



## Introduction

Sample preparation is one of the essential steps of bottom-up proteomics. The PreOmics® iST-S sample preparation kit is designed to streamline this process for low amounts of starting material, enabling researchers to achieve optimal results with reduced steps and hands-on time. For specific recommendations and optimization, visit [www.preomics.com/resources](http://www.preomics.com/resources) or contact [info@preomics.com](mailto:info@preomics.com).

## Kit Contents

The kit includes all essential components for proteomic sample preparation: denaturing, reducing, and alkylating agents, enzymes, cartridges, and wash buffers for peptide cleanup.

Component	Cap	Quantity	Buffer Properties				Description	Storage
			Organic	Acidic	Basic	Neutral		
DIGEST-L	●	1x					Trypsin/LysC mix to digest proteins.	-20°C
RESUSPEND	●	1x 1 mL				●	For reconstituting lyophilized proteolytic enzymes.	RT
LYSE A	●	1x 1 mL				●	For denaturing, reducing, and alkylating proteins.	RT
LYSE B	●	1x 1 mL				●	For denaturing, reducing, and alkylating proteins.	RT
STOP	●	4x 1 mL	●	●			For stopping enzymatic activity.	RT
WASH 1	●	2x 1 mL	●	●			For removing hydrophobic contaminants.	RT
WASH 2	●	2x 1 mL		●			For removing hydrophilic contaminants.	RT
ELUTE	●	2x 1.4 mL	●		●		For eluting peptides from the cartridge.	RT
LC-LOAD	○	1x 1.4 mL		●			For loading peptides on reversed-phase LC-MS column.	RT
CARTRIDGE		12x					Cartridges with SPE sorbent for peptide purification sealed with silicone mat.	RT
WASTE TUBE		12x					2 mL tube for collecting waste after washing steps.	RT
COLLECTION TUBE		12x					1.5 mL tube for collecting peptides after elution.	RT
ADAPTER		12x					Enables a cartridge to be placed into a tube. It can be used for further experiments.	RT

## Pre-Requisites

Common lab equipment is required for the sample preparation.

Equipment	Quantity and Description
PIPETTE	Careful sample handling and pipetting reduce contamination and improve quantification.
REACTION PLATE	96-well micro plates are recommended (e.g., Eppendorf LoBind® twin.tec® PCR Plates, catalogue number 0030129.512).
SEALING MAT	Prevents sample contamination and evaporation (e.g., AXYGEN Sealing Mat, product number: AM-96-PCR-RD).
SAMPLE	Pelleted & adherent cells (equivalent to 1–10 µg total protein).
HEATING SHAKER	Two heating shakers for multi-well plates are recommended to support protein denaturation and digestion.
CENTRIFUGE	Benchtop centrifuges for plates and 1.5/2 mL standard tubes are required.
VACUUM EVAPORATOR	To evaporate volatile buffers from the eluate before LC-MS.

## Procedure



## Method

### 1. LYSE

- 1.1. The **LYSE** volume needed per sample is 40  $\mu\text{L}$ . Calculate the total volume of **LYSE** required for your experiment. Prepare the calculated **LYSE** volume by mixing equal volumes of **LYSE-A** (yellow circle) and **LYSE-B** (brown circle) in a suitable vessel. Briefly vortex the **LYSE** mixture.
- 1.2. Add 40  $\mu\text{L}$  **LYSE** to 1–10  $\mu\text{g}$  of protein sample.
- 1.3. Place **SEALING MAT** on the **REACTION PLATE** and place it on a pre-heated **HEATING SHAKER** (80°C; 1,000 rpm; 10min).
- 1.4. Optional: Quick spin to remove condensation from the lid (RT; max. 300 rcf; 10 sec).
- 1.5. Allow samples to cool down to room temperature.

