

1 **Same class, different activity:**
2 **Delamanid and pretomanid have comparable bactericidal activity but pretomanid potently**
3 **inhibits *Mycobacterium tuberculosis* ribosomal rRNA synthesis**
4

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24 Keywords: antibiotic, tuberculosis, pharmacodynamic, concentration response, dose ranging,
25 bactericidal activity, sterilizing activity

26 Short title: Differing activity of delaminid and pretomanid
27

28 ABSTRACT

29 **Background.** The nitroimidazoles delamanid and pretomanid play an important role in
30 contemporary tuberculosis treatment. It is unclear whether delamanid and pretomanid have
31 meaningfully different activity since both reduce *Mycobacterium tuberculosis* colony forming
32 units (CFU) similarly in animal models. The RS ratio is a pharmacodynamic marker of ongoing
33 rRNA synthesis that has been associated with treatment-shortening (*i.e.*, sterilizing) activity.

34 **Methods.** Using *Mycobacterium tuberculosis Erdman*, we conducted dose-ranging studies in
35 aerobic axenic culture and in the conventional BALB/c mouse high-dose aerosol infection model
36 to compare bactericidal and RS ratio activity of delamanid and pretomanid.

37 **Results.** *In vitro* concentration-response curves showed that delamanid and pretomanid had
38 similar RS ratio effect at maximal concentration but pretomanid was more potent, achieving 90%
39 of the maximal effect (RS-EC₉₀) at a lower concentration (390 ng/mL) than delamanid (810
40 ng/mL). In mice, delamanid and pretomanid had similar effects on CFU. Human-equivalent
41 doses of delamanid (6 mg/kg) and pretomanid (50 mg/kg) resulted in plasma C_{max} concentrations
42 well below (210 ng/mL) and well above (7,825 ng/mL) the RS-EC₉₀, respectively. Delamanid
43 displayed no discernable RS ratio response, even at 16-times the human-equivalent dose. Higher
44 pretomanid doses resulted in significantly greater RS ratio effects.

45 **Conclusions.** We found that delamanid and pretomanid have similar bactericidal activity but
46 pretomanid has superior RS ratio activity. Meaningful differences between drugs within the same
47 class were not captured by conventional CFU-based pharmacodynamics, supporting the value of
48 measuring orthogonal drug effects such as the RS ratio.

49 **LAY SUMMARY**

50 Antibiotics in the nitroimidazole class are used in treatment of drug-resistant tuberculosis.
51 There are two approved nitroimidazole antibiotics: delamanid and pretomanid. For decades, it
52 has been unclear whether delamanid and pretomanid are interchangeable or whether they affect
53 the bacterium *M. tuberculosis* differently. Most studies of the effect of antibiotics count the
54 number of bacterial colonies that form on a culture plate. “Colony forming units” tell us about
55 change in bacterial burden but does not give information about bacterial health. A new way of
56 thinking about antibiotic effect is the RS ratio. The RS ratio is a test that measures how much
57 ribosomal RNA synthesis is ongoing. Ribosomal RNA synthesis is a “vital sign” of bacterial
58 health and activity. The key finding of this study is that although the two nitroimidazole
59 antibiotics look the same in terms of their effect on bacterial burden, they have different effects
60 on bacterial health. This information deepens understanding of differences between two
61 clinically important antibiotics. It also shows that antibiotics testing should consider not only
62 bacterial burden but also new tests of bacterial health.

63 INTRODUCTION

64 There is an urgent need for shorter, more effective tuberculosis (TB) treatments. An
65 important new class of anti-TB drugs is the nitroimidazoles. Two nitroimidazoles, delamanid and
66 pretomanid, received US Food and Drug Administration approval for drug-resistant TB in 2014
67 and 2019, respectively, and pretomanid is a component of World Health Organization
68 recommended treatment of multiple-drug resistant TB.¹

69 Despite intensive preclinical and clinical evaluation, there remains considerable
70 uncertainty about whether delamanid and pretomanid have a meaningfully different activity or
71 are interchangeable.² As comprehensively reviewed recently, delamanid and pretomanid are
72 structurally similar and share a dual mode of action which includes inhibition of mycolic acid
73 synthesis and respiratory poisoning. However, delamanid inhibits two mycolic acid classes
74 (ketomycolates and methoxymycolates) while pretomanid inhibits only one (ketomycolates).³⁻⁵
75 Additionally, while both appear to poison *M. tuberculosis* (*Mtb*) respiration, delamanid does so
76 via an NAD-adduct⁶ while pretomanid generates reactive nitrogen.^{7,8} When administered at an
77 identical dose in murine models, delamanid exhibits greater bactericidal activity (*i.e.*, greater
78 reduction in colony forming units (CFU)) than does pretomanid. However, when given at lower
79 doses designed to mimic human AUC, the bactericidal activity of delamanid and pretomanid are
80 similar.² Because pre-clinical studies have not clearly differentiated the activity of delamanid and
81 pretomanid, there has been persistent uncertainty regarding optimal usage in humans. As a result,
82 considerable resources are currently being expended to support head-to-head testing of
83 delamanid versus pretomanid in human clinical trials.^{9,10}

