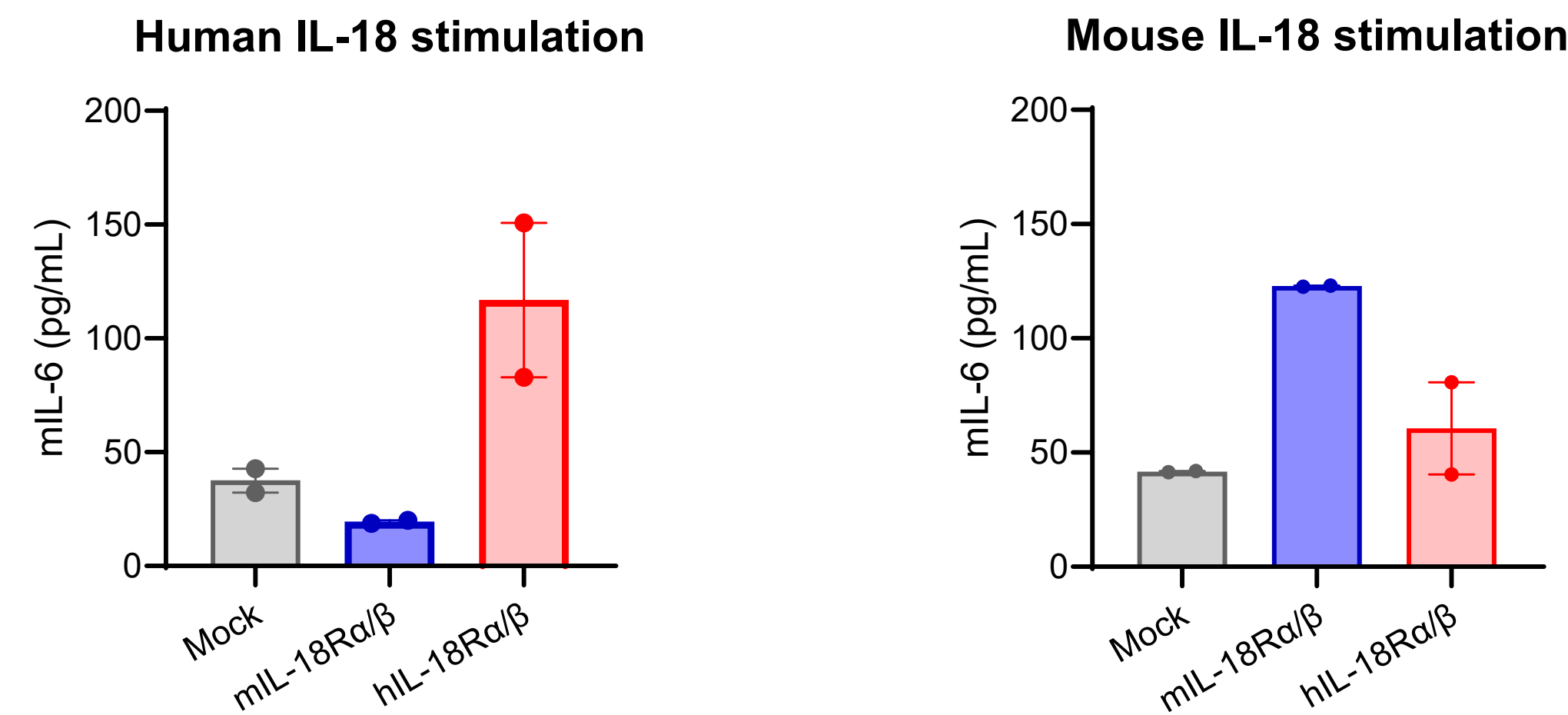


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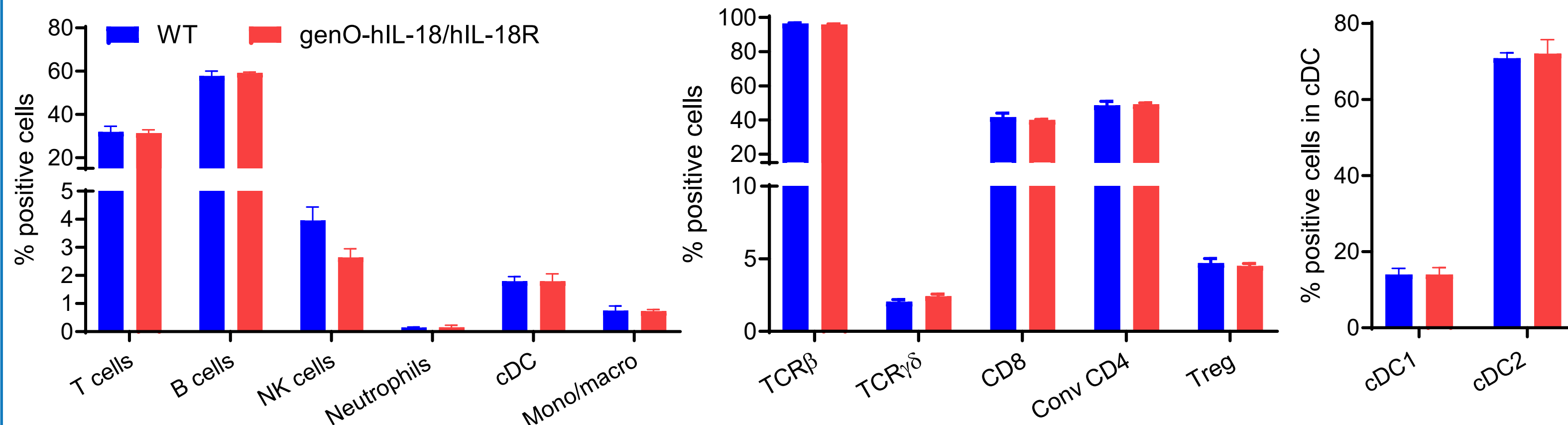
**Background:** Interleukin-18 (IL-18) has emerged as a promising immunomodulatory cytokine in immuno-oncology due to its ability to enhance both innate and adaptive immune responses. It promotes IFN- $\gamma$  production by T and NK cells, thereby amplifying anti-tumor immunity, especially when combined with checkpoint inhibitors or engineered pro-drugs to resist natural inhibition by IL-18BP. Recent studies suggest that IL-18-based therapies may overcome resistance mechanism in “cold” tumors, making them responsive to immunotherapies<sup>1</sup>. Humanized mouse models expressing human IL-18R are essential to accurately assess efficacy and guide development of human-directed IL-18-based therapies. Therefore, we describe here a new IL-18/IL-18R double humanized mouse model to assess the efficacy of therapeutics targeting the IL-18/IL-18R axis.

## 1. Mouse IL-18 partially induces signaling through human IL-18R $\alpha/\beta$ , requiring humanization of both IL-18 and IL-18R $\alpha/\beta$ to maintain functional IL-18/IL-18R axis



B16F10 cells stably expressing mouse or human IL-18R $\alpha/\beta$  were stimulated with recombinant mouse or human IL-18 (10 ng/mL) for 24 hours. Mouse IL-6 secretion in the supernatant was then assessed by ELISA.

