



Humanized FcγR/FcRn mouse model exhibits immunological responses comparable to wild-type mice

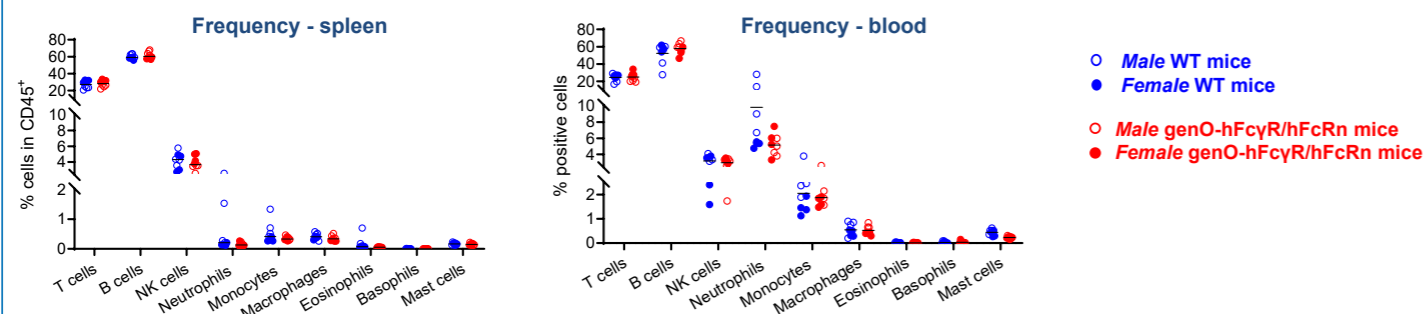
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Abstract #3803

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Background: Therapeutic antibodies have revolutionized cancer treatment by leveraging immune mechanisms such as Fc-effector functions, which depend on interactions between IgG and Fcγ receptors (FcγR). However, preclinical evaluation of antibody pharmacokinetics (PK) and pharmacodynamics (PD) remains challenging due to species-specific differences in FcγR and FcRn expression and function. We previously reported a FcγR humanized model that expresses a human-like pattern of FcγRs (including FcγRI, FcγRIIA, FcγRIIB, FcγRIIIA and FcγRIIIB – genO-hFcγR). These receptors are functional and enable accurate evaluation of Fc-effector functions such as ADCC and B-cell depletion. The model demonstrated Fc-dependent activity of therapeutic IgG, allowing differentiation between antibodies with regular versus enhanced FcγR binding and supports ranking of Fc-engineered antibodies in preclinical studies¹. FcRn was also humanized in the model to enhance the translatability of PK studies by enabling human FcRn-mediated IgG recycling, while maintaining PD assessment of Fc-engineered therapeutic antibodies. Herein, we characterized the immunological competence of the humanized genO-hFcγR/FcRn mouse model under inflammatory conditions by comparing its immune responses with those of wild-type (WT) mice following defined immunological challenges.

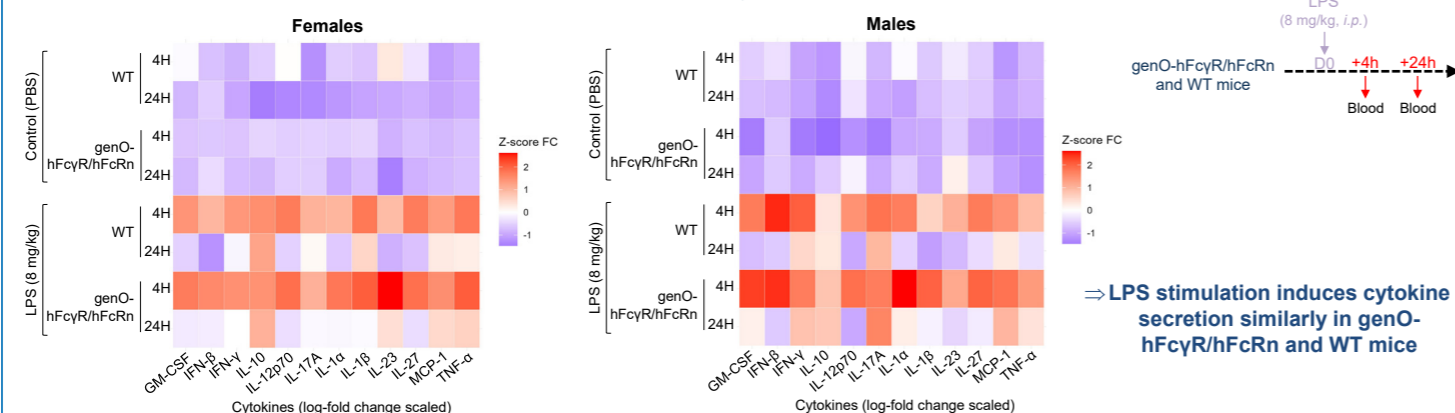
1. Humanization of Fcγ receptors and FcRn does not alter immune cell distribution in genO-hFcγR/hFcRn compared to WT mice in spleen and blood