84 One reason the activity of delamanid and pretomanid have been difficult to differentiate
85 in preclinical studies is that efficacy has generally been estimated in terms of CFU. CFU
86 enumerates the burden of *Mtb* capable of growth on agar but provides little information about the
87 effect of drugs on *Mtb* physiology. In the conventional BALB/c high-dose aerosol (HDA) mouse
88 infection model, which is the workhorse of *in vivo* efficacy testing, early bactericidal activity
89 does not reliably predict time to achieve durable cure.¹¹ Therefore, long-term murine relapse
90 studies are conventionally used to quantify treatment shortening in terms of treatment time
91 required to prevent relapse in 95% of mice (T₉₅).¹²

92 An alternative to CFU is measuring “pathogen health,” meaning how drugs affect *Mtb*
93 physiology.^{13–18} Drugs or regimens that reduce CFU to equivalent degrees may force unique
94 patterns of bacterial injury and physiologic adaptation.^{16,17} One measure of pathogen health is the
95 RS ratio[®] assay which quantifies ongoing *Mtb* rRNA synthesis.¹⁷ The capacity of drugs and
96 regimens to rapidly and profoundly suppress the RS ratio in mice has been associated with their
97 ability to more rapidly achieve non-relapsing cure (*i.e.*, treatment shortening).¹⁷ Here, we used
98 both CFU and RS ratio to compare the effects of delamanid and pretomanid *in vitro* and in the
99 BALB/c HDA mouse model.

100

101 **METHODS**

102 *In vitro experiments*

103 Cultures of *Mtb* Erdman were propagated in 7H9 medium containing 850 mg/L NaCl, 0.2%
104 glycerol, 0.2% glucose, 0.5% BSA, and 0.05% tween80. *Mtb* cultures were grown to mid-log

105 and then transferred into sterile glass test tubes (20 by 125 mm) containing a 12 by 4.5 mm stir
106 bar at an $OD_{600} = 0.05$ and a volume of 5 mL. Cultures were agitated at ~200 rpm under the
107 control of a rotary magnetic tumble stirrer at 37°C in 5% CO₂. Sterile 1000X stocks of delamanid
108 and pretomanid prepared in DMSO were added after 18h of outgrowth. An initial 5-fold dilution
109 series starting from 10 µg/mL of each drug was performed to determine an approximate
110 concentration at which each drug achieved 90% of their respective RS- E_{max} values (RS-EC₉₀).
111 Along with this, a concentration 10-fold higher than the observed MIC value for each drug in
112 this model, 16 µg/mL and 4 µg/mL for delamanid and pretomanid respectively, was also
113 included. After which, a 2-fold dilution series starting from 8X the RC-EC₉₀ was performed
114 starting from 6.4 µg/mL and 3.2 µg/mL for delamanid and pretomanid respectively. RNA was
115 collected from cultures both prior to drug exposure and after 48h of drug exposure as described
116 ¹⁴. RNA collection, isolation, and ddPCR were performed as described ^{14,17}. RS ratio effect dose
117 response was defined using an E_{max} 4-parameter variable slope sigmoid response model as
118 described ¹⁴. Dose response curves were generated and analyzed using GraphPad Prism 9.
119 Curves were fit to the observed data using least squares regression and medium convergence
120 criteria (maximum iterations = 1000). RS ratio percent effect corresponded to the percent
121 decrease in RS ratio after 48h drug exposure relative to controls prior to drug exposure.

