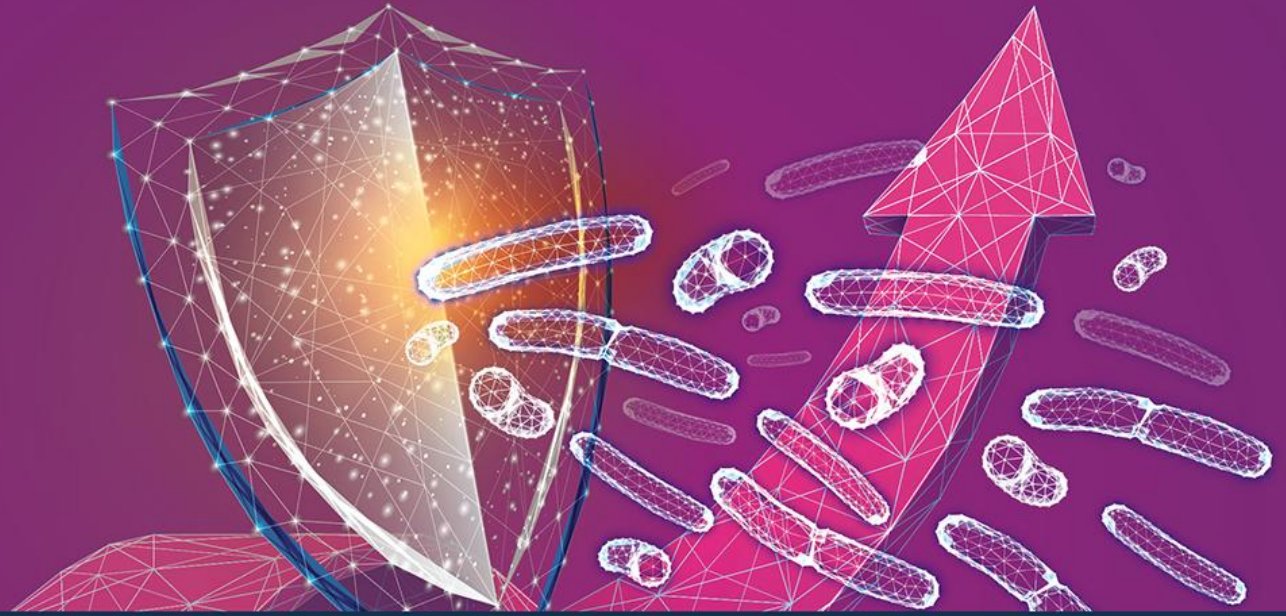




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27-29 October | Washington, DC



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A Global Approach to New Technology Introduction and Validation: Automated Colony Counting

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AstraZeneca



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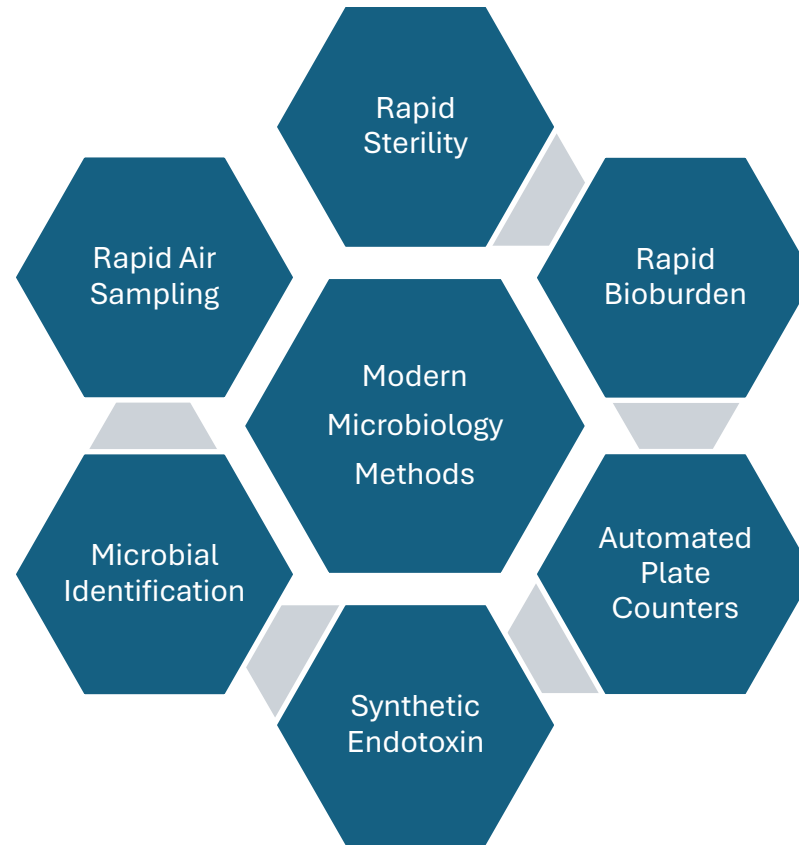
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So much technology, so little time



Reasons for the Global Introduction of Technology: Lessons Learned Through Experience

Local Validation
teams often want
additional validation

Local QA often want
additional
requirements

Local IT have
additional
requirements

Local Health & Safety
requirements vary
globally

Equipment that never
gets unpacked

Equipment that is
unable to be validated

Equipment tagged out
of service on benches
taking up space (still
being calibrated)