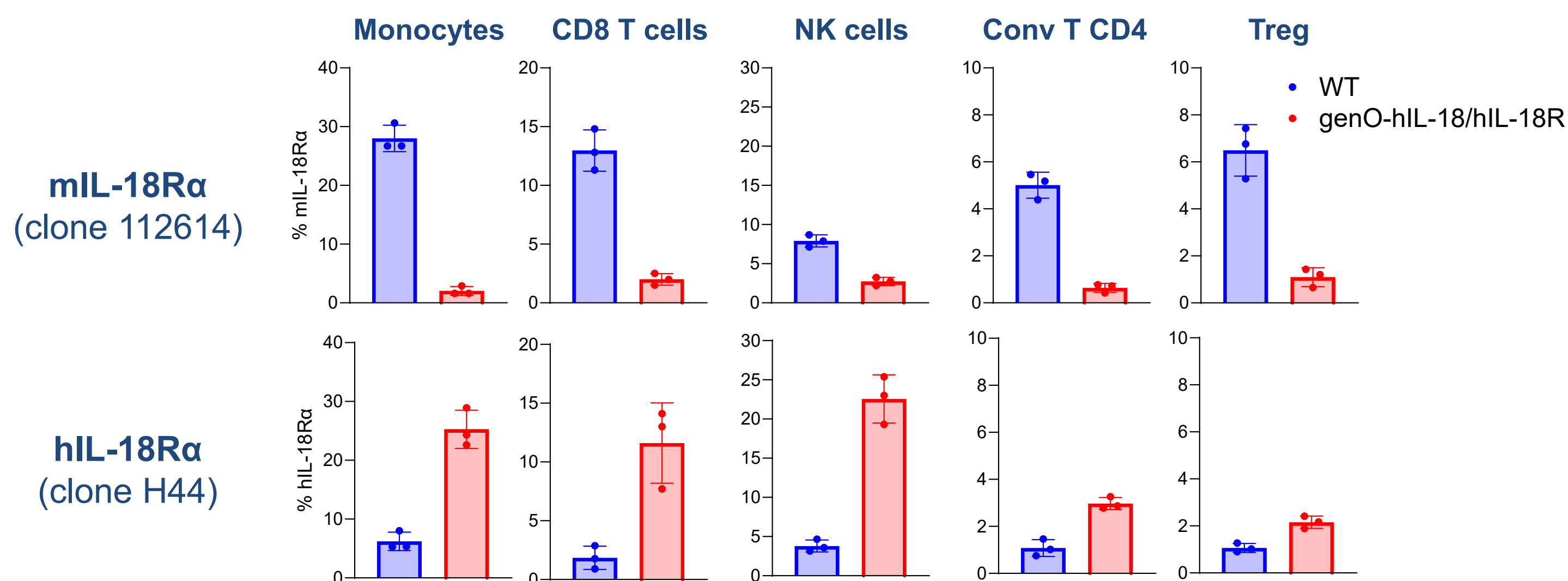
## 2. Humanization of both IL-18 and IL-18R does not alter immune cell distribution



Freshly isolated splenocytes, gated on live CD3<sup>+</sup> CD19<sup>-</sup> (T cells) and CD3<sup>-</sup> CD19<sup>+</sup> (B cells), in CD3<sup>-</sup> CD19<sup>-</sup> cells: NKp46<sup>+</sup> (NK cells), CD317<sup>+</sup> (pDC), Ly-6G<sup>+</sup> (Neutrophils), Ly-6G<sup>-</sup> CD11c<sup>+</sup> MHCII<sup>+</sup> (cDC) and Ly-6G<sup>-</sup> Ly-6C<sup>+</sup> CD11b<sup>+</sup> (Monocytes/macrophages). T cell subsets gated on CD3<sup>+</sup> TCR $\beta$ <sup>+</sup> TCR $\gamma\delta$ <sup>-</sup>: CD4<sup>+</sup> CD8<sup>+</sup> (CD8), CD4<sup>+</sup> CD25<sup>+</sup> FoxP3<sup>+</sup> (convCD4) and CD4<sup>+</sup> CD25<sup>+</sup> FoxP3<sup>-</sup> (Treg). In cDC: CD8<sup>+</sup> CD11b<sup>-</sup> (cDC1) and CD8<sup>-</sup> CD11b<sup>+</sup> (cDC2).