122 *Murine experiments*

123 Animal studies were performed at Colorado State University (CSU) in ABSL-3
124 containment in accordance with CSU Institutional Animal Care and Use Committee (reference
125 number: 4179) guidelines. Six- to eight-week-old female pathogen-free BALB/c mice (Jackson
126 Laboratories) were infected by aerosol (Glas-Col) with *Mtb* Erdman to achieve deposition of

127 ~3.7 log₁₀ CFU in the lungs one day following high-dose aerosol infection (day -10).¹⁹
128 Treatment by oral gavage, five days per week was initiated 11 days post aerosol (day 0).
129 Pretomanid (Chemshuttle) was prepared in the CM-2 formulation as previously described.¹⁹
130 Delamanid (ChemShuttle) was prepared in 5% gum Arabic (Sigma). Prepared pretomanid or
131 delamanid were administered 5 of 7 days per week by oral gavage in 0.2 mL volume quaque die
132 (QD; once per day) at 10, 50, or 100 mg/kg and 6, 10, 50, and 100 mg/kg, respectively. Mice
133 were humanely euthanized on day 0 as a pre-treatment control, and on treatment days 5, 12 and
134 26. Lungs were flash frozen under liquid nitrogen for RNA and CFU enumeration. Details on RS
135 ratio assessment were described previously.^{13,17} For CFU, left, inferior, and post-caval lung lobes
136 (2/3rds by weight) were homogenized using the Bertin Precellys CKMix50-7 mL lysis kit [in 4.5
137 mL phosphate-buffered saline (PBS) with 10% bovine serum albumin (BSA)]. Lung
138 homogenates were plated as serial dilutions on 0.4% charcoal-supplemented 7H11 agar (i.e.,
139 Middlebrook 7H11 agar plates supplemented 0.2% [v:v] glycerol, 10% [v:v] oleic acid-albumin-
140 dextrose-catalase (OADC) supplement, and 0.01 mg/mL cycloheximide, and 0.05 mg/mL
141 carbenicillin). For RNA, flash frozen superior and middle lung lobes were homogenized using
142 the Bertin Precellys CKMix50-7 mL lysis kit in 1.5 mL CAMM-RPS buffer.^{13,17}

143 *Statistical analysis*

144 The RS ratio effect at maximal concentration (RS-E_{max}) and concentration required to achieve
145 90% of the maximal attainable effect (RS-EC₉₀) were calculated as previously described.²⁰ All *p*-
146 values for pairwise comparisons were calculated using the Wilcoxon rank-sum test. Results with
147 *p* < 0.05 were considered statistically significant. Analyses and graphics were conducted using R

148 version 4.5.1 (R Development Core Team, Vienna, Austria <https://www.r-project.org>) and
149 RStudio version 2025.05.1+513 (<https://posit.co>).

150 **RESULTS**

151 *RS ratio activity in vitro*

152 In *in vitro* dose-response curves,¹⁴ delamanid and pretomanid had indistinguishable RS
153 ratio effects at maximal concentration (RS-E_{max}), indicating equivalent RS ratio efficacy (**Fig**
154 **1a-b**). However, the dose required to achieve 90% of the maximal attainable effect (RS-EC₉₀)
155 was higher for delamanid (810 ng/mL) than for pretomanid (390 ng/mL). Achieving the same
156 effect at a lower concentration indicates that pretomanid has more potent RS ratio activity.

157 *Dose ranging in BALB/c mice*

158 In a murine dose ranging study, no dose of delamanid reached the *in vitro* delamanid RS-
159 EC₉₀ of 810 ng/mL (**Fig 1a, Fig 2a**). For example, the 100 mg/kg delamanid dose (*i.e.*, >16-
160 times higher than the established human equivalent 6 mg/kg dose) resulted in a C_{max} of 707
161 ng/mL.

162 By contrast, the standard human-equivalent pretomanid dose (50 mg/kg) resulted in a
163 C_{max} of 7,825 ng/mL, far exceeding the *in vitro* pretomanid RS-EC₉₀ of 310 ng/mL (**Fig 1b, Fig**
164 **2b**). At the standard human equivalent dose of 50 mg/kg, the trough pretomanid concentration
165 was 78 ng/mL. The highest dose of pretomanid (100 mg/kg) resulted in higher C_{max} and trough
166 concentrations than standard dosing although the trough concentration remained lower than the
167 *in vitro* pretomanid RS-EC₉₀.

168 *RS ratio activity in mice*

169 Consistent with the lower-than-RS-EC₉₀ plasma concentrations described above,
170 delamanid had no discernable RS ratio dose-response relationship in mice (**Fig 2c**). By contrast,
171 increasing pretomanid doses resulted in progressively greater decreases in the RS ratio (**Fig 2d**).
172 When compared at human-equivalent doses (*i.e.*, 6 mg/kg and 50 mg/kg for delamanid and
173 pretomanid, respectively), pretomanid had significantly greater RS ratio activity than delamanid
174 ($P=0.008$ at all timepoints). When delamanid and pretomanid were given at an identical dose
175 (resulting in a delamanid dose substantially exceeding human exposures), pretomanid continued
176 to have significantly greater RS ratio activity than delamanid (**Table 1**). For example, when both
177 drugs were given at a dose of 50 mg/kg, pretomanid had significantly greater RS ratio activity
178 than delamanid ($P=0.008$ at all treatment days).