New technology
doesn't replace the
existing, both are kept



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Technology Drivers



LICENSE TO
OPERATE



DIGITAL

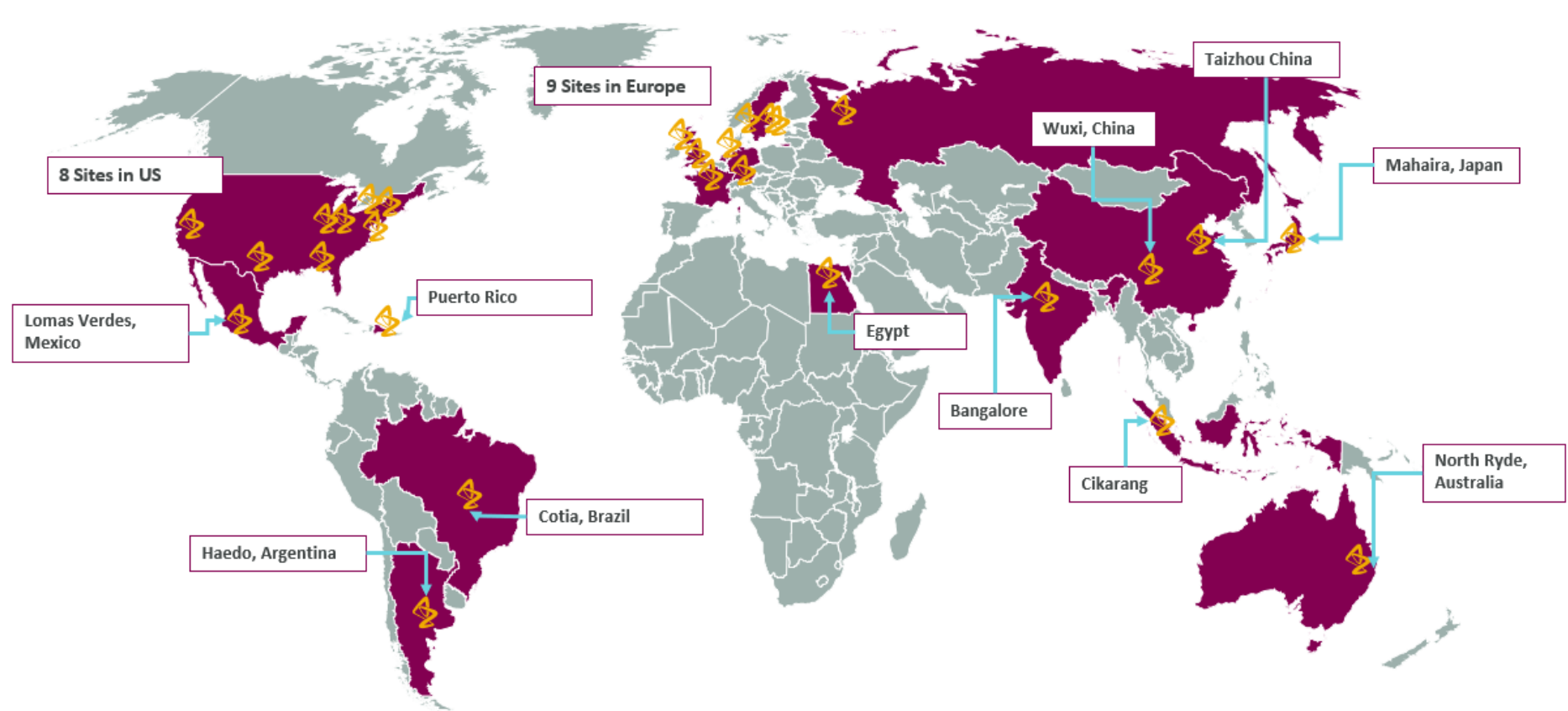


EFFICIENCY

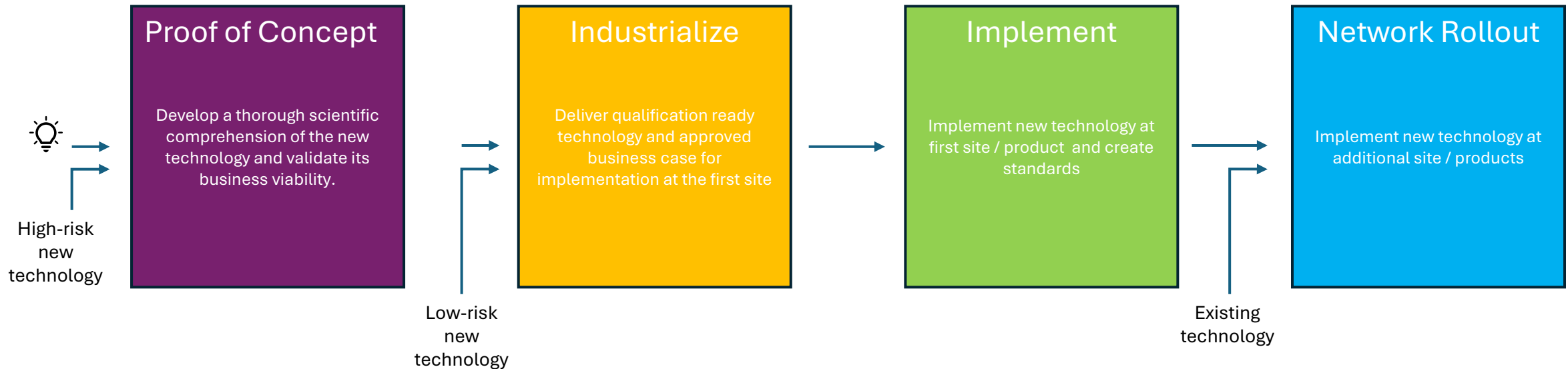


GREAT PLACE
TO WORK

AZ Microbiome



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Case Study

Automated Colony Counting for Environmental Monitoring





Why are AstraZeneca interested?

- 1 Up to 30,000 EM agar plates are read manually and verified every month at large AZ sites
- 2 Annual EM data from aseptic manufacturing facilities shows that >98% of plates are negative
- 3 Occasionally humans make mistakes
- 4 Resolves data integrity challenges



Benefits of this technology

1

APAS processes ~200 plates/hour and sorts them into categories

2

Only plates with growth or processing errors are second checked- vastly reducing technician time

3

Data automatically transferred to LIMS system – manual transcription and chance of error removed

4

Current process plates destroyed on day of reading – All images stored in APAS for 45 days

5

Not media supplier restricted, and different incubation practices can be accommodated