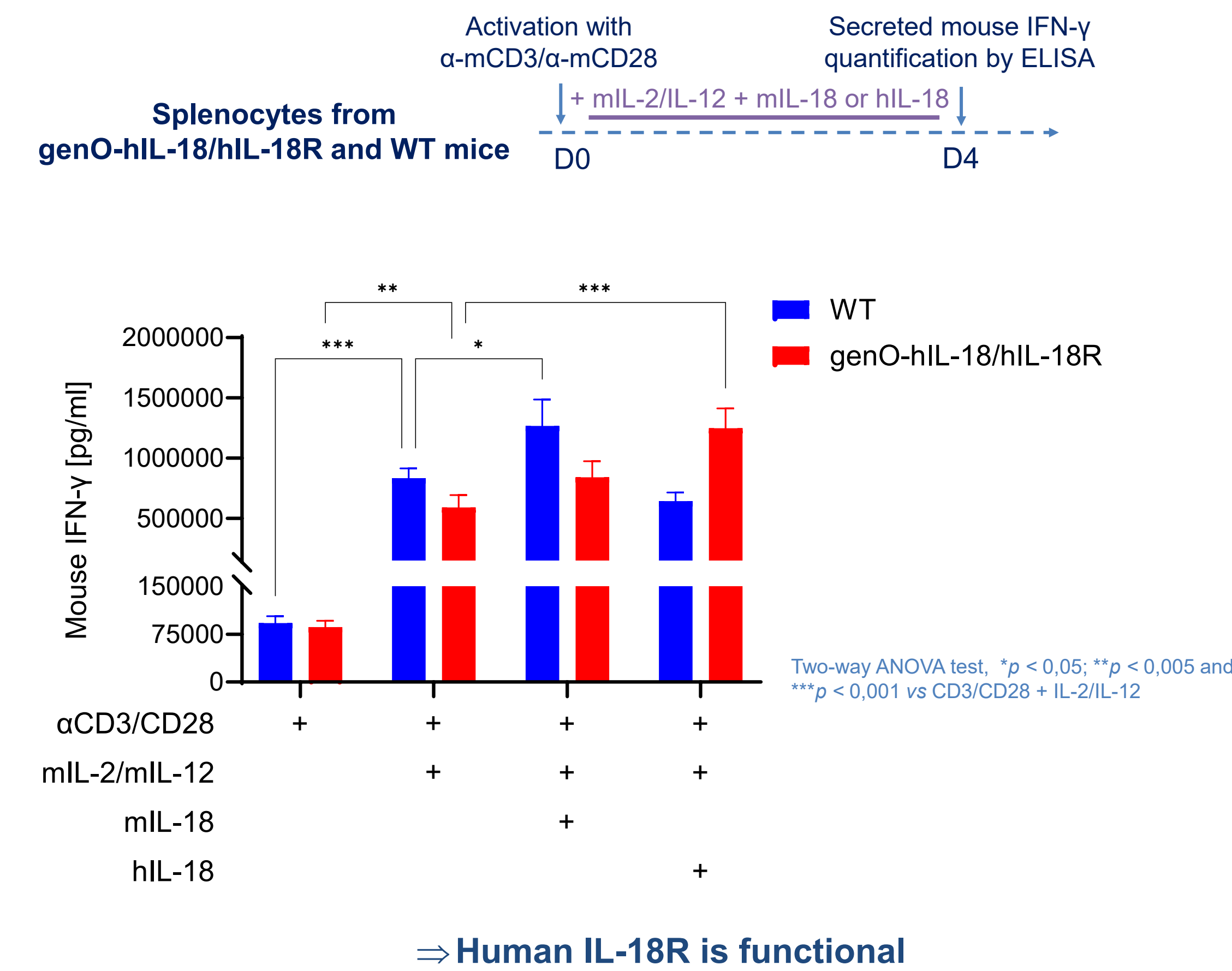
⇒ Similar results of immune profiling in blood and bone marrow – data not shown