179 ***Bactericidal activity in mice***

180 Both delamanid and pretomanid displayed a dose-response relationship in their effect on
181 CFU (**Fig 2e-f**). Consistent with previous studies, delamanid and pretomanid had
182 indistinguishable bactericidal activity when compared at respective human-equivalent doses at
183 day 5 ($P=0.75$) and day 26 ($P=0.15$). At day 12, the human-equivalent pretomanid dose
184 decreased CFU marginally but significantly more than the human-equivalent delamanid dose
185 ($P=0.04$). When delamanid and pretomanid were given at the same dose, delamanid generally
186 had significantly greater bactericidal activity (**Table 2**). For example, when both drugs were
187 given at 50 mg/kg, delamanid reduced CFU significantly more than pretomanid at day 12
188 ($P=0.01$) and at day 26 ($P=0.008$).

189 **DISCUSSION**

190 Our *in vitro* and murine studies revealed a hitherto unreported difference between the
191 effects of delamanid and pretomanid. Although the two drugs had similar bactericidal activity,

192 we found that pretomanid had more potent RS ratio activity than delamanid. Both in
193 monotherapy and in combination regimens, pretomanid decreased the RS ratio more than
194 delamanid. These results highlight that there exist significant differences between drugs that are
195 not captured by the conventional PD marker (CFU burden). Molecular measures of *Mtb*
196 physiology such as the RS ratio may support drug and regimen development by differentiating
197 between the activity of drugs or regimens that have the same effect on CFU.

198 Conventional preclinical testing has left a state of equipoise in which it is unclear whether
199 delamanid and pretomanid have different activity or are interchangeable.² Quantifying drug
200 activity in preclinical models has traditionally depended heavily on enumeration of *Mtb* growth
201 on agar plates.¹¹ Our previous work has suggested that measurement of properties other than
202 pathogen burden may augment understanding of drug effects.^{13,15,15-17} Specifically, the RS ratio
203 measures a fundamental indicator pathogen health and activity via assessment of ongoing rRNA
204 synthesis rather than bacterial burden. We have shown that the RS ratio provides orthogonal
205 information that is distinct from CFU¹⁸ and may distinguish between drugs or regimens that
206 have comparable bactericidal activity.^{17,19} Understanding of drug effects is often influenced by
207 the PD marker used. Here, the traditional marker of drug effect (CFU) suggested that delamanid
208 is significantly more potent than pretomanid whereas the RS ratio suggested the opposite.

209 TB drug development has been impeded by a “portability” problem in which
210 conventional *in vitro* microbiologic measures of drug activity (*i.e.*, minimum inhibitory
211 concentration (MIC) and minimum bactericidal activity (MBC) generally do not translate
212 directly to *in vivo* drug activity. Translation from *in vitro* microbiologic assays to *in vivo* results
213 has required combinations of specialized conditions. Examples include testing in *ex vivo*
214 caseum²¹ or conditions meant to mimic other *in vivo* conditions followed by integrative

215 modeling.²² By contrast, the *in vitro* RS ratio activity of delamanid and pretomanid was directly
216 concordant with their *in vivo* RS ratio activity in mice. Specifically, *in vitro* studies showed that
217 delamanid required a higher concentration than pretomanid to suppress the RS ratio. Murine
218 studies showed that the delamanid RS-EC₉₀ was not achieved and, correspondingly, there was
219 minimal RS ratio effect. If confirmed with additional drug classes, portability of RS ratio results
220 from *in vitro* to mouse could de-risk and accelerate the progression from early drug discovery to
221 animal studies. It would provide drug developers with greater confidence in advancing animal
222 testing and enable dose-projection needed to achieve RS ratio activity.

223 This work has several limitations. Our *in vitro* concentrations-response analysis indicated
224 that delamanid achieved the same RS ratio efficacy (RS-E_{max}) as pretomanid if provided at
225 sufficiently high concentration. With the intention of exceeding the *in vitro* delamanid RS-EC₉₀,
226 we treated mice with delamanid 100 mg/kg, a dose >16-times higher than human equivalent
227 dose. However, even delamanid administered to mice at 100 mg/kg failed to reach the RS-EC₉₀
228 concentration in plasma and, correspondingly, we observed no RS ratio effect in mice. This
229 suggests an even higher delamanid dose would be required to achieve an RS ratio effect and
230 highlights the relatively lower RS ratio potency of delamanid. Second, this work was performed
231 in a single mouse model (BALB/c). Future assessment in other models, including the C3HeB/FeJ
232 mouse, would provide added value in terms of the impact of advanced disease severity and
233 heterogeneous pathology.²³