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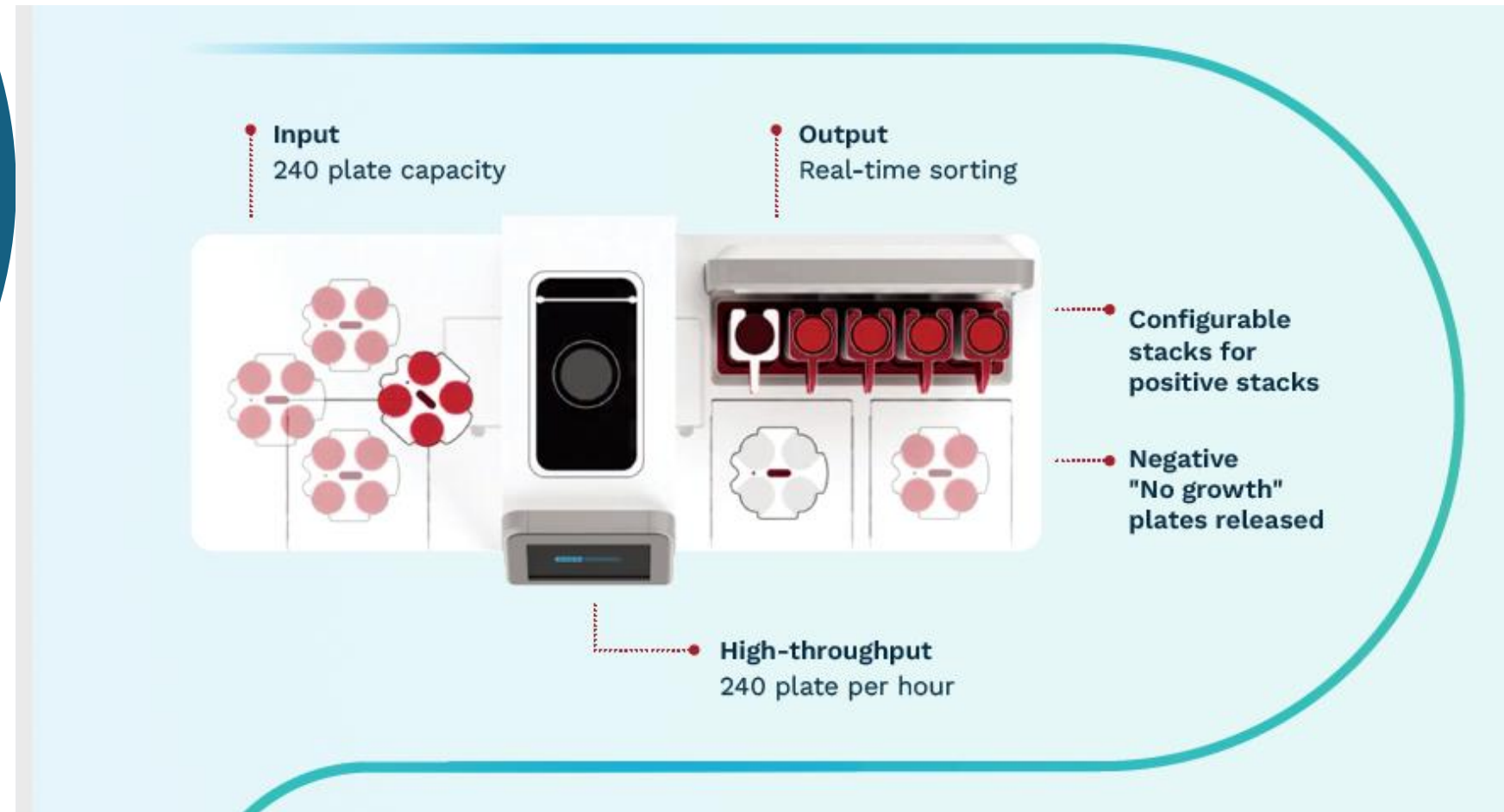
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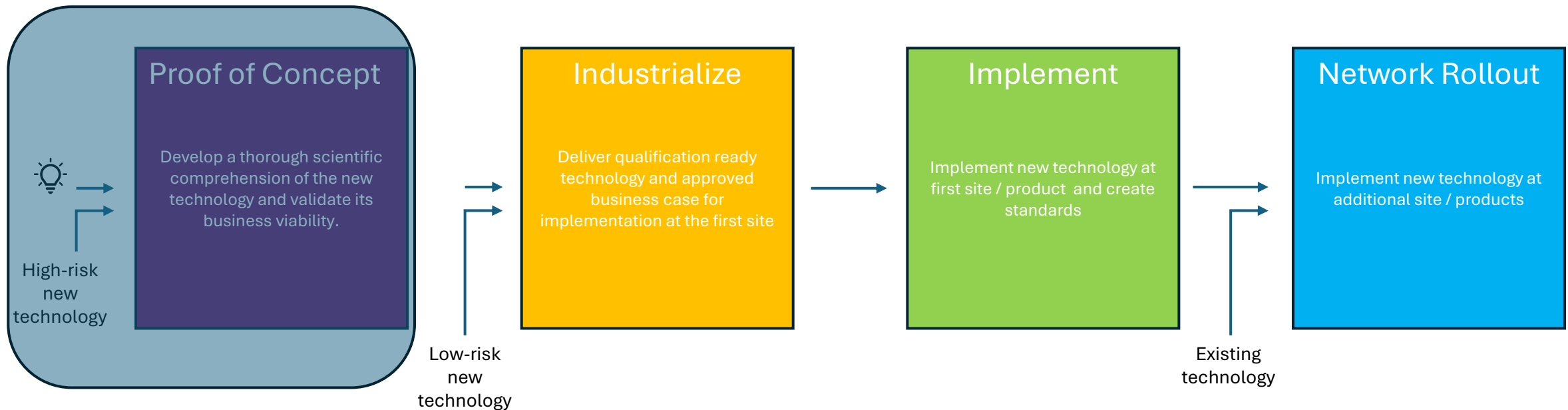
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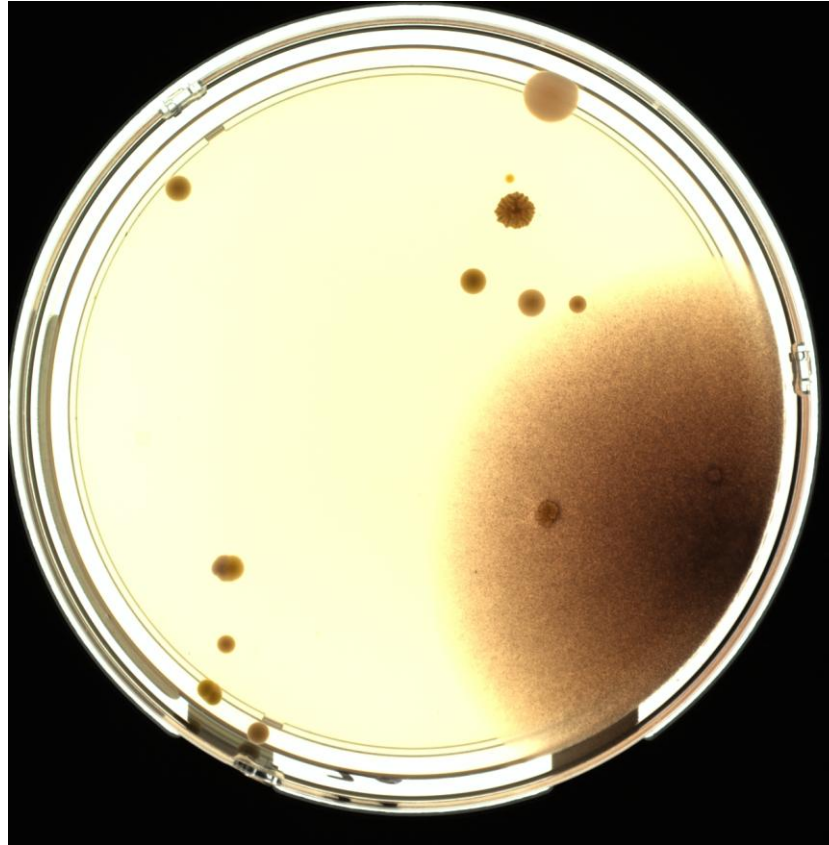
Topographical view



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How many CFU on this plate?



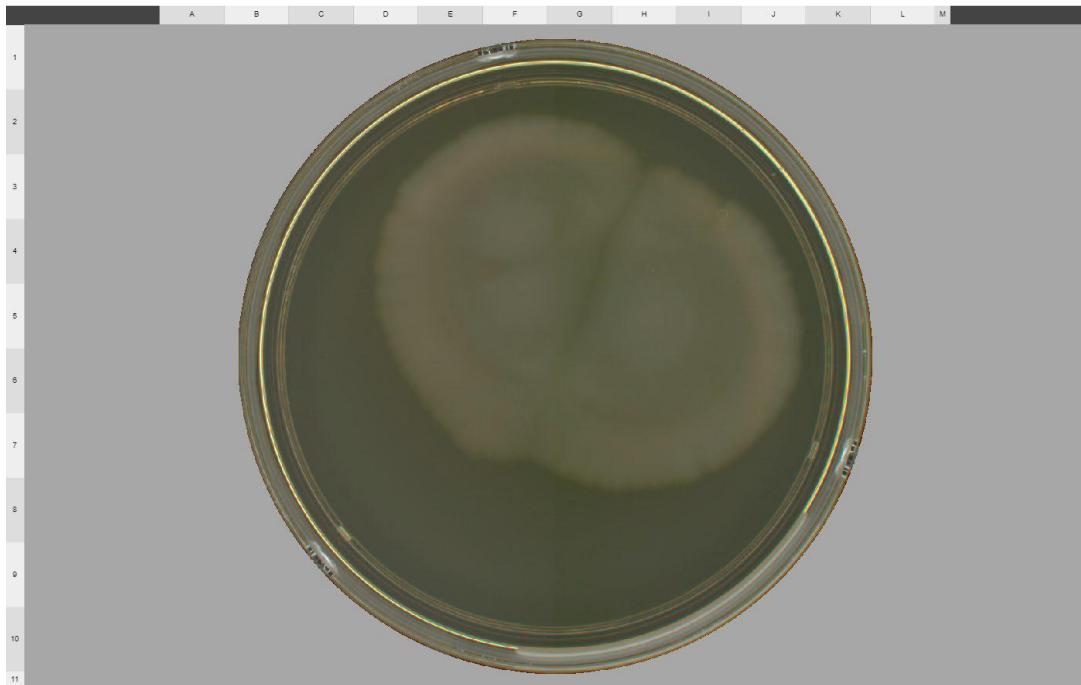
Key Learnings Proof of Concept

Counting is impacted by areas of growth confluence where it is difficult to accurately count and can also be subjective between technicians

Here APAS counts 44 CFU vs actual 5 CFU



Key Learnings from Proof of Concept



For some species, larger and older colonies have significant morphological textural features that influence APAS counting algorithms e.g., *Bacillus* & *Aspergillus*

