



Novel FcγR humanized mouse models enabling efficacy and PK/PD of therapeutic antibodies

Fabiane SÔNEGO¹, Angela PAPPALARDO¹, Gaele MARTIN¹, Rachel COURTOIS², Séverine AUGIER², Julie GORRY¹, Yacine CHERIF¹, Ariane MOREL², Patricia ISNARD-PETIT¹, Kader THIAM¹

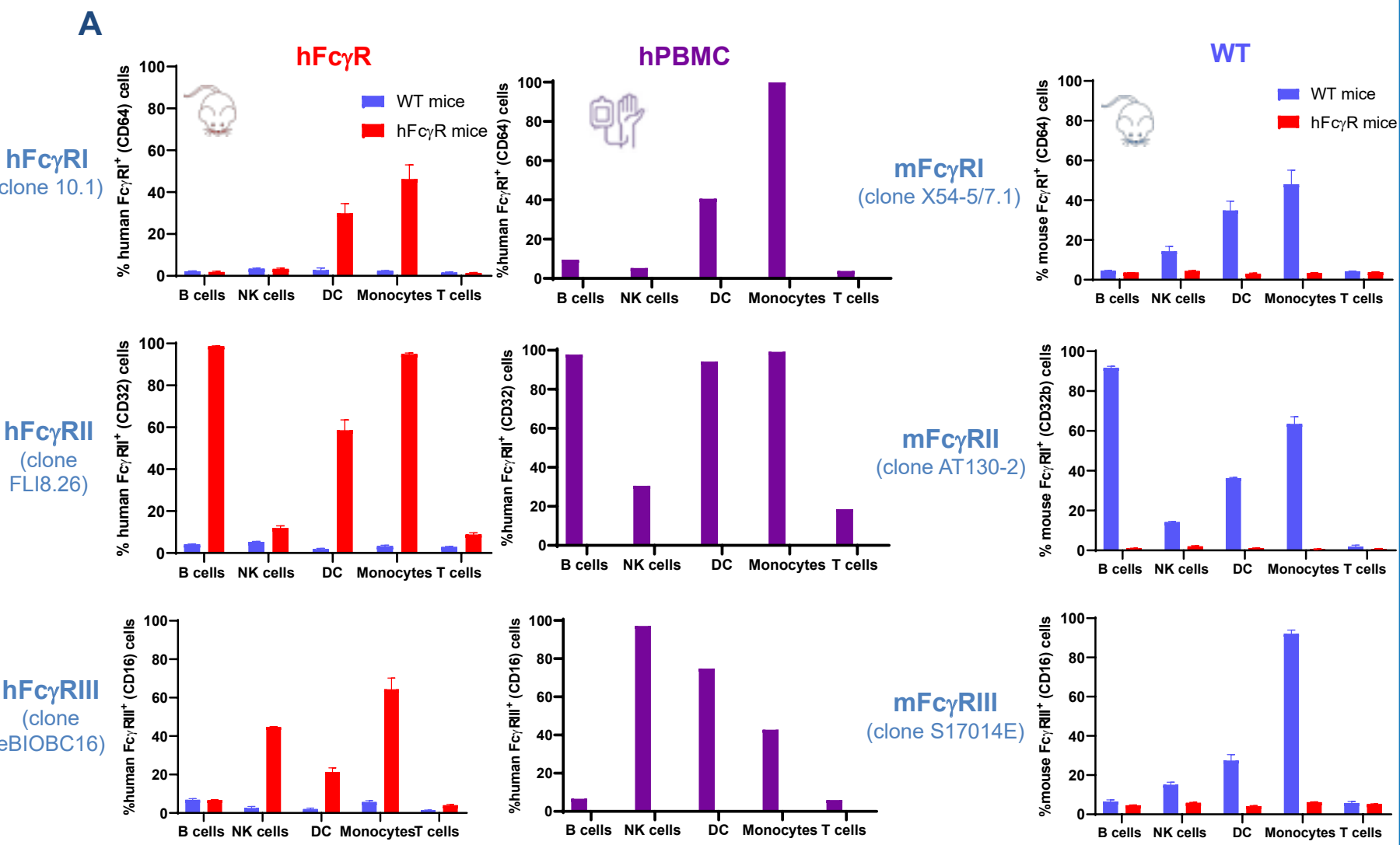
¹genOway, Lyon, France
²Innate Pharma, Marseille, France