234 This work provides a new perspective on the activity of two nitroimidazoles which play
235 important roles in contemporary TB treatment regimens. CFU, the conventional culture-based
236 measure of pathogen burden, showed delamanid and pretomanid had similar bactericidal activity
237 when administered at their respective human-equivalent dose, but the RS ratio revealed that they

238 differed in their capacity to inhibit bacterial rRNA synthesis, a fundamental physiologic process.
239 RS ratio activity has previously been associated with treatment-shortening activity in the
240 BALB/c mouse model,¹⁷ suggesting this previously unappreciated difference between delamanid
241 and pretomanid is consequential. Given the high cost and long timelines of TB regimen
242 development, we believe that there is risk in continuing to “put all eggs in a single [PD] basket”
243 by focusing exclusively on CFU burden. Instead, we propose that preclinical TB drug and
244 regimen evaluation should include not only CFU burden but also molecular measures of
245 pathogen physiologic health.

246 **Funding.** NDW acknowledges funding from Veterans Affairs 1I01BX004527-01A1. NW, MV
247 and GR acknowledge funding from NIH UM1 AI179699. The funders had no role in study
248 design, data collection and analysis, decision to publish, or preparation of the manuscript.

249 **Conflicts of Interest.** The authors have no conflicts of interest.

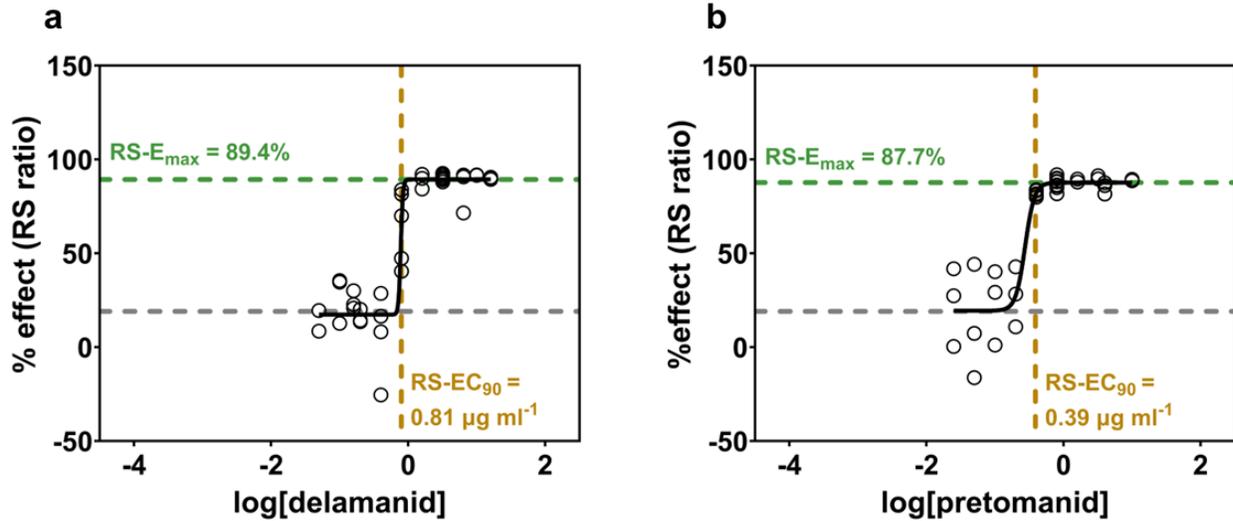
250 REFERENCES

- 251 1. Saukkonen JJ, Duarte R, Munsiff SS, Winston CA, Mammen MJ, Abubakar I, et al.
252 Updates on the Treatment of Drug-Susceptible and Drug-Resistant Tuberculosis: An
253 Official ATS/CDC/ERS/IDSA Clinical Practice Guideline. *Am J Respir Crit Care Med.*
254 2025;211(1):15–33.
- 255 2. Mudde SE, Upton AM, Lenaerts A, Bax HI, De Steenwinkel JE. Delamanid or pretomanid?
256 A Solomonic judgement! *J Antimicrob Chemother.* 2022;77(4):880–902.
- 257 3. Stover CK, Warrener P, VanDevanter DR, Sherman DR, Arain TM, Langhorne MH, et al.
258 A small-molecule nitroimidazopyran drug candidate for the treatment of tuberculosis.
259 *Nature.* 2000;405(6789):962–6.
- 260 4. Matsumoto M, Hashizume H, Tomishige T, Kawasaki M, Tsubouchi H, Sasaki H, et al.
261 OPC-67683, a nitro-dihydro-imidazooxazole derivative with promising action against
262 tuberculosis in vitro and in mice. *PLoS Med.* 2006;3(11):e466.
- 263 5. Matsumoto M, Hashizume H, Tsubouchi H, Sasaki H, Itotani M, Kuroda H, et al. Screening
264 for novel antituberculosis agents that are effective against multidrug resistant tuberculosis.
265 *Curr Top Med Chem.* 2007;7(5):499–507.
- 266 6. Hayashi M, Nishiyama A, Kitamoto R, Tateishi Y, Osada-Oka M, Nishiuchi Y, et al.
267 Adduct formation of delamanid with NAD in mycobacteria. *Antimicrob Agents Chemother.*
268 2020;64(5):10–1128.
- 269 7. Manjunatha U, Boshoff HI, Barry CE. The mechanism of action of PA-824: novel insights
270 from transcriptional profiling. *Commun Integr Biol.* 2009;2(3):215–8.
- 271 8. Zeng S, Zhang J, Sun M, Zhang X, Cook GM, Zhang T. Nitric oxide-dependent electron
272 transport chain inhibition by the cytochrome bc 1 inhibitor and pretomanid combination
273 kills *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother.* 2021;65(9):10–1128.
- 274 9. Boeree M, Lange C, Thwaites G, Paton N, de Vruh R, Barros D, et al. UNITE4TB: a new
275 consortium for clinical drug and regimen development for TB. *Int J Tuberc Lung Dis.*
