# **PREOMICS**

## iST8x

#### **Plant Tissue**



#### Introduction

Sample preparation is one of the essential steps of bottom-up proteomics. The PreOmics® iST sample preparation kit is designed to assist researchers achieving best results with few sample preparation steps and little hands-on time. For specific recommendations and optimization, visit www.preomics.com/faq or contact info@preomics.com.

#### **Kit Contents**

The kit includes all essential components for proteomic sample preparation: denaturing, reducing, and alkylating agents, enzymes, cartridges, and wash buffers for peptide cleanup.

Component	Сар	Quantity	<b>Buffer Properties</b>		es	Description	Storage	
iST 8x (P.O.000	01)		Organic	Acidic	Basic	Neutral		
DIGEST		2x					Trypsin/LysC mix to digest proteins.	-20°C
RESUSPEND	$\bigcirc$	1x 2 mL				•	For reconstituting lyophilized proteolytic enzymes.	RT
LYSE		1x 1 mL			•		For denaturing, reducing, and alkylating proteins.	RT
STOP		1x 1 mL	•	•			For stopping enzymatic activity.	RT
WASH 1		1x 2 mL	•	•			For removing hydrophobic contaminants.	RT
WASH 2		1x 2 mL		•			For removing hydrophilic contaminants.	RT
ELUTE		1x 2 mL	•		•		For eluting peptides from the cartridge.	RT
LC-LOAD	$\circ$	1x 1 mL		•			For loading peptides on reversed-phase LC-MS column.	RT
CARTRIDGE		8x					Cartridges with SPE sorbent for cleaning up peptides from 1–100 $\mu g$ protein starting material.	RT
WASTE TUBE		8x					2 mL tube for collecting waste after washing steps.	RT
COLLECTION TUBE		8x					1.5 mL tube for collecting peptides after elution.	RT
ADAPTER		8x					Enables a cartridge to be placed into a tube.	RT
Additional reagents, kits and instruments from PreOmics:								
WASH 0 (P.O.00095)		10 mL	•	•			Cleans peptides from phytochemicals. Please order 1x WASH 0 buffer 10 mL (P.O.00095) from PreOmics, or ask for the buffer recipe at info@preomics.com.	RT

#### **Pre-Requisites**Common lab equipment is required for the sample preparation.

Equipment	Quantity and Description
PIPETTE	Careful sample handling and pipetting reduces sample contaminations and improves quantification.
SAMPLE	Plant tissue containing $1-100~\mu g$ protein starting material and cryomilled. Protein content strongly depends on tissue type. Visit our FAQ for more information on tissue starting amounts. Alternatively, use BeatBox® to homogenize intact plant material. Contact PreOmics for an adapted protocol.
REACTION TUBE	1.5 mL microcentrifuge low protein binding tubes are recommended (e.g., Eppendorf Protein LoBind® Tubes, catalogue number EU: 0030108116, US: 022431081).
HEATING SHAKER	Two heating shakers for tubes are recommended for protein denaturation and digestion.
CENTRIFUGE	Benchtop centrifuge for 1.5 or 2 mL tubes is required for peptide loading, washing, and elution.
SONICATOR	For tissue homogenization and lysis (e.g., Diagenode Bioruptor®).
VACUUM EVAPORATOR	To evaporate volatile buffers from the eluate before LC-MS.
ULTRASONIC BATH	Optional: can be used to resuspend peptides.

#### **Procedure**



#### Method

## 1. LYSE

- 1.1. Add 100 μL LYSE to 1–100 μg of protein sample in a REACTION TUBE, place it in a pre-heated HEATING SHAKER (95°C; 1,000 rpm; 10 min). \*NOTE1\*
- 1.2. Optional: Spin down droplets (RT; max. 300 rcf; 10 sec).
- 1.3. Shear the sample in a SONICATOR (10 cycles; 30 sec ON/OFF).
- 1.4. Optional: Spin down droplets (RT; max. 300 rcf; 10 sec).

#### 2. DIGEST

- 2.1. Optional: Spin down lyophilized enzyme mix in the DIGEST tube (RT; max. 300 rcf; 10 sec).
- 2.2. Add 210 μL **RESUSPEND** to **DIGEST** (1 tube for 4 reactions), shake (RT; 500 rpm; 10 min) and pipette up/down. \*NOTE2\*
- 2.3. Add 50 μL **DIGEST** to REACTION TUBE and place it in a pre-heated HEATING SHAKER (37°C; 500 rpm; 3 h).
- 2.4. Add 100 μL STOP to REACTION TUBE (precipitation may occur), shake (RT; 500 rpm; 1 min) and pipette up/down.\*SP\*
- 2.5. Spin sample in CENTRIFUGE (16,000 rcf; 1 min).
- 2.6. Assemble ADAPTER and CARTRIDGE to the top of WASTE TUBE. Label all tubes and transfer supernatant to the CARTRIDGE.

#### 3. PURIFY

- 3.1. Centrifuge the CARTRIDGE (RT; 3,800 rcf; 1-3 min). If needed, adjust time to ensure complete flow-through.
- 3.2. Add 200 μL WASH 0 to CARTRIDGES, repeat step 3.1.
- 3.3. Add 200 μL WASH 1 to CARTRIDGE, repeat step 3.1.

- 3.4. Add 200 μL WASH 2 to CARTRIDGE, repeat step 3.1. \*SP\*
- 3.5. Transfer assembled ADAPTER and CARTRIDGE onto clean COLLECTION TUBE. Discard WASTE TUBE. Label all tubes.
- 3.6. Add 100 μL ELUTE to CARTRIDGE, repeat step 3.1, keep flow-through in COLLECTION TUBE.
- 3.7. Repeat step 3.6, keep flow-through in the same **COLLECTION TUBE**.
- 3.8. Discard CARTRIDGE and place COLLECTION TUBE in a VACUUM EVAPORATOR (45°C; until completely dry). \*SP\*
- 3.9. Reconstitute peptides by adding LC-LOAD to COLLECTION TUBE. Adjust the volume according to specific requirements. For example, add 50 µL LC-LOAD to 100 µg protein starting material.
- 3.10. Sonicate COLLECTION TUBE in an ULTRASONIC BATH (5 min) or shake (RT; 500 rpm; 5 min).
- 3.11. Spin **COLLECTION TUBE** in a CENTRIFUGE (RT; 16,000 rcf; 5 min). Transfer the supernatant to a clean vial and avoid touching the bottom of the **COLLECTION TUBE** during transfer. \*NOTE3\*

*NOTE1*	Buffer volumes can be adjusted according to protein starting amounts.			
	Lysis temperature should be between 60-95°C. Visit our FAQ website for more information.			
*NOTE 2*	Resuspended DIGEST can be stored for up to two weeks at 4°C. For longer storage periods, visit our FAQ.			
*NOTE 3*	At this point, peptide concentration can be measured, or the sample can be directly injected for LC-MS			
	analysis. Visit our FAQ for recommendations on peptide quantitation assays.			
*SP* - Storage Point:	At this point, close the peptide containing <b>COLLECTION TUBE</b> or <b>CARTRIDGE</b> using the silicone lid.			
	Peptides can be frozen at -20°C for two weeks. Dried peptides, prior to reconstitution in <b>LC-LOAD</b> , can			
	also be stored long-term at -80°C.			

## Data analysis

Consider the following as fixed modifications in your database search:

MODIFICATION	DESCRIPTION	COMPOSITION	SPECIFICITY	MASS	UNIMOD#
ALKYLATION	Carbamidomethyl on cysteine	C <sub>2</sub> H <sub>3</sub> NO	[C]	+57Da	4

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