		Study Period	Screening	Pre-Treatment [d]		Safety Follow Up															ET
		Study Week	-8 to -7	-6 to -1		1				2		3	4	6	8	10	12	18	24	36	48/ES	
		Study Day (Visit Window)		Apheresis	Pre-infusion -10 to -4	1	2	4 or 5	7±1	11±1	14±2	21±2	28±2	42±2	56±2	70±2	84±3	126±7	168±7	252±7	336±7	
Label	Collection	Testing																				
ADA	7 mL SST	Serum sample for ADA			X								X		X		X				X	X
Exploratory Markers - Serum	7.0 mL SST	Serum sample for exploratory markers			X		X		X		X	X	X	X	X	X	X	X	X	X	X	X
PBMC (collection) [b]	10.0 mL NaHep	PBMC for Cellular Immunogenicity, Exploratory Biomarkers, RCL	X(x2) Ex2		X (x4) Cx1 Ex2 Rx1		X(x2) Ex2	X(x2) Ex2	X(x2) Ex2	X(x2) Ex2	X(x2) Ex2	X(x2) Ex2	X (x3) Cx1 Ex2	X(x2) Ex2	X (x3) Cx1 Ex2	X(x2) Ex2	X (x3) Cx1 Ex1 Rx1	X(x2) Ex2	X (x3) Ex2 Rx1	X(x2) Ex2	X (x3) Cx1 Ex1 Rx1	X (x2) Cx1 Ex1
PK ddPCR	3.0 mL K2EDTA	Blood sample for PK(ddPCR)			X		X[a]	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X
Exploratory Markers - Plasma [c]	4.0 mL K2EDTA	Plasma sample for exploratory markers	X		X(x2)		X		X		X	X	X	X	X	X	X	X	X	X	X	X

Footnotes

General Note: Samples collected outside the listed scheduled visits should use the Unscheduled Visit forms provided at the back of the requisition binder.

[a] The PK sample collected on study Day 2 should be collected at approximately the same time of day that the infusion of study drug took place on study Day 1 (+/- 1 hour). An un scheduled PK sample should be collected within 30 minutes of a suspected infusion related event.

[b] 4 tubes will be collected at Pre-Treatment. 3 tubes will be collected at Day 28, Day 56, Day 84, Day 168 and Day 336. For all other visits only 2 tubes will be collected. X = total, C = Cellular Immunogenicity, E = exploratory biomarkers, R = RCL, number of tubes for each collection designated for each visit as outlined in protocol SOA.

[c] 2 tubes will be collected at Pre-Treatment. For all other visits only 1 tube will be collected.

[d] The Pre-Treatment visit samples may be redrawn any number of times. For redraws of the Pre-Treatment visit samples please use the Pre-Treatment Reassessment Visit forms in the back of the requisition binder next to the Unscheduled Visit forms.

Order of Draw

		Study Period	Screening	Pre-Treatment [e]		Safety Follow Up															ET
		Study Week	-8 to -7	-6 to -1		1				2		3	4	6	8	10	12	24	36	48/ES	
		Study Day (Visit Window)		Apheresis	Pre-infusion 10 to -4	1	2	4 or 5	7±1	11±1	14±2	21±2	28±2	42±2	56±2	70±2	84±3	168±7	252±7	336±7	
Label	Collection	Testing																			
Synovial Fluid Supernatant (d)	4.0 mL NaHep			X (a)									X (b) (c)				X (b) (c)				X (c)
Synovial Fluid Viable Cells (d)				X (a)									X (b) (c)				X (b) (c)				X (c)
Cryo Synovial Biopsy (d)	Bard Mission Needles and 12-Well Plates			X (a)									X (b) (c)								X (c)
FFPE Synovial Biopsy (d)				X (a)									X (b) (c)								X (c)

Footnotes

General Note: Samples collected outside the listed scheduled visits should use the Unscheduled Visit forms provided at the back of the requisition binder.

(a) Synovial biopsy and/or synovial fluid collection during the Pre-treatment period should be performed after enrollment and at least one week prior to the planned dosing visit.

(b) The synovial biopsy/fluid collection can be performed at or up to 1 week after the scheduled assesment visit.

(c) The timing of the synovial biopsy and/or synovial fluid collection in may be changed based on evaluation of data from the first dose escalation cohot.

(d) The synovial biopsy sample/fluid collection is optional at all timepoints.

[e] The Pre-Treatment visit samples may be redrawn any number of times. For redraws of the Pre-Treatment visit samples please use the Pre-Treatment Reassessment Visit forms in the back of the requisition binder next to the Unscheduled Visit forms.