### 2. DIGEST

- 2.1. Optional: Spin down lyophilized enzyme mix in the **DIGEST-L** tube (RT; max. 300 rcf; 10 sec).
- 2.2. Add 300  $\mu\text{L}$  **RESUSPEND** (yellow circle) to **DIGEST-L** (red circle) (1 tube for 24 reactions) and shake (RT; 500 rpm; 10 min).
- 2.3. Add 10  $\mu\text{L}$  **DIGEST-L** (red circle) to each well.
- 2.4. Place **SEALING MAT** on the **REACTION PLATE**. Place **REACTION PLATE** on a pre-heated **HEATING SHAKER** (37°C; 1000 rpm; 1 h).
- 2.5. Optional: Spin to remove condensation from the lid (RT; max. 300 rcf; 10 sec).
- 2.6. Add 60  $\mu\text{L}$  **STOP** (black circle) to each **REACTION VESSEL**, shake (RT; 300 rpm; 1 min).
- 2.7. Place **CARTRIDGES** in **WASTE TUBES** by using **ADAPTERS**.

### 3. PURIFY

- 3.1. Optional: Priming before transferring the samples to the **CARTRIDGES** by adding 200  $\mu\text{L}$  **STOP**. Place **CARTRIDGES** in a centrifuge and spin (RT; 1000 rcf; 1 min). Make sure the **CARTRIDGES** do not dry out afterwards and directly continue with step 3.2. **\*NOTE1\***
- 3.2. Transfer the samples to the **CARTRIDGES**.
- 3.3. Place **CARTRIDGES** in a **CENTRIFUGE** and spin (1,000 rcf; 1 min). If needed, adjust time to ensure complete flow-through.
- 3.4. Add 100  $\mu\text{L}$  **WASH 1** (blue circle) to the **CARTRIDGES** and centrifuge (2,250 rcf; 1 min).
- 3.5. Add 100  $\mu\text{L}$  **WASH 2** (green circle) to the **CARTRIDGES** and centrifuge (2,250 rcf; 1 min).
- 3.6. Discard **WASTE TUBES**. Place **CARTRIDGES** in **COLLECTION TUBES** using **ADAPTERS**.
- 3.7. Add 75  $\mu\text{L}$  **ELUTE** (pink circle) to the **CARTRIDGES**. Place **CARTRIDGES** in a **CENTRIFUGE** and spin (1,000 rcf; 1 min). Keep flow-through in the **COLLECTION TUBES**.
- 3.8. Add further 75  $\mu\text{L}$  **ELUTE** (pink circle) to the **CARTRIDGES** and centrifuge (1,000 rcf; 1 min). Keep flow-through in the same **COLLECTION TUBES**.
- 3.9. Discard **CARTRIDGES** and place **COLLECTION TUBES** in a **VACUUM EVAPORATOR** (RT; until completely dry). **\*SP\***
- 3.10. Reconstitute peptides by adding **LC-LOAD** (white circle). **\*NOTE2\***
- 3.11. Adjust the peptide concentration according to the **LC-MS** column loading requirements. **\*NOTE3\***  
It is recommended to centrifuge the samples prior to **LC-MS** analysis (RT; 2,250 rcf; 10 min).

- \*NOTE1\* Priming may increase protein identifications, especially for protein input amounts below 5 µg.
- \*NOTE2\* Using 20 µL of LC-LOAD will typically result in a peptide concentration of 0.2 µg/µL for 5 µg and 0.3 µg/µL for 10 µg sample input.
- \*NOTE3\* For accurate adjustment of peptide injection amount, a peptide quantification should be performed. It is recommended using a quantitative fluorometric assay (e.g., Pierce Quantitative Fluorometric Peptide Assay Kit, Thermo Scientific™, catalogue number 23290), with an input of 2 µL peptide sample + 8 µL LC-LOAD.
- \*SP\* - Storage Point: Dried peptides, prior to reconstitution in LC-LOAD, can be stored long-term at -80°C.

## Data analysis

Consider the following as fixed modifications in your database search:

MODIFICATION	DESCRIPTION	COMPOSITION	SPECIFICITY	MASS	UNIMOD #
ALKYLATION	Carbamidomethyl on cysteine	C <sub>2</sub> H <sub>3</sub> NO	[C]	+57DA	4

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