AACR 2024
Abstract #5341

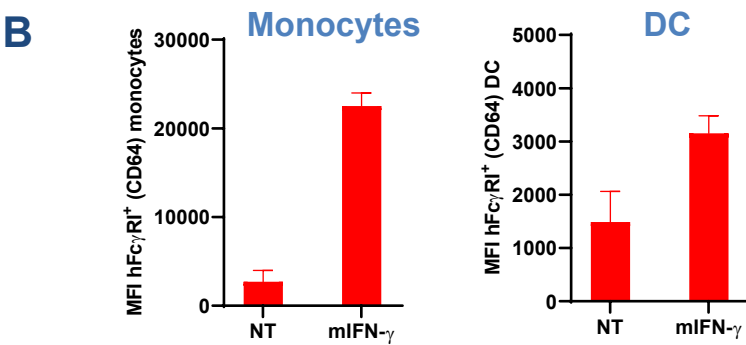
Background: Therapeutic antibodies have revolutionized treatment of cancer. Enhanced activity of therapeutic IgG can be achieved by the modulation of Fc binding to Fcγ receptors (FcγR) which will modulate the Fc-effector function triggered upon crosslinking target effector cells by the therapeutic antibodies.

We report here a novel mouse model expressing humanized FcRI, FcγRIIA, FcγRIIB, FcγRIIIA and FcγRIIIB, instead of the mouse versions. This model was intercrossed with the previously described hSA/hFcRn mouse model (Viuff et al., 2016), which has shown to improve translatability of PK assessment of therapeutics with extended half-life via FcRn recycling and/or hSA binding.

1. Expression of FcγRs



⇒ hFcγR expression pattern in hFcγR mice is consistent with the expression in human PBMC

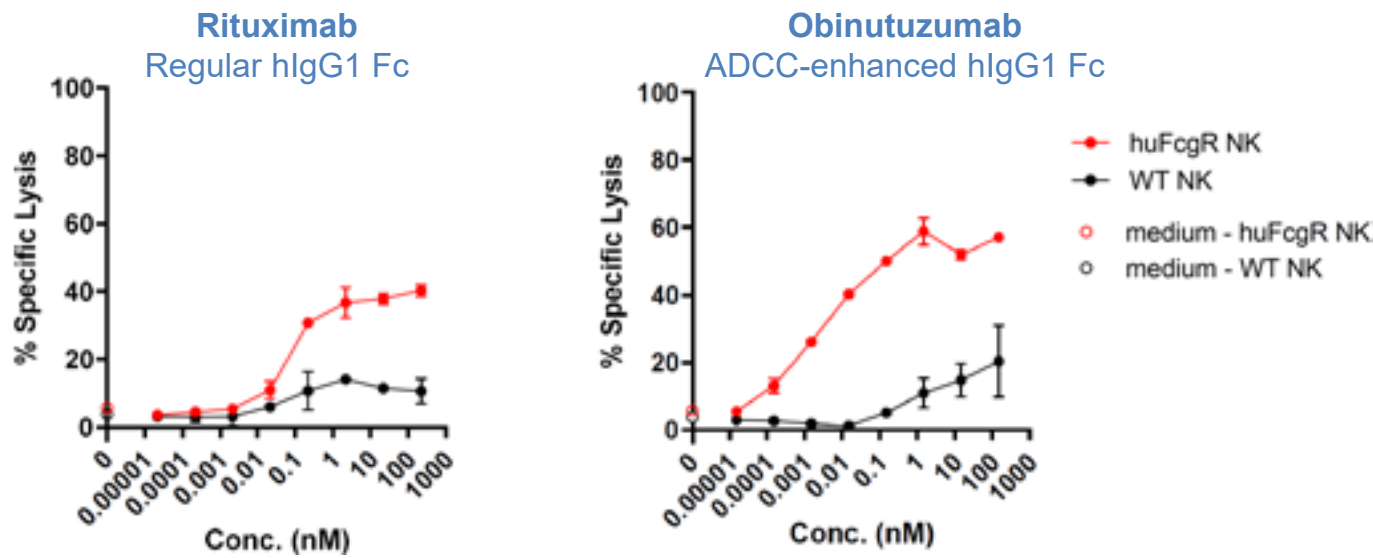


⇒ Expression of hFcγR is increased upon treatment with IFN-γ on monocytes and DC as described in human cells (Bournazos et al., Microbiol Spectr., 2016) – Regulation of FcγRI expression is kept