276 2021;25(11):886.
- 277 10. Lin Y, van Der Laan LE, Karlsson MO, Garcia-Prats AJ, Hesselning AC, Svensson EM.
278 Model-Informed Once-Daily Dosing Strategy for Bedaquiline and Delamanid in
279 Children, Adolescents and Adults with Tuberculosis. *Clin Pharmacol Ther.* 2024.
- 280 11. Gumbo T, Lenaerts AJ, Hanna D, Romero K, Nuermberger E. Nonclinical models for
281 antituberculosis drug development: a landscape analysis. *J Infect Dis.*
282 2015;211(suppl_3):S83–95.

- 283 12. Berg A, Clary J, Hanna D, Nuermberger E, Lenaerts A, Ammerman N, et al. Model-based
284 meta-analysis of relapsing mouse model studies from the critical path to tuberculosis drug
285 regimens initiative database. *Antimicrob Agents Chemother.* 2022;66(3):e01793-21.
- 286 13. Walter ND, Ernest JP, Dide-Agossou C, Bauman AA, Ramey ME, Rossmassler K, et al.
287 Lung microenvironments harbor *Mycobacterium tuberculosis* phenotypes with distinct
288 treatment responses. *Antimicrob Agents Chemother.* 2023;67(9):e00284-23.
- 289 14. Reichlen MJ, Born SE, Lyons MA, Rossmassler K, Reid J, Robertson GT, et al.
290 Standardized RS ratio metrics to assess tuberculosis antimicrobial efficacy and potency.
291 *Antimicrob Agents Chemother.* 2023;67(1):e01483-22.
- 292 15. Wynn EA, Dide-Agossou C, Reichlen M, Rossmassler K, Al Mubarak R, Reid JJ, et al.
293 Transcriptional adaptation of *Mycobacterium tuberculosis* that survives prolonged multi-
294 drug treatment in mice. *Mbio.* 2023;14(6):e02363-23.
- 295 16. Wynn EA, Dide-Agossou C, Al Mubarak R, Rossmassler K, Eknitphong V, Bauman AA,
296 et al. Emergence of antibiotic-specific *Mycobacterium tuberculosis* phenotypes during
297 prolonged treatment of mice. *Antimicrob Agents Chemother.* 2025:e01310-24.
- 298 17. Walter ND, Born SE, Robertson GT, Reichlen M, Dide-Agossou C, Eknitphong VA, et al.
299 *Mycobacterium tuberculosis* precursor rRNA as a measure of treatment-shortening activity
300 of drugs and regimens. *Nat Commun.* 2021;12(1):2899.
- 301 18. Dide-Agossou C, Bauman AA, Ramey ME, Rossmassler K, Al Mubarak R, Pauly S, et al.
302 Combination of *Mycobacterium tuberculosis* RS ratio and CFU improves the ability of
303 murine efficacy experiments to distinguish between drug treatments. *Antimicrob Agents*
304 *Chemother.* 2022;66(4):e02310-21.
- 305 19. Lenaerts AJ, Gruppo V, Marietta KS, Johnson CM, Driscoll DK, Tompkins NM, et al.
306 Preclinical testing of the nitroimidazopyran PA-824 for activity against *Mycobacterium*
307 *tuberculosis* in a series of in vitro and in vivo models. *Antimicrob Agents Chemother.*
308 2005;49(6):2294–301.
- 309 20. Reichlen MJ, Born SE, Lyons MA, Rossmassler K, Reid J, Robertson GT, et al.
310 Standardized RS ratio metrics to assess tuberculosis antimicrobial efficacy and potency.
311 *Antimicrob Agents Chemother.* 2023;67(1):e01483-22.
- 312 21. Sarathy JP, Via LE, Weiner D, Blanc L, Boshoff H, Eugenin EA, et al. Extreme drug
313 tolerance of *Mycobacterium tuberculosis* in caseum. *Antimicrob Agents Chemother.*
314 2018;62(2):10–1128.
- 315 22. Goh JJ, Patel A, Ngara B, van Wijk RC, Strydom N, Wang Q, et al. Predicting tuberculosis
316 drug efficacy in preclinical and clinical models from in vitro data. *iScience.* 2025.
- 317 23. Irwin SM, Driver E, Lyon E, Schrupp C, Ryan G, Gonzalez-Juarrero M, et al. Presence of
318 multiple lesion types with vastly different microenvironments in C3HeB/FeJ mice