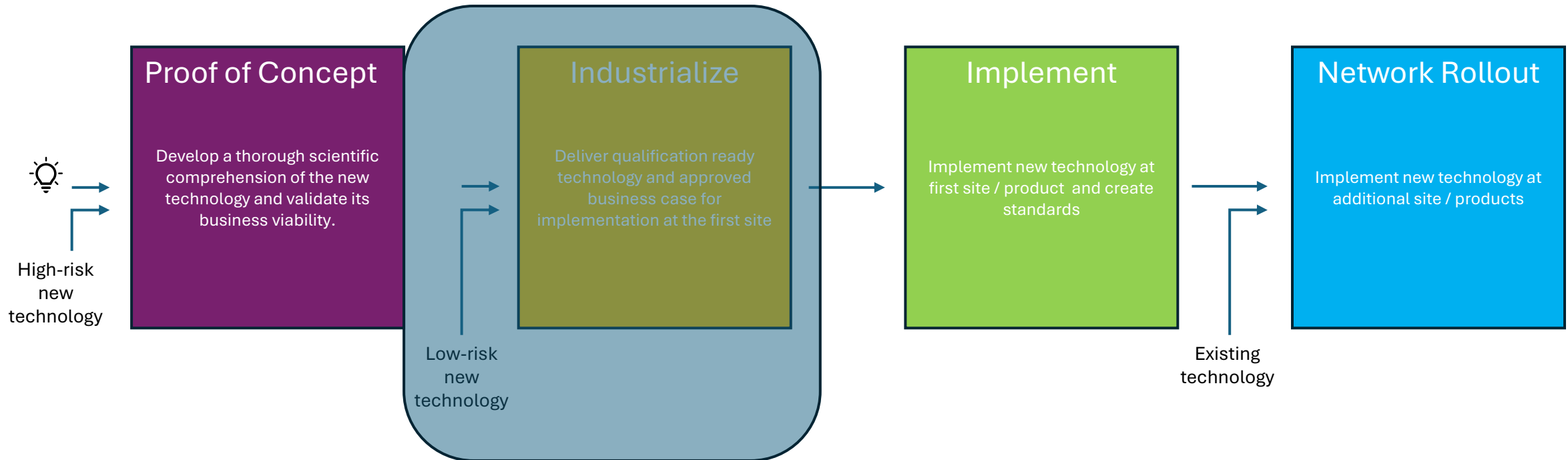
Day 3

APAS counts 3 CFU vs actual 3 CFU

Day 5

APAS counts 26 CFU vs actual 3 CFU

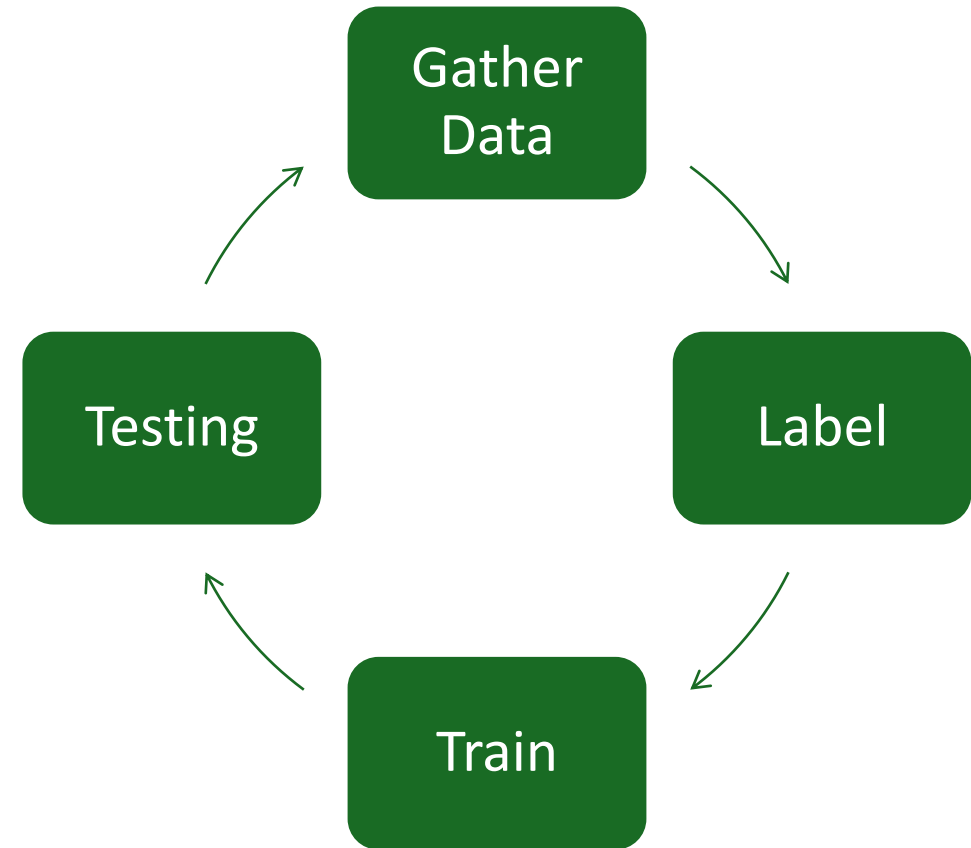
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Industrializing the technology using CCS AI & ML

Challenges to plate reading

- Colony variability
- Agar supplier differences
- Plate labelling by technicians
- Colonies located on the edge or rim
- Condensation
- Plate issues and sampling faults



Industrializing the technology using CCS AI & ML



Data Collection >8000 plates read from AZ, images analyzed, and algorithm developed



Colony Variability: Different morphologies, colony colors, swarming, molds, different *Bacillus* sp, 'wild' and ATCC strains



Agar Plate Variability: Multiple batches of different media suppliers, to accommodate batch to batch variability different plate labelling, different barcodes



Count Variability: Range of counts from 0 CFU – 250 CFU

Supplier Primary Validation

- Primary Validation as per the principles of USP <1223> and Ph.Eur 5.1.6, but isn't really an alternative method per se; it is a different way of reading the results from a traditional method
- Linearity, Precision, Specificity, Accuracy, Robustness, Ruggedness, Operational range, Limit of Detection, Limit of Quantification, Repeatability
- CCS completed Primary Validation March 2024



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Secondary Validation at AZ

Stage 1 Establish Expected Performance

- Positive plates ‘contrived’ by exposing plates in general labs and containing enough negative plates to keep the humans ‘reading’ in representative manner.

Stage 2 Establish Actual Performance

- Actual plates from production environmental monitoring compared to APAS result with the current manual method (two people reading each plate)
- The desired target is non-inferiority to manual read (zero false negatives).

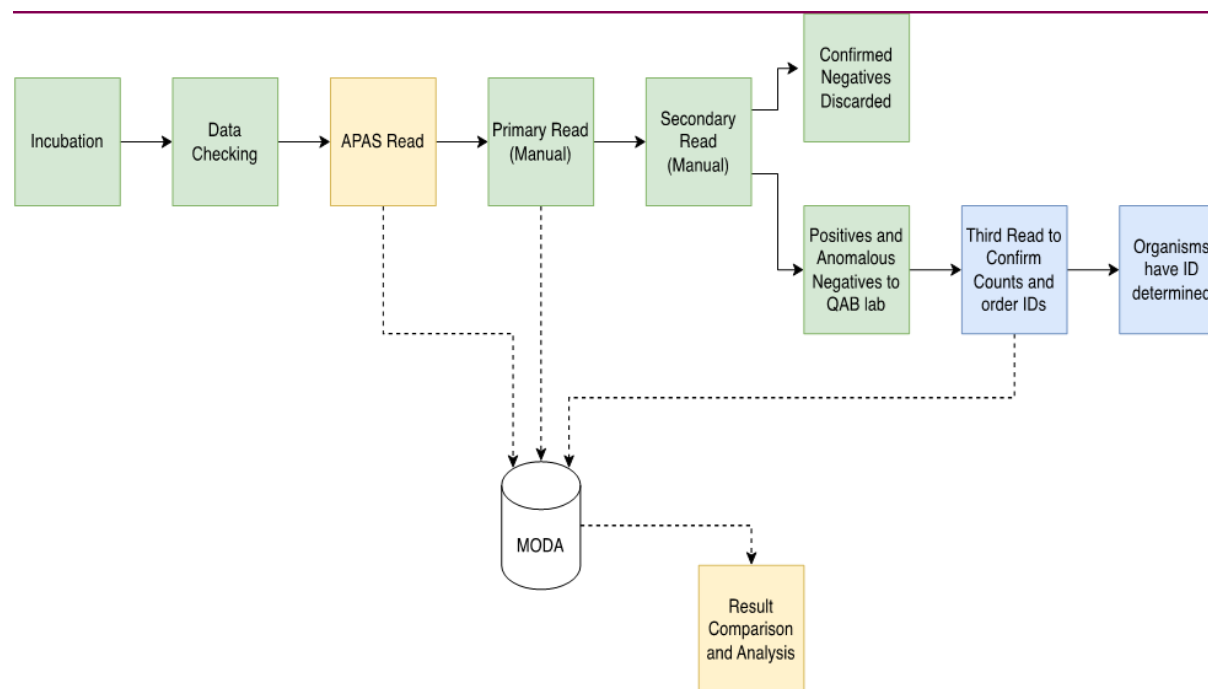
Secondary Validation Study

Establish Actual Performance

APAS instrument used as primary reader for real EM plates.