2. Functionality of hFcγR

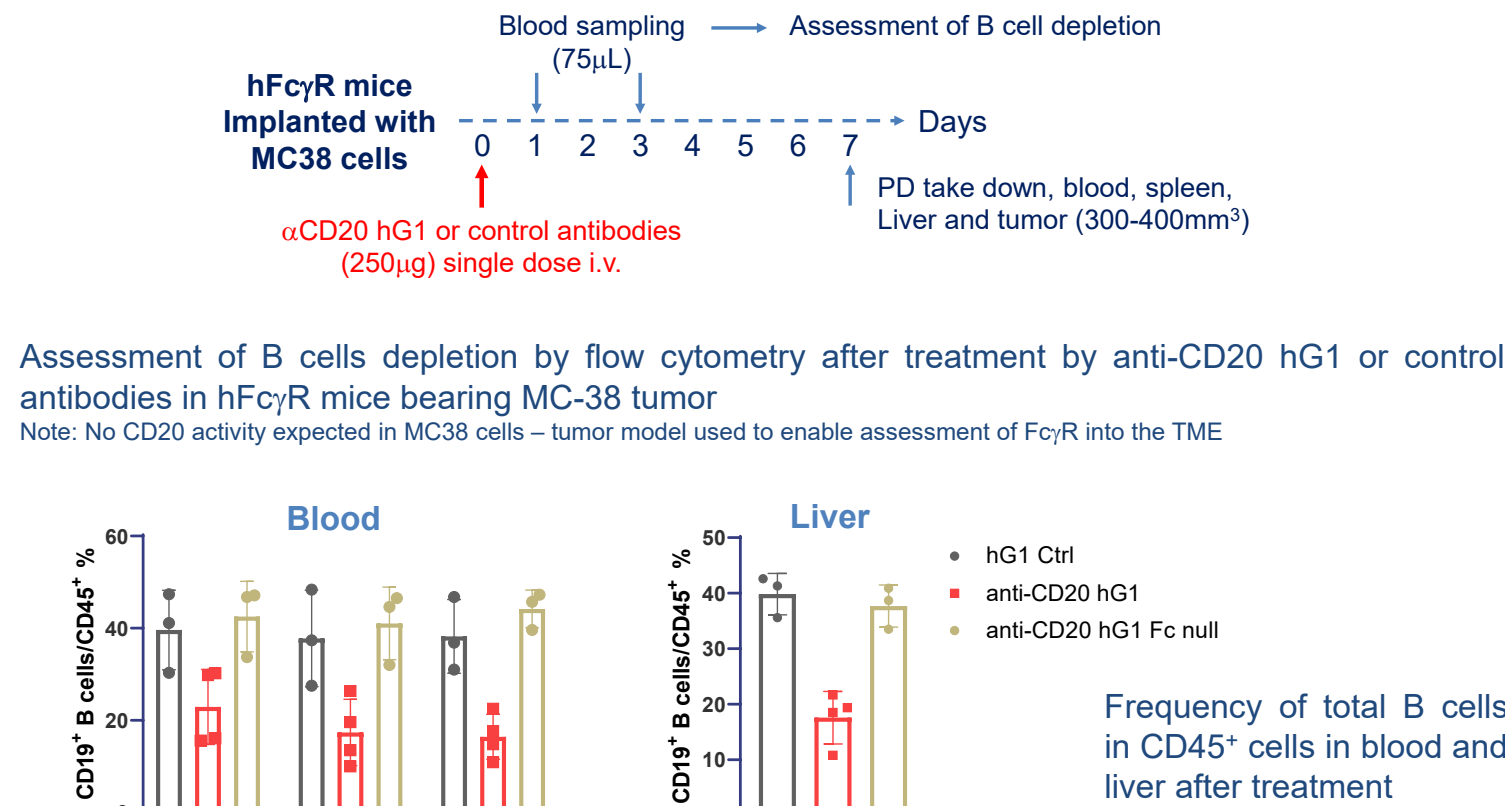
2.1. Ex vivo ADCC by NK cells

- NK cells isolated from the spleen of WT or hFcγR mice loaded with B16F-10-CD20 cells loaded with ⁵¹Cr (ratio 10:1) in the presence of Rituximab or Obinutuzumab
- Antibody-dependent cell cytotoxicity (ADCC) was measured by gamma radiation count in the supernatant



