST can be stored for up to two weeks at 4°C. For longer storage periods, visit our FAQ.			
NOTE 3	At this point, peptide concentration can be measured, or the sample can be directly injected for LC-MS			
	analysis. Visit our FAQ for recommendations on peptide quantitation assays.			
SP - Storage Point:	At this point, close the peptide containing COLLECTION TUBE or CARTRIDGE using the silicone lid.			
	Peptides can be frozen at -20°C for two weeks. Dried peptides, prior to reconstitution in LC-LOAD, can			
	also be stored long-term at -80°C.			

Data analysis

Consider the following as fixed modifications in your database search:

MODIFICATION	DESCRIPTION	COMPOSITION	SPECIFICITY	MASS	UNIMOD#
ALKYLATION	Carbamidomethyl on cysteine	C ₂ H ₃ NO	[C]	+57Da	4

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