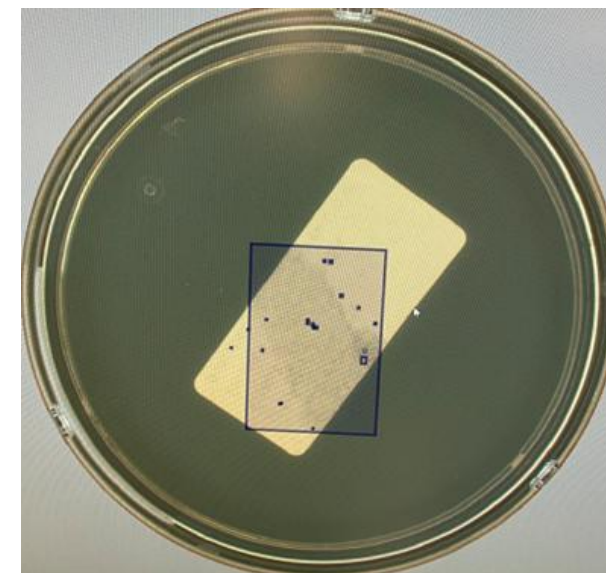
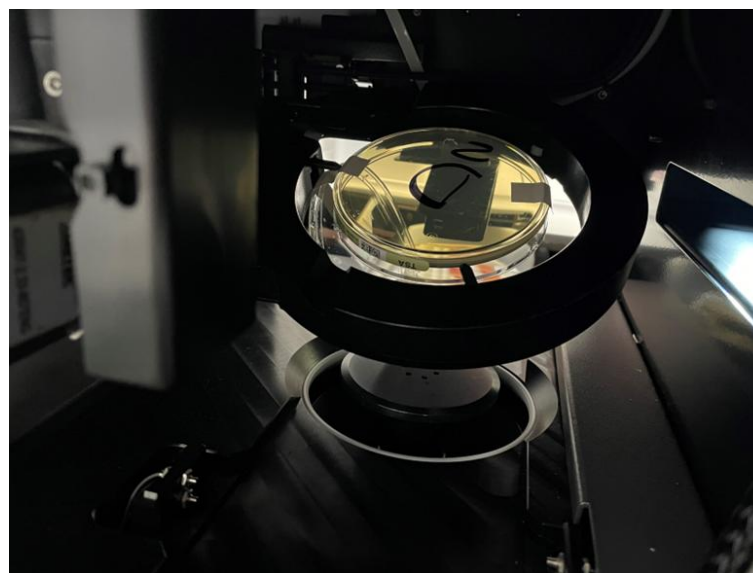
ALL plates checked by humans and results corrected where necessary

Rate of corrections tracked and used to form acceptance criteria for AZ validation



Key Learning Points

Current practices using tape caused mechanical issues driving higher false positive rate



Key Learning Points

Simple solution to utilise clip and bags used at other AZ sites introduced via change control.



Key Learning Points

- Pilot primary validation study has shown difficulties in counting accurately at higher end of the count range especially with some organisms.
- However, this is also seen in humans, where differences of 20-40 colonies have been observed.
- IS THIS IMPORTANT?
- APAS would sort these plates as requiring human review.

Update : Latest software is much improved in this aspect, however in the real world all positive plates are flagged for review.



Figure 3. Example of *B. spizizenii* growth demonstrating variable morphology, size, and confluence

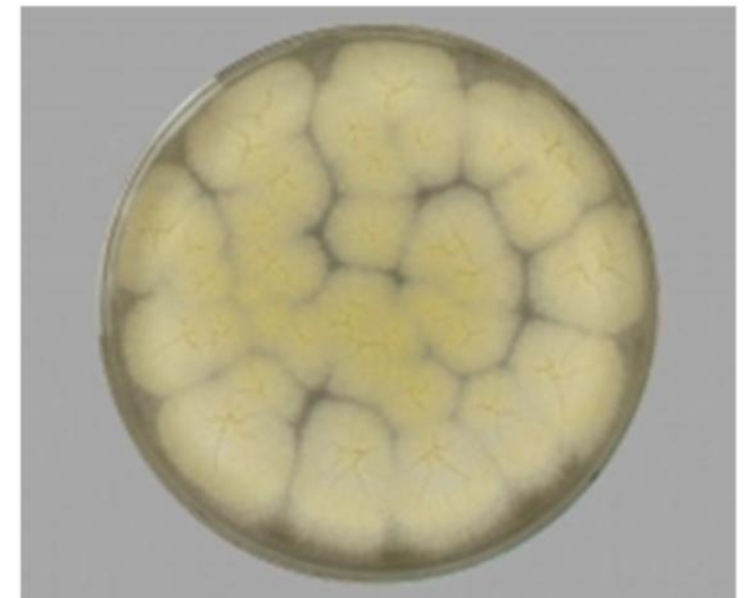
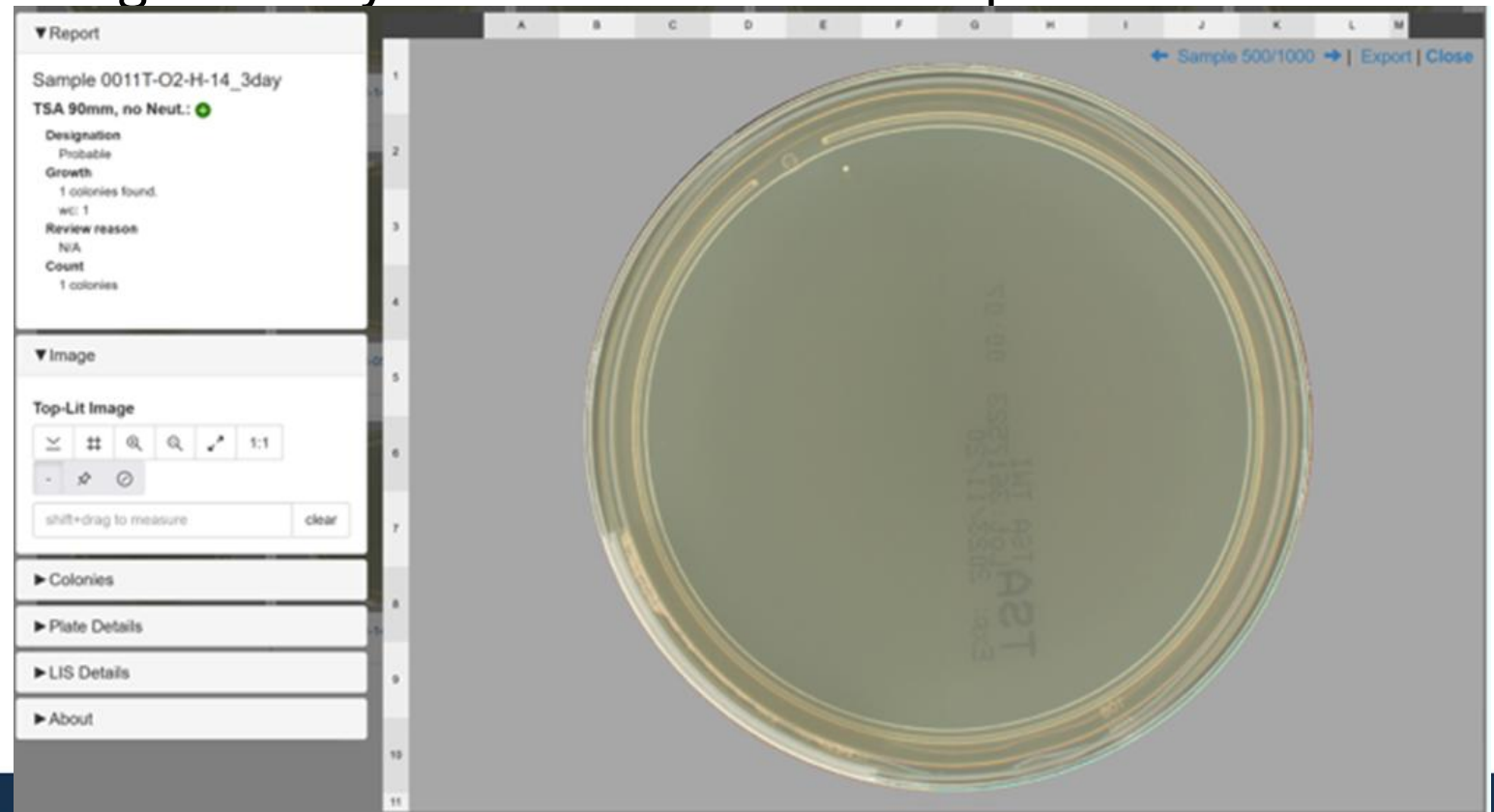
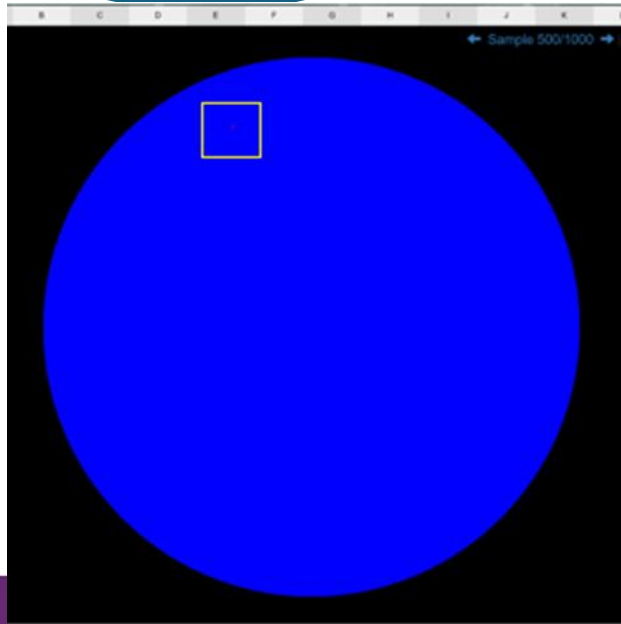