## 3. Human IL-18R $\alpha$ expression on immune cells from genO-hIL-18/hIL-18R mice



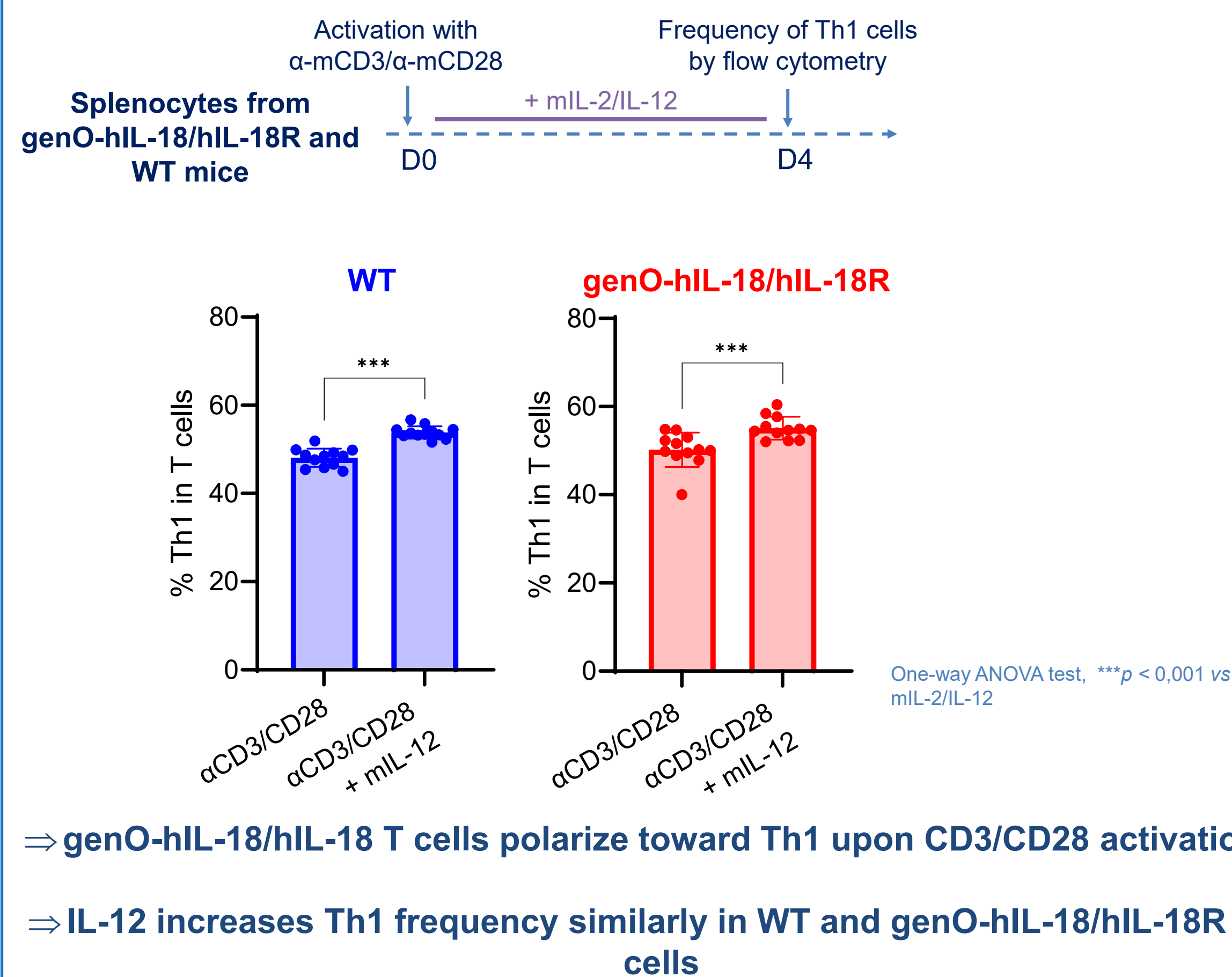
Freshly isolated splenocytes, gated on single live CD3<sup>+</sup> CD19<sup>-</sup> NKp46<sup>-</sup> CD317<sup>-</sup> Ly-6G<sup>-</sup> Ly-6C<sup>+</sup> Ly-6C<sup>+</sup> CD11b<sup>+</sup> (Monocytes/macrophages), CD3<sup>+</sup> CD19<sup>-</sup> NKp46<sup>+</sup> (NK cells), CD3<sup>+</sup> TCR $\beta$ <sup>+</sup> TCR $\gamma\delta$ <sup>-</sup> CD4<sup>+</sup> CD8<sup>+</sup> (CD8 T cells), CD3<sup>+</sup> TCR $\beta$ <sup>+</sup> TCR $\gamma\delta$ <sup>-</sup> CD4<sup>+</sup> CD25<sup>+</sup> FoxP3<sup>+</sup> (convCD4), CD3<sup>+</sup> TCR $\beta$ <sup>+</sup> TCR $\gamma\delta$ <sup>-</sup> CD4<sup>+</sup> CD25<sup>+</sup> FoxP3<sup>-</sup> (Treg).