Sample	Tube	Collection & Processing	Aliquots	Shipping	Notes
ADA	 **7.0 mL SST**	Fill tube completely with blood. Immediately after collection, gently invert tubes 5 times. Allow tubes to clot for a minimum of 30 minutes post collection, keeping tubes vertical. The SST tube should NOT be refrigerated prior to centrifugation. Within 60 minutes of collection, centrifuge at 1,800- 2,200xg (RCF not RPM) for 10-15 minutes to allow the gel barrier to migrate between the cell and serum layers. If the serum is not separated from the blood after centrifugation, re-centrifuge the sample until the serum is separated Using a transfer pipette, aliquot 0.5mL of serum each into the first 2 x 2.0 mL cryovials and divide the remaining serum between the other 2 x 2.0mL cryovials, leaving a small amount on top of the separator gel. Discard the collection tube. Store at ≤-20°C until shipping Ship cryovials to MRL frozen in weekly batches.	**4 x 2.0 mL Sarstedt microtube**	**Ship to MRL frozen in weekly batches**	
Exploratory markers - Serum	 **7.0 mL SST**	Fill tube completely with blood. Immediately after collection, gently invert tubes 5 times. Allow tubes to clot for a minimum of 30 minutes post collection, keeping tubes vertical. The SST tube should NOT be refrigerated prior to centrifugation. Within 60 minutes of collection, centrifuge at 1,800- 2,200xg (RCF not RPM) for 10-15 minutes to allow the gel barrier to migrate between the cell and serum layers. If the serum is not separated from the blood after centrifugation, re-centrifuge the sample until the serum is separated Using a transfer pipette, aliquot 0.5mL of serum each into the first 2 x 2.0 mL cryovials and divide the remaining serum between the other 2 x 2.0mL cryovials, leaving a small amount on top of the separator gel. Discard the collection tube. Store at ≤-20°C until shipping Ship cryovials to MRL frozen in weekly batches.	**4 x 2.0 mL Sarstedt microtube**	**Ship to MRL frozen in weekly batches**	
PBMC (collection)	 **10.0 mL NaHep**	Fill all tubes completely with blood. Gently invert 8-10 times to avoid clotting. Do NOT Centrifuge. Do NOT open tube. Ship tube to MRL ambient on day of collection		**Ship to MRL ambient on day of collection**	
Exploratory markers - Plasma	 **4.0 mL K2EDTA**	Fill all tubes completely. Mix tubes after collection by gently inverting 8-10 times and immediately place on wet ice. Within 60 minutes of collection, centrifuge at 4°C at 1800g - 2200g (RCF not RPM) for 10-15 minutes. Using a transfer pipette, aliquot 0.5 mL of plasma into each of 4(8 for Pre-Treatment) x 2.0 mL cryovials and freeze until shipment. Store at ≤-20°C until shipping Ship cryovials to MRL frozen in weekly batches.	**4 or 8 x 2.0 mL Sarstedt microtube**	**Ship to MRL frozen in weekly batches**	

Sample	Tube	Collection & Processing	Aliquots	Shipping	Notes
PK ddPCR	 **3.0 mL K2EDTA**	Fill tube completely as a partial fill results in over anticoagulated plasma, impacting results of testing. Mix tube immediately after collection, gently inverting 8-10 times. Do NOT Centrifuge. Using a pipette, divide blood equally between 2 separate aliquots Freeze immediately on dry ice and keep at ≤-70°C until shipping. Ship cryovials to MRL frozen in weekly batches.	**2 x 2.0 mL Sarstedt microtube**	**Ship to MRL frozen in weekly batches**	
Synovial Fluid Supernatant	 **4.0 mL NaHep**	See Biopsy Manual for instruction.	**2 x 2.0 mL Sarstedt microtube**	**Ship to MRL frozen in weekly batches**	
Synovial Fluid Viable Cells	 **4.0 mL NaHep**	See Biopsy Manual for instruction.	**1 x 2.0 mL Corning Cryovials**	**Ship to MRL frozen in weekly batches**	
Cryo Synovial Biopsy	**Bard Mission Needles and 12-Well Plates**	See Biopsy Manual for instruction.	**2 x 2.0 mL Corning Cryovials**	**Ship to MRL frozen in weekly batches**	
FFPE Synovial Biopsy	**Bard Mission Needles and 12-Well Plates**	See Biopsy Manual for instruction.	**1 x 20.0 mL tube**	**Ship to Mosaic Refrigerated (4C) day of collection**	**DO NOT SEND TO MRL**