Figure 4. Example of *A. brasiliensis* growth changes over time demonstrating counting challenges

Key Learning Points

- APAS primary function is to sort 'Growth' from 'No Growth'
- Remember, over 98% of plates are zero cfu (AZ facility)
- The difference between 0 and 1 is massive in Grade A, the difference between 15 and 19 is negligible.
- Single colony detection the most important factor.





0012-SA-33-0.5-1
0 colonies found.



0017T-31-09
2 colonies found.



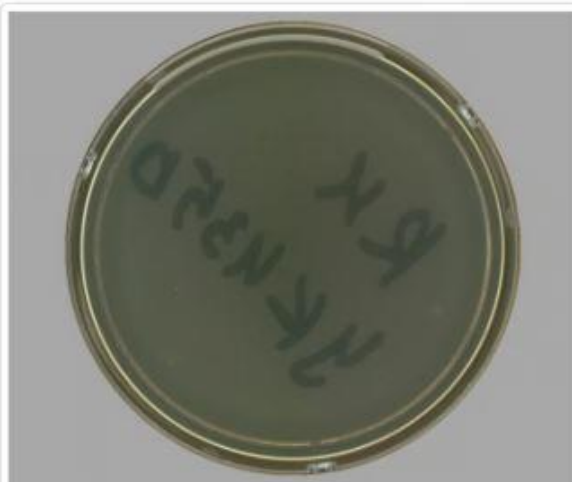
0006BN-A1-U4-2
50 colonies found, 38 bacterial, 12 mould.



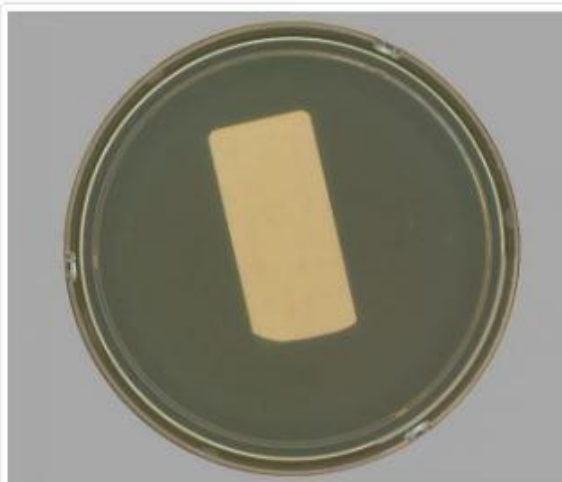
0006BN-A1-U2-1
30 colonies found, 28 bacterial, 2 mould.



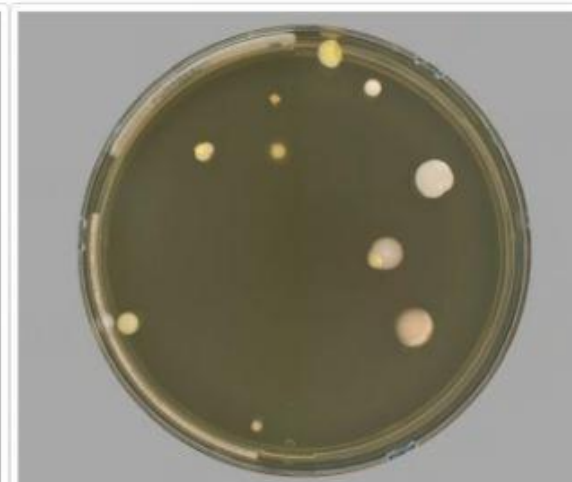
0006TN-D-U4-1
239 colonies found, 214 bacterial, 25 mould.



21003134863-910313
0 colonies found.

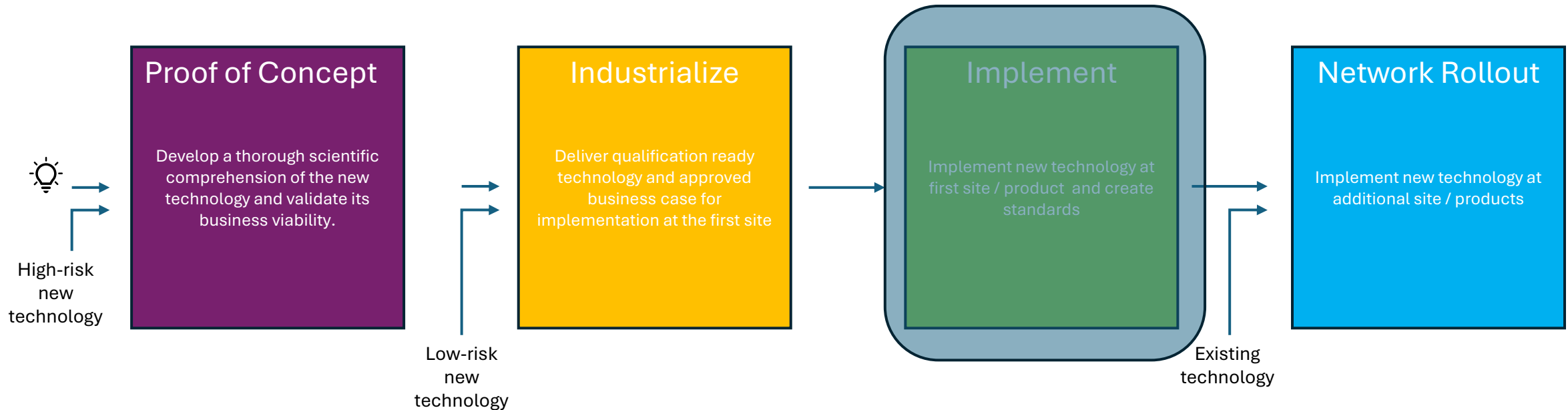


21005290102-910529
0 colonies found.