319 following aerosol infection with *Mycobacterium tuberculosis*. *Dis Model Mech.*
320 2015;8(6):591–602.

321 **Figures 1a-b**



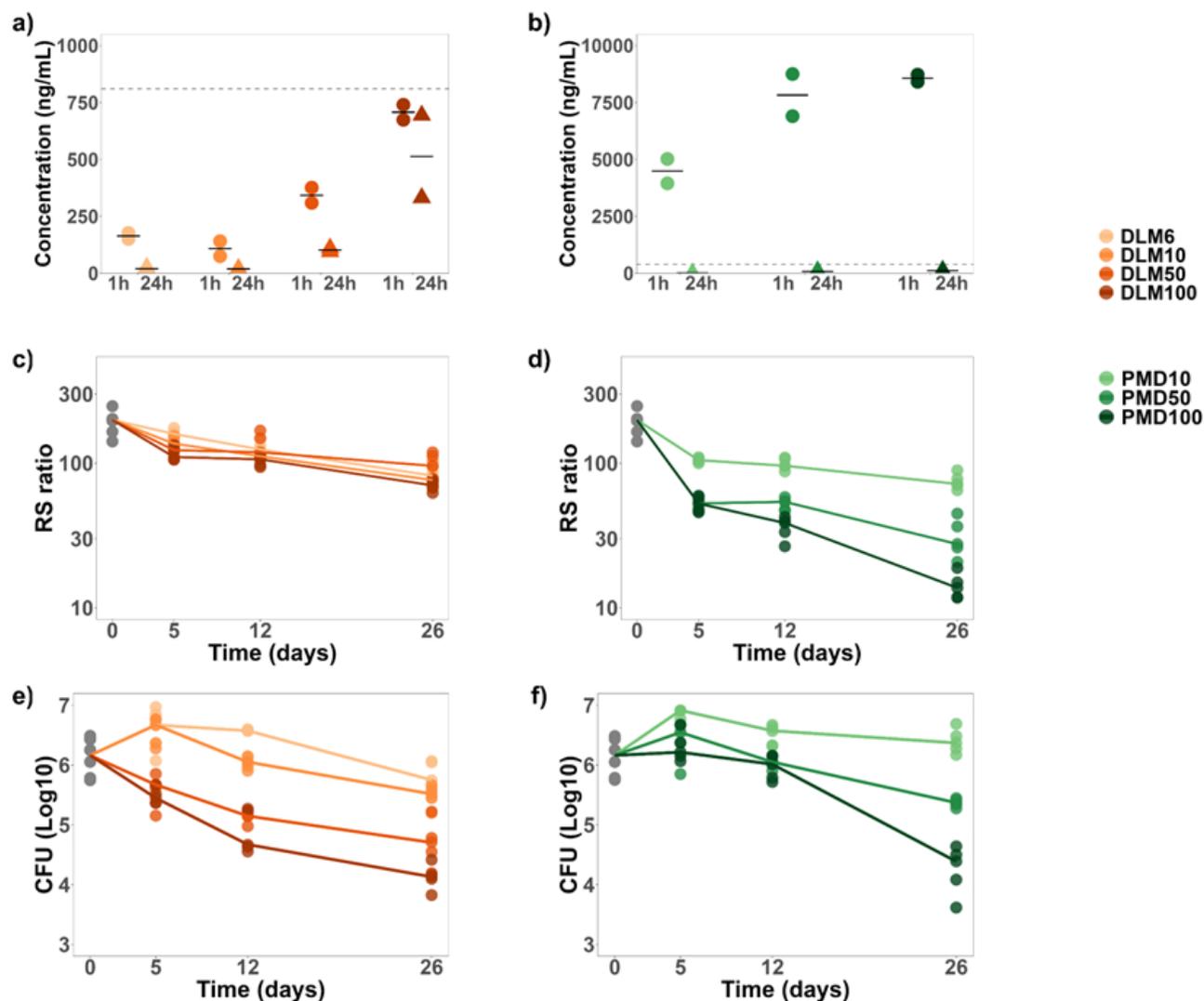
323 **Figure 1:** *In vitro* RS ratio dose-response sigmoid E_{\max} models of delamanid and pretomanid.
324 Solid black dashed lines depict best-fit curves. Green horizontal dashed lines depict the $RS-E_{\max}$.
325 Grey horizontal dashed lines depict mean RS ratio response for no drug at 48 h. Yellow vertical
326 dashed lines depict $RS-EC_{90}$. Circles represent values from individual samples. Data for
327 pretomanid was published previously in ref ¹⁴. Dose-response curves were fitted to observed data
328 using least squares regression and medium convergence criteria (maximum iterations = 1000).