## 4. Human IL-18 induces IFN- $\gamma$ secretion in splenocytes from genO-hIL-18/hIL-18R mice



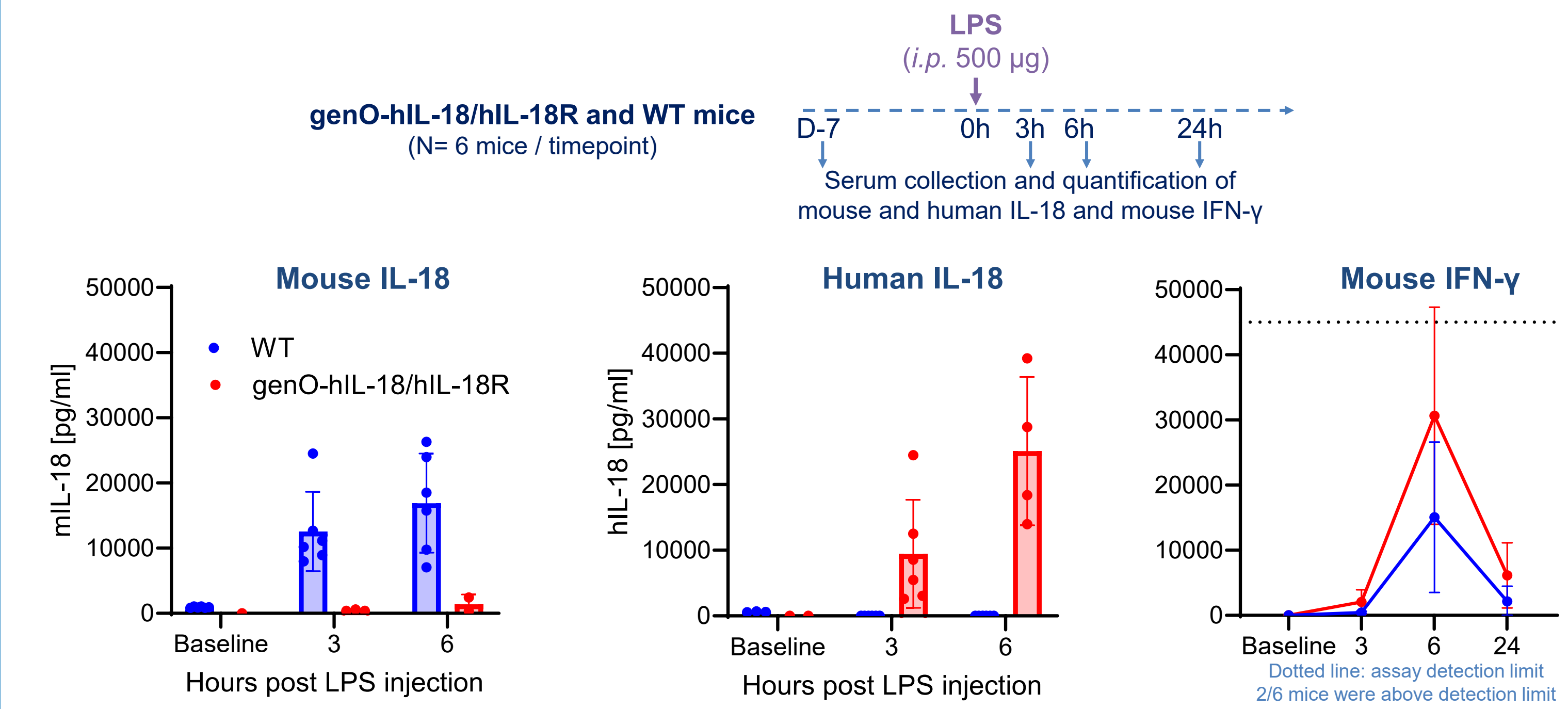
⇒ Human IL-18R is functional

## 5. Th1 differentiation is unaltered in genO-hIL-18/hIL-18R mice



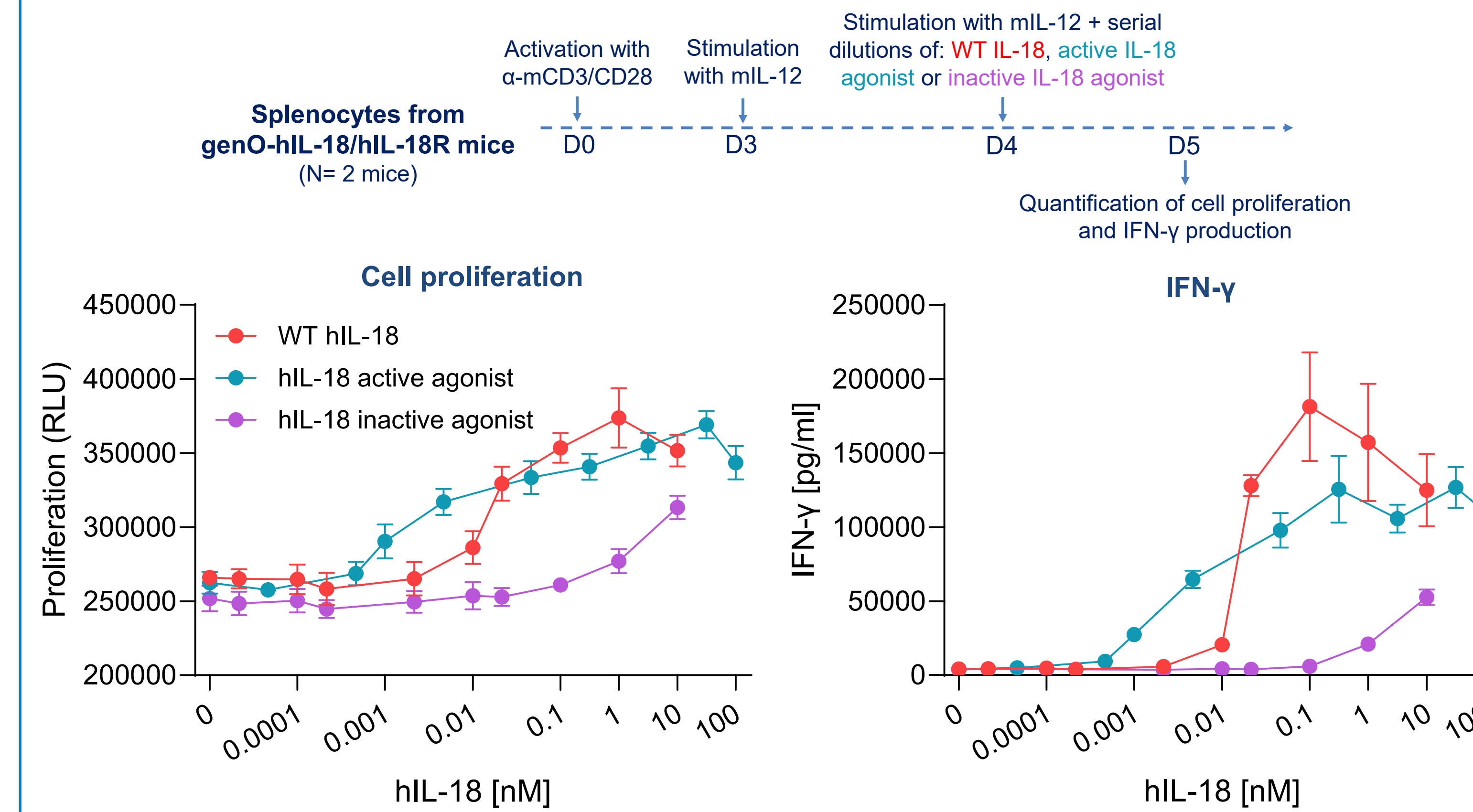
⇒ genO-hIL-18/hIL-18 T cells polarize toward Th1 upon CD3/CD28 activation  
⇒ IL-12 increases Th1 frequency similarly in WT and genO-hIL-18/hIL-18R T cells

## 6. In vivo LPS-induced IL-18 and IFN- $\gamma$ release



⇒ LPS-induced human IL-18 secretion in genO-hIL-18/hIL-18R mice mirrors mouse IL-18 secretion in WT mice  
⇒ IFN- $\gamma$  is secreted in a similar kinetic in genO-hIL-18/hIL-18R and WT mice  
⇒ mL-18BP expression is increased by LPS similarly in genO-hIL-18/hIL-18R and WT mice (data not shown)

## 7. Ex vivo efficacy of IL-18 agonists



⇒ Splenocytes from genO-hIL-18/hIL-18R mice treated with engineered hIL-18 exhibit enhanced cell proliferation and increased IFN- $\gamma$  secretion compared to inactive hIL-18

**Conclusion:** Altogether, these results support the suitability of the genO-hIL-18/hIL-18R mouse model for assessment of new therapies targeting this axis.

**Reference:** 1. Kessel C, Rossig C, Abken H. Weal and woe of interleukin-18 in the T cell therapy of cancer. *Journal for ImmunoTherapy of Cancer*. 2025;13:e010545. <https://doi.org/10.1136/jitc-2024-010545>