0006BN-D-U3-2
11 colonies found.

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Comparison of Growth Detection for Manual and APAS

Table 3 – Comparison of microbial growth detection for Manual and APAS plate reads when APAS image and review of agar plates is used as the 'Truth state'.

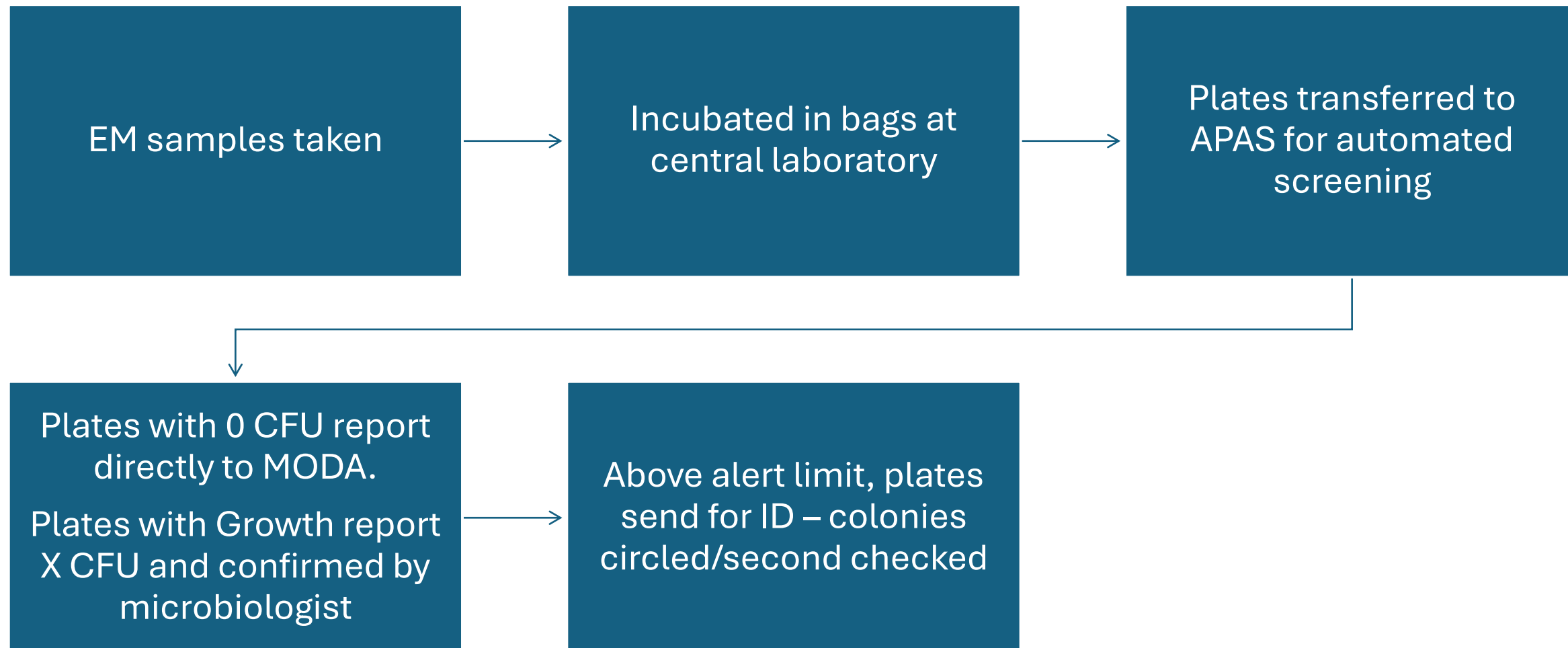
	Number of 90mm plates analysed	Number of plates with growth ^a	Number of plates with growth detected by Manual	Number of plates with growth detected by APAS	Accuracy at growth detection Manual	Accuracy at growth detection APAS	Manual False Negative Rate	APAS False Negative Rate
Stage 1	2556	1924	1851 ^b	1872	96.2%	97.3%	3.8%	2.7%
Stage 2	8548	492	475 ^c	474	96.5%	96.3%	3.5%	3.7%
Overall	11104	2416	2326	2346	96.3%	97.1%	3.7%	2.9%

^a – Presence of growth determined by analysis of each APAS image (top lit and bottom lit images) and the agar plates.

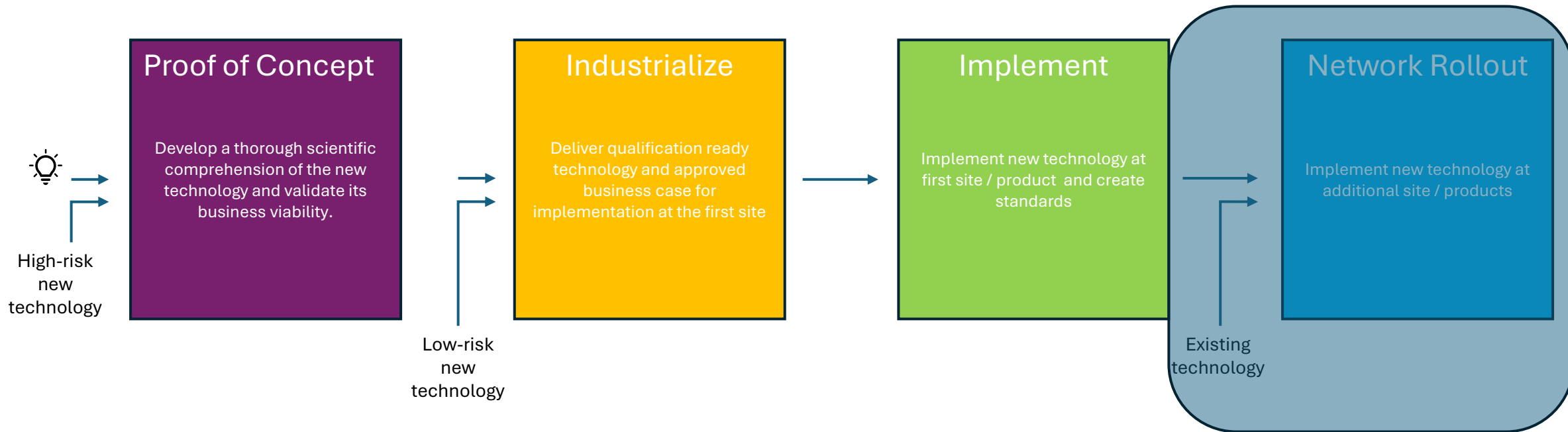
^b - In Stage 1 there were 3 separate and independent manual counts. If any of the 3 manual readers missed growth on a plate, it was counted as a miss/non-detect event. For APAS a miss/non-detect event was when APAS did not 'box' and detect microbial growth which was visible on the image/plate.

^c – In Stage 2 the Manual count was defined by the current plate reading process which is the agreement of 2 separate reads e.g. read and checked result entered in to MODA. A miss/non-detect for manual readers in this instance was when there was microbial growth on the image/agar plate, but the result in MODA was recorded as 0 cfu. For APAS a miss/non-detect event was when APAS had not 'boxed' and detected microbial growth which was visible on the image.