329

330

331

332 **Figures 2a-f**



333

334 **Figure 2.** Pharmacokinetics and pharmacodynamic markers during treatment with varying
335 concentrations of delamanid and pretomanid (a–b) C_{max} and trough concentrations following
336 treatment with varying doses of (a) delamanid and (b) pretomanid after 1 h (circles) and 24 h
337 (triangles). Horizontal bars indicate median values. Dotted gray lines indicate the RS-EC₉₀
338 concentration determined *in vitro*. (c–d) Change in RS ratio during treatment with varying doses
339 of (c) delamanid and (d) pretomanid. (e–f) Change in CFU during treatment with varying doses
340 of (e) delamanid and (f) pretomanid. For c–f, circles represent values from individual mice and
341 lines connect median values.

342

343 **Table 1.** Median RS ratio at various durations and doses of delamanid and pretomanid.
 344 Wilcoxon *P*-values comparing equal dosing are shown.

Day/Dose	Median RS ratio		<i>P</i> -value
	Delamanid	Pretomanid	
Day 5			
6 mg/kg*	159.3	--	---
10 mg/kg	136.4	104.9	0.008
50 mg/kg [¶]	123.0	52.7	0.008
100 mg/kg	110.2	52.5	0.008
Day 12			
6 mg/kg*	124.2	--	---
10 mg/kg	111.6	95.9	0.055
50 mg/kg [¶]	118.9	54.0	0.008
100 mg/kg	106.3	38.6	0.008
Day 26			
6 mg/kg*	81.9	--	---
10 mg/kg	76.4	95.9	0.31
50 mg/kg [¶]	95.7	38.6	0.008
100 mg/kg	70.2	38.6	0.008

345 * Standard human-equivalent doses [plasma AUC] are 6 mg/kg delamanid, and [¶] 50 mg/kg
 346 for pretomanid.

347

348 **Table 2.** Change in CFU relative to pre-treatment control (\log_{10}). Positive and negative values
 349 indicate an increase or decrease in CFU relative to pre-treatment control, respectively. Wilcoxon
 350 *P*-values comparing equal dosing are shown.
 351

Day/Dose	Decrease in CFU (\log_{10})		<i>P</i> -value
	Delamanid	Pretomanid	
Day 5			
6 mg/kg*	+ 0.51	---	
10 mg/kg	+ 0.51	+ 0.75	0.18
50 mg/kg [†]	- 0.49	+ 0.38	0.065
100 mg/kg	- 0.71	+ 0.05	0.002
Day 12			
6 mg/kg*	+ 0.41	---	
10 mg/kg	+ 0.11	+ 0.41	0.03
50 mg/kg [†]	- 1.01	- 0.11	0.002
100 mg/kg	- 1.49	- 0.11	0.009
Day 26			
6 mg/kg*	- 0.41	---	
10 mg/kg	- 0.65	+ 0.21	0.004
50 mg/kg [†]	- 1.46	- 0.79	0.04
100 mg/kg	- 2.03	- 1.78	0.59

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 353 * Standard human-equivalent doses are 6 mg/kg for delamanid, and [†] 50 mg/kg for
 354 pretomanid.
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