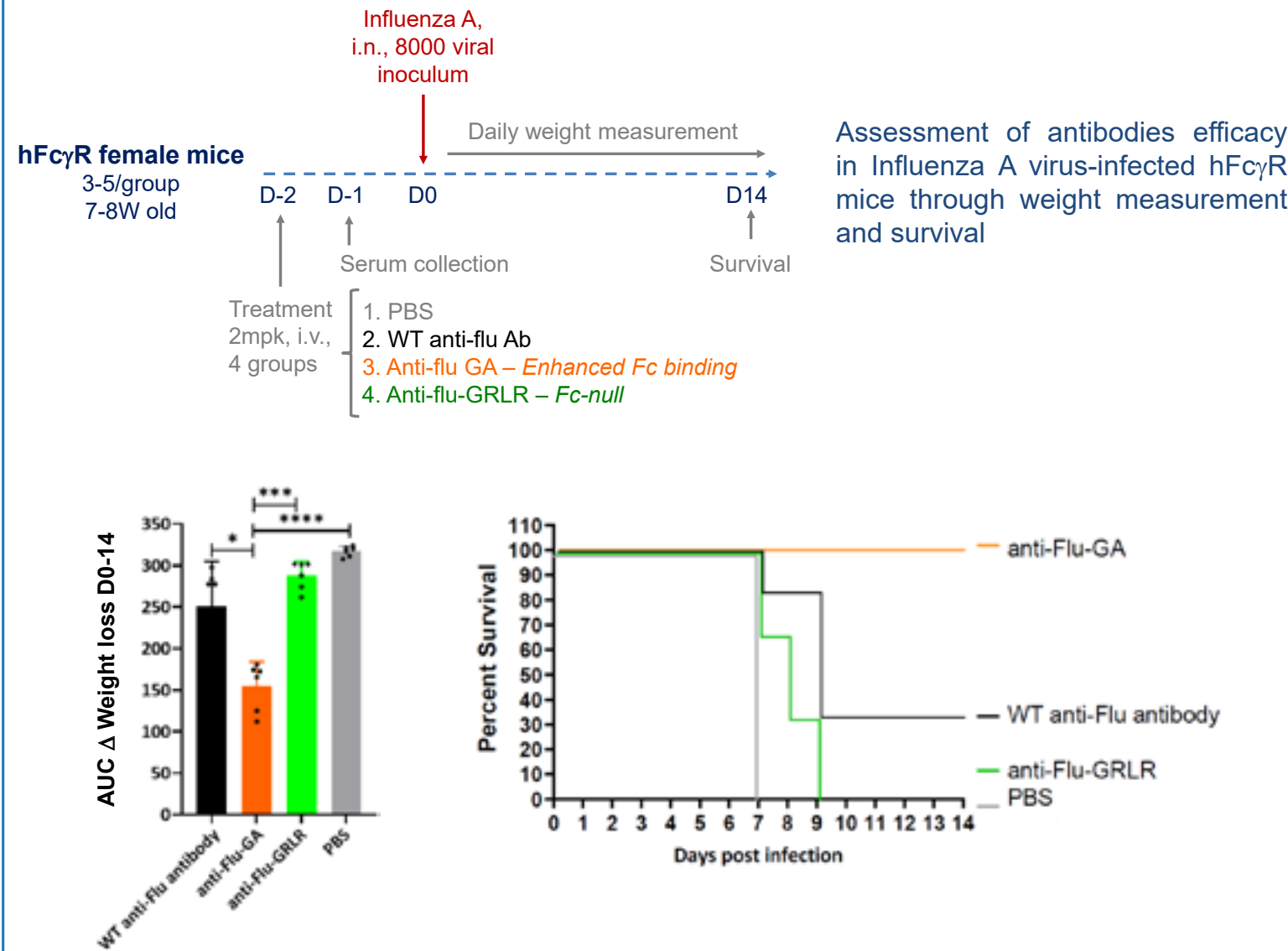
⇒ Rituximab induces higher ADCC in NK cells from hFcγR mice compared to WT mice
⇒ Obinutuzumab, featuring an engineered Fc portion to enhance binding to FcγRIII (Mössner et al., Blood, 2010), induces higher lysis than Rituximab (regular Fc) in NK cells from hFcγR mice