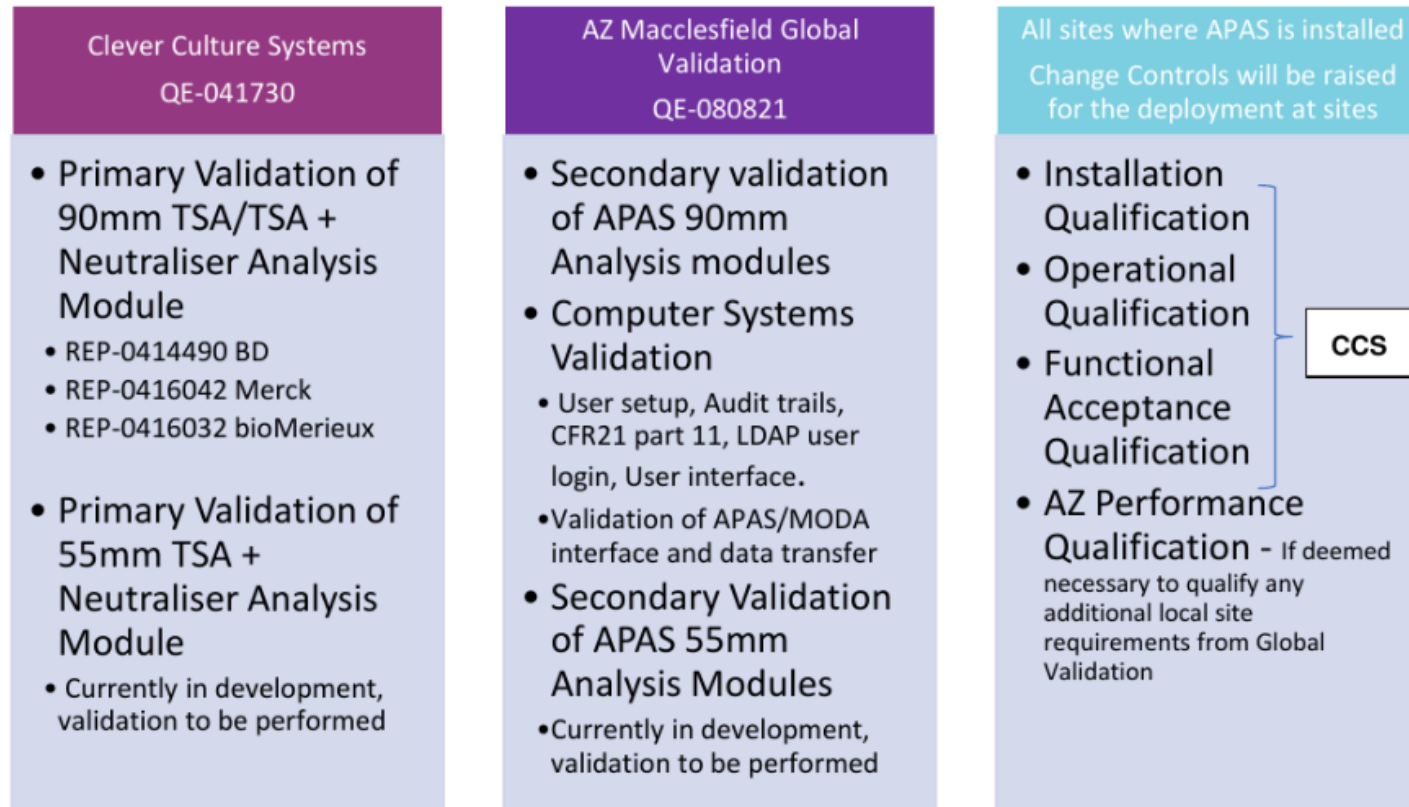
Automated Colony Counting Workflow



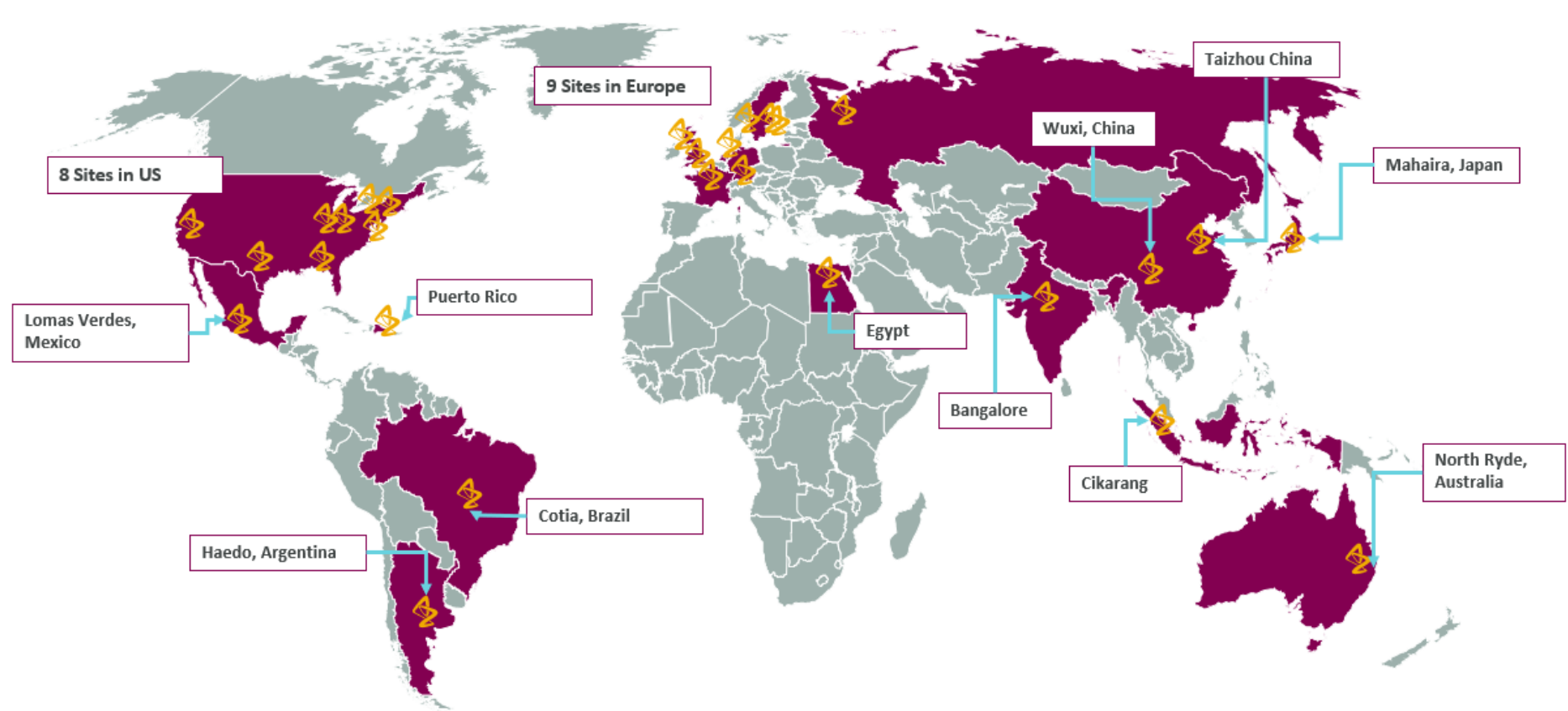
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Network Rollout Validation Strategy



AZ Microbiome



Conclusions



Accurate colony counting is impacted by areas of growth confluence. The “correct” colony count is also subjective between technicians



APAS estimates CFU for plates with growth and sorts them as ‘requiring human review’. Therefore, the false negative rate is more important



APAS performance met our acceptance criteria and is equivalent and non-inferior to the manual method



Using APAS technology brings additional benefits such as data integrity improvements,



elimination of human error, standardization, job role enhancement for microbiologists and the potential that with advancements and improvements, the technology will become even better.

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AstraZeneca

Jack Brown – Corporate
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Clever Culture
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Steve Giglio – Scientific
Director, Clever
Culture Systems

Miriam Guest – Former
Principal Scientist,
AstraZeneca (now
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