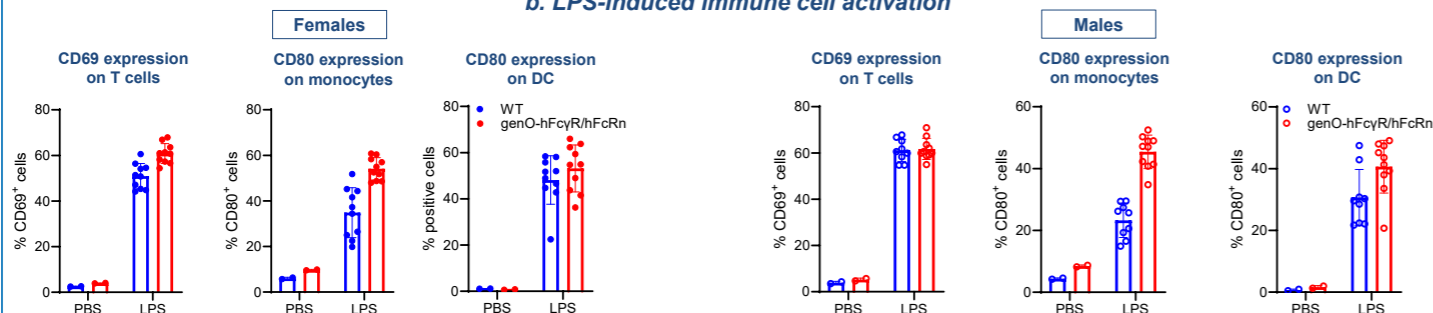
Freshly isolated splenocytes, gated on live CD45⁺ CD200R⁺ FcεRI⁺ (Basophils), CD45^{low} CD200R⁺ FcεRI⁺ (Mast cells); in CD45⁺ CD200R⁻ CD3⁺ CD19⁻ (T cells) and CD3⁺ CD19⁺ (B cells); in CD3⁺ CD19⁻ cells: NKp46⁺ (NK cells), Ly-6G⁺ (Neutrophils), Ly-6G⁺ SIGLEC-F⁺ (Eosinophils), Ly-6G⁺ SIGLEC-F⁻ Ly-6C⁺: F4/80⁻ (Monocytes) and F4/80⁺ (Macrophages).