2.2. In vivo B cells depletion in hFcγR mice



⇒ Anti-CD20 depletes approximately 50% of B cells in blood and liver from hFcγR mice
⇒ B cell depletion is dependent on Fc effector function (no activity in Fc-null anti-CD20 human IgG1)

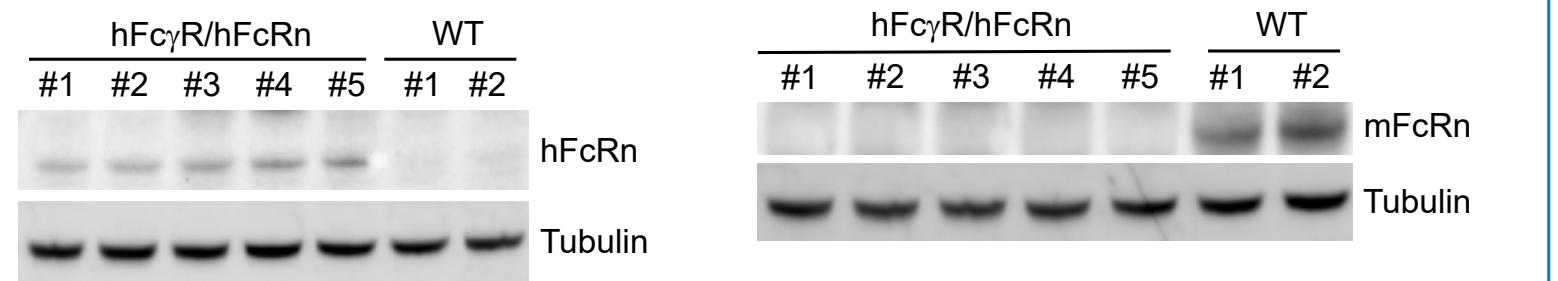
2.3. Enhanced protected effect of Fc-enhance-prophylactic antibody in hFcγR mice



⇒ Anti-Flu mAb increases survival of Influenza A-infected hFcγR mice
⇒ Anti-Flu antibody with enhanced Fc binding improves protective activity and outperforms the WT antibody
⇒ No protective activity shown by Fc-null version of anti-Flu mAb

3. Model upgrades

- hFcγR was intercrossed with the hFcRn mouse model to enable more translatable PK/PD assessment
- Expression of mouse or human FcRn in lung from hFcγR/hFcRn mice was determined by Western Blot analysis



⇒ hFcRn is expressed in lung from hFcγR/hFcRn mice while mFcRn is expressed in lung from WT mice
⇒ Preliminary results show an enhanced PK translatability (data not shown)

Model upgrade	Rational
hFcγR/hSA/hFcRn	Better translatability of PK study of therapeutics with enhanced half-life through binding to hSA or hFcRn
hFcγR/hlgG1	Tolerability to human IgG1 therapeutic antibodies
hFcγR/hSA/hFcRn/Rag1 ^{-/-}	Assessment of translational PK/PD in immunodeficient model for engraftment with human cells
hFcγR/hTFRC	Better biodistribution of therapeutics to the brain through TFRC-mediated drug shuttling
hFcγR/Pan hCD3	Assessment of anti-CD3's efficacy
hFcγR/ immune checkpoint humanized models from genOway's catalog	Flexibility of therapeutics assessment: bispecific or combotherapies targeting immune checkpoints

Conclusion: Humanized FcγR model represents a useful tool for testing efficacy of Fc-engineered antibodies in several therapeutic domains: autoimmunity, immuno-oncology and viral infection.

References:

- Viuff D, Antunes F, Evans L, et al., Generation of a double transgenic humanized neonatal Fc receptor (FcRn)/albumin mouse to study the pharmacokinetics of albumin-linked drugs. J Control Release. 2016;10.1016
- Bournazos S, Wang TT, Ravetch JV. The Role and Function of Fcγ Receptors on Myeloid Cells. Microbiol Spectr. 2016 Dec;4(6):10.1128/microbiolspec.MCHD-0045-2016, 10.1128
- Mössner E, Brünker P, Moser S, et al., Increasing the efficacy of CD20 antibody therapy through the engineering of a new type II anti-CD20 antibody with enhanced direct and immune effector cell-mediated B-cell cytotoxicity. Blood. 2010 Jun 3;115(22):4393-402, 10.1182