⇒ Similar results of immune profiling in bone marrow – data not shown

2. In vivo response to LPS is similar in genO-hFcγR/hFcRn and WT mice



b. LPS-induced immune cell activation

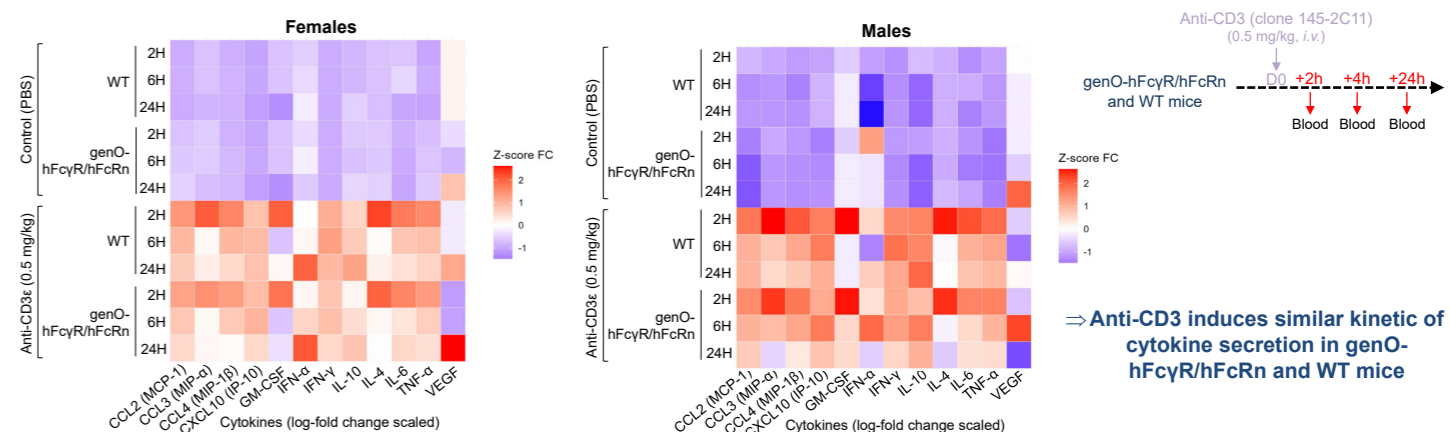


Freshly isolated splenocytes, T cells (gated on live CD3⁺CD19⁻ TCRαβ⁺) and CD3⁺CD19⁻NKp46⁻CD11b⁺ (monocytes) and CD3⁺CD19⁻NKp46⁻CD11c⁺ (DC cells) 24h post LPS injection (8 mg/kg, i.p.).

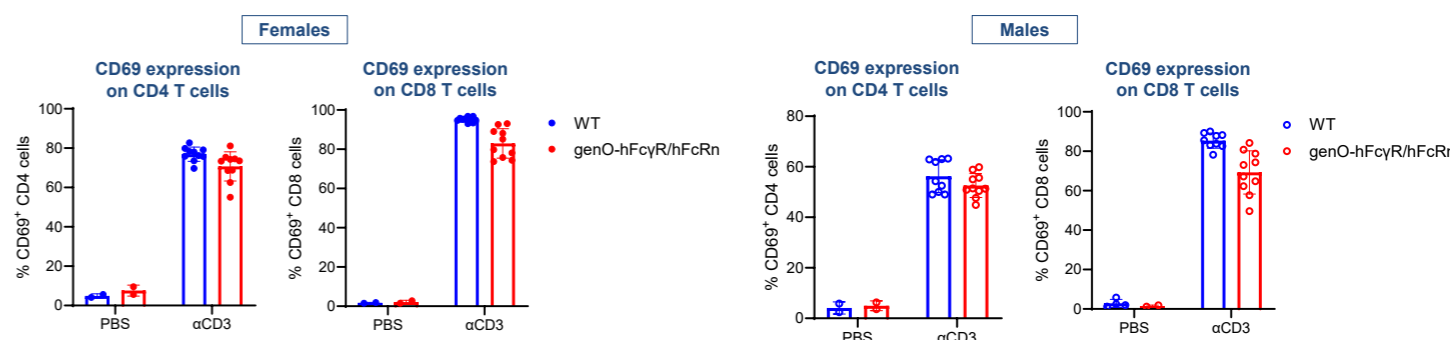
⇒ LPS induces expression of CD69 on T cells and CD80 on monocytes and DC in both genO-hFcγR/hFcRn and WT mice, suggesting immune cells activation, which is in line with the increased cytokine production

3. In vivo response to anti-mouse CD3 is similar in genO-hFcγR/hFcRn and WT mice

a. Anti-mouse CD3-induced cytokine secretion



b. Anti-mouse CD3-induced immune cell activation



Freshly isolated splenocytes, T cells (gated on live CD3⁺CD19⁻ TCRαβ⁺), CD4 (CD4⁺ CD8⁻) and CD8 (CD4⁻ CD8⁺) cells, 24h post anti-CD3 injection (clone 145-2C11, 0.5 mg/kg, i.v.).

⇒ Overall similar induction of CD69 expression on CD4 and CD8 T cells from genO-hFcγR/hFcRn mice compared to WT mice, in line with cytokine production induced by anti-CD3

Conclusion: These results demonstrate that genO-hFcγR/FcRn mice are immunocompetent and mount immune responses comparable to those of wild-type mice. This model is a robust and reliable tool for studying Fc-engineered therapeutic antibodies in a context that preserves native immune functionality. The genO-hFcγR/hFcRn model is also being improved to enable tolerability to hIgG1 antibodies, and flexibility of therapeutics testing through expression of human immune checkpoints.

Reference: 1. Van Damme KFA, Sichien D, Van der Borcht K, et al. Cross-species cellular mapping and humanization of Fcγ receptors to advance antibody modeling. *Sci Immunol.* 2026;11(115):eady7328. doi:10.1126/sciimmunol